

(kDa) 250

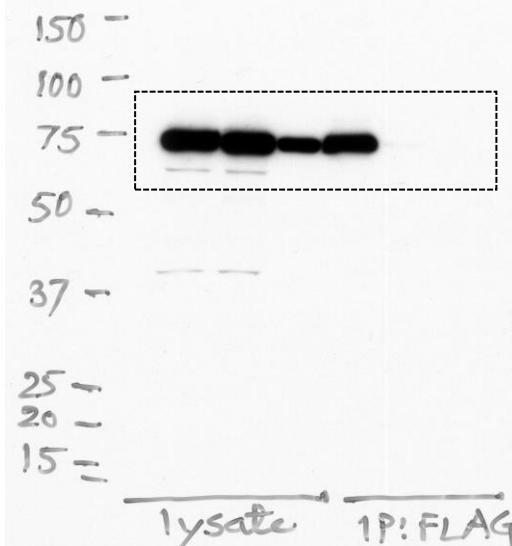


Figure 2B Upper  
Myc-myoclonin1

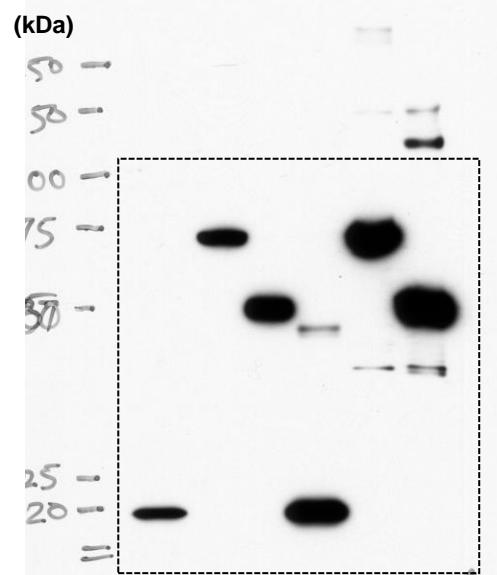


Figure 2B Lower  
FLAG-IP<sub>3</sub>R1/C, /N and Endophilin

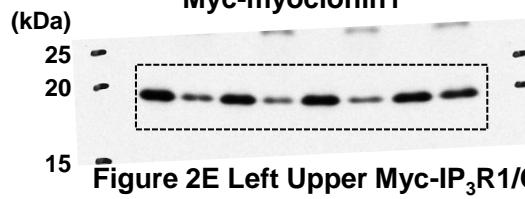


Figure 2E Left Upper Myc-IP<sub>3</sub>R1/C

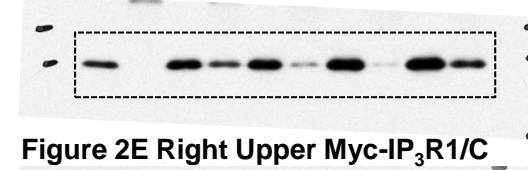


Figure 2E Right Upper Myc-IP<sub>3</sub>R1/C

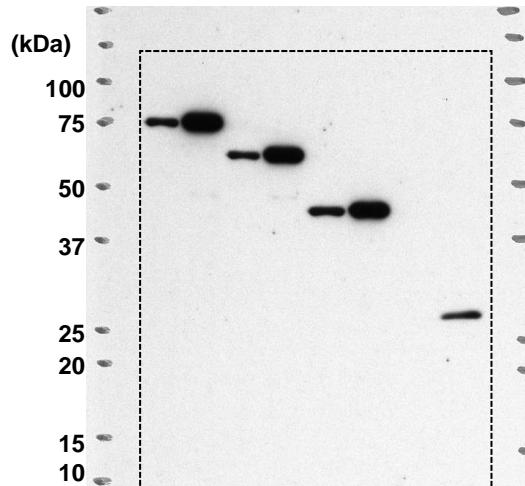


Figure 2E Left Lower  
FLAG-myoclonin1 deletion forms

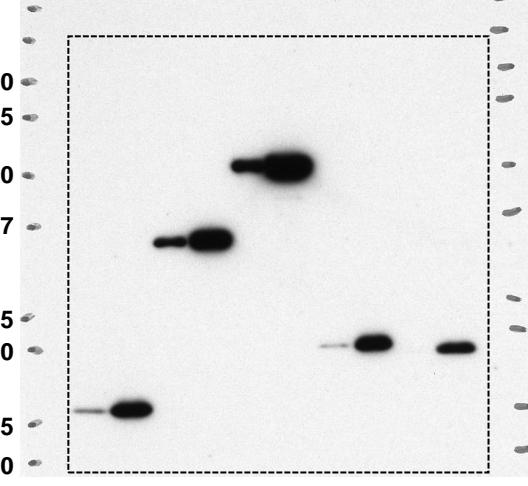


Figure 2E Right Lower  
FLAG-myoclonin1 deletion forms

(kDa)

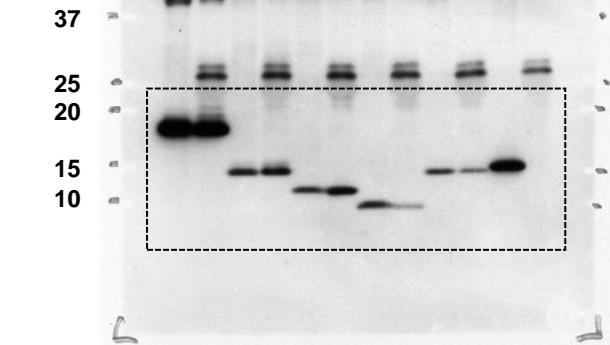


Figure 2C Left Upper  
Myc-IP<sub>3</sub>R1 deletion forms

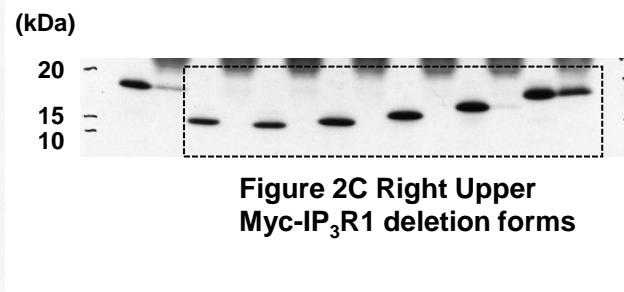


Figure 2C Right Upper  
Myc-IP<sub>3</sub>R1 deletion forms

(kDa)

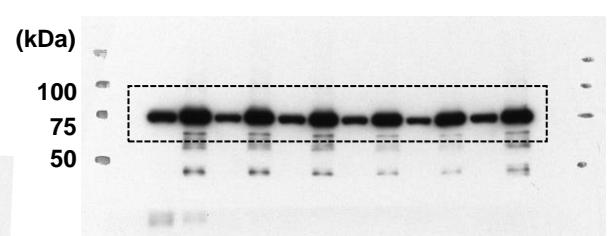


Figure 2C Left Lower  
FLAG-myoclonin1

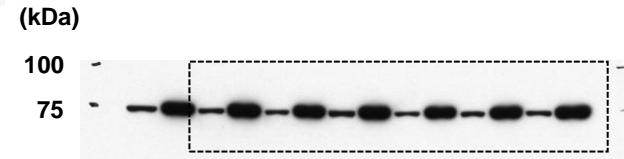
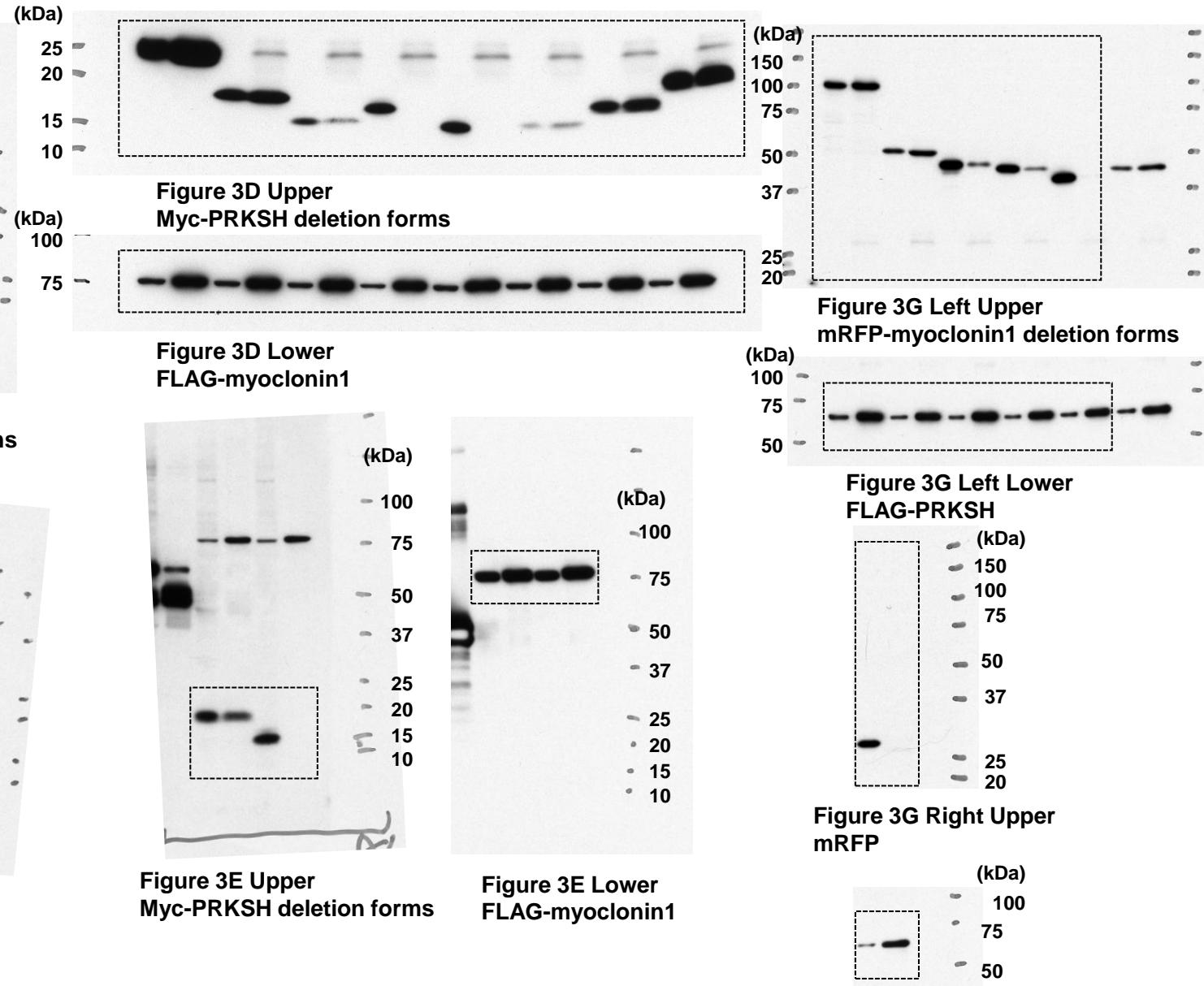
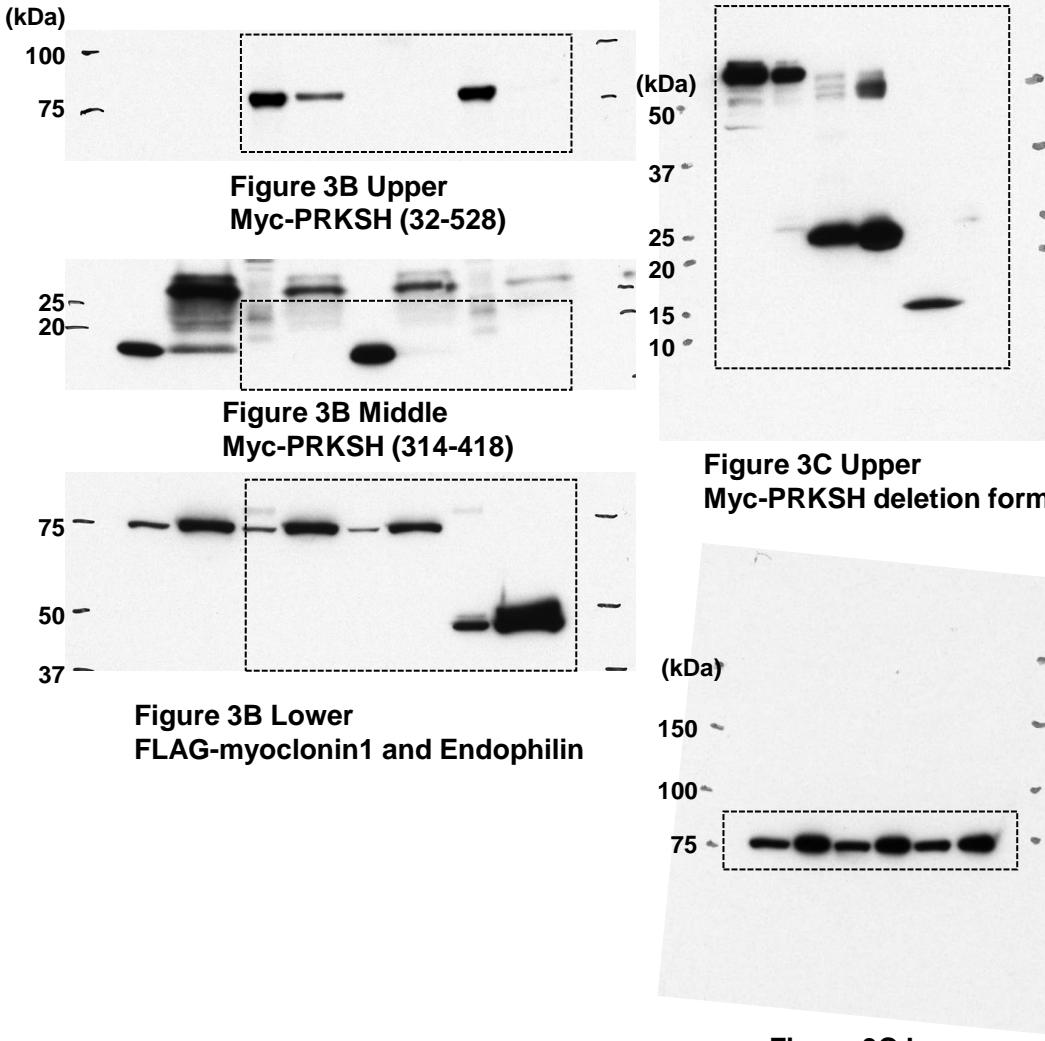


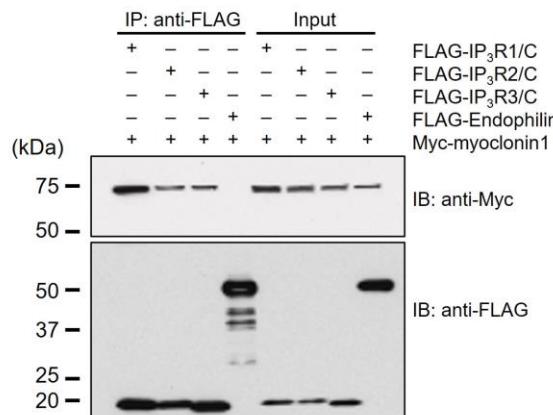
Figure 2C Right Lower  
FLAG-myoclonin1



**Figure S1. Full size western blot images.** Original scanned western blot data that were used to generate Figure 2 and 3. Dashed rectangles in the images indicate the locations of the cropped images.

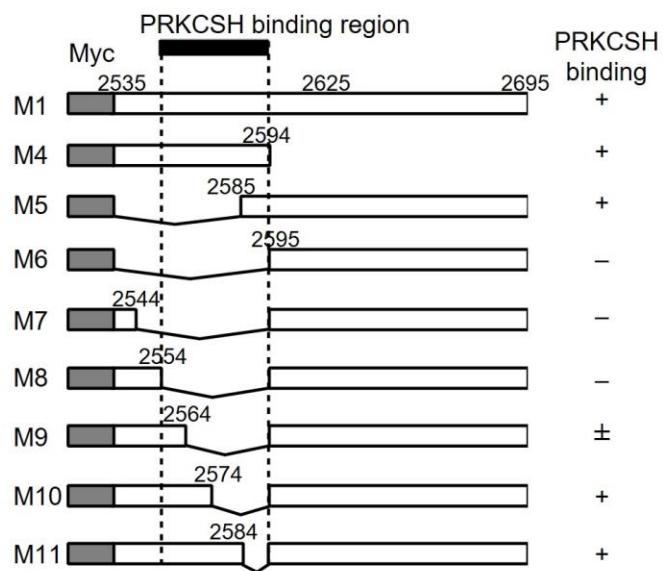
**A**

myoclonin1 binding region						
<i>Homo sapiens</i>	IP <sub>3</sub> R1	2565	KFDNKTVT <del>FEEHIKEEHN</del> MWHYLCFIVLVVKVD <del>STEYTGPESYVAEMIKERNLDWFPRMRA</del>		2625	
<i>Mus musculus</i>	IP <sub>3</sub> R1	2564	KFDNKTVT <del>FEEHIKEEHN</del> MWHYLCFIVLVVKVD <del>STEYTGPESYVAEMIRERNLDWFPLRMRA</del>		2624	
<i>Rattus norvegicus</i>	IP <sub>3</sub> R1	2579	KFDNKTVT <del>FEEHIKEEHN</del> MWHYLCFIVLVVKVD <del>STEYTGPESYVAEMIRERNLDWFPLRMRA</del>		2639	
<i>Gallus gallus</i>	IP <sub>3</sub> R1	2581	KFDNKTVT <del>FEEHIKEEHN</del> MWHYLCFIVLVVKVD <del>STEYTGPESYVAEMIKERNLDWFPRMRA</del>		2641	
<i>Bos taurus</i>	IP <sub>3</sub> R1	2579	KFDNKTVT <del>FEEHIKEEHN</del> MWHYLCFIVLVVKD <del>STEYTGPESYVAEMIKERNLDWFPRMRA</del>		2639	
<i>Homo sapiens</i>	IP <sub>3</sub> R2	2571	KFDNKTVS <del>FEEHIK</del> SEHN <del>MWHY</del> L <del>F</del> IVLVVKD <del>PTEYTGPESYVAOMIVEKNLDWFPRMRA</del>		2631	
<i>Mus musculus</i>	IP <sub>3</sub> R2	2571	KFDNKTVS <del>FEEHIK</del> SEHN <del>MWHY</del> L <del>F</del> IVLVVKD <del>PTEYTGPESYVAOMITEKNLDWFPRMRA</del>		2631	
<i>Rattus norvegicus</i>	IP <sub>3</sub> R2	2571	KFDNKTVS <del>FEEHIK</del> SEHN <del>MWHY</del> L <del>F</del> IVLVVKD <del>PTEYTGPESYVAOMITEKNLDWFPRMRA</del>		2631	
<i>Gallus gallus</i>	IP <sub>3</sub> R2	2570	KFDNKTVS <del>FEEHIK</del> SEHN <del>MWHY</del> L <del>F</del> IVLVVKD <del>PTEYTGPESYVAOMIVEKNLDWFPRMRA</del>		2630	
<i>Bos taurus</i>	IP <sub>3</sub> R2	2571	KFDNKTVS <del>FEEHIK</del> SEHN <del>MWHY</del> L <del>F</del> IVLVVKD <del>PTEYTGPESYVAOMIVEKNLDWFPRMRA</del>		2631	
<i>Homo sapiens</i>	IP <sub>3</sub> R3	2547	KFDNKTVS <del>FEEHIK</del> DEHN <del>MWN</del> Y <del>L</del> <del>F</del> IVLVVRVKNKT <del>DYTGPESYVAOMIKKNKNLDWFPRMRA</del>		2607	
<i>Mus musculus</i>	IP <sub>3</sub> R3	2546	KFDNKTVS <del>FEEHIK</del> DEHN <del>MWN</del> Y <del>L</del> <del>F</del> IVLVVRVKNKT <del>DYTGPESYVAOMIKKNKNLDWFPRMRA</del>		2606	
<i>Rattus norvegicus</i>	IP <sub>3</sub> R3	2546	KFDNKTVS <del>FEEHIK</del> DEHN <del>MWN</del> Y <del>L</del> <del>F</del> IVLVVRVKNKT <del>DYTGPESYVAOMIKKNKNLDWFPRMRA</del>		2606	
<i>Gallus gallus</i>	IP <sub>3</sub> R3	2540	KFDNKTVS <del>FEEHIK</del> DEHN <del>MWN</del> Y <del>L</del> <del>F</del> IVLVVRVKNKT <del>DYTGPESYVAOMIKKNKNLDWFPRMRA</del>		2600	
<i>Bos taurus</i>	IP <sub>3</sub> R3	2540	KFDNKTVS <del>FEEHIK</del> DEHN <del>MWN</del> Y <del>L</del> <del>F</del> IVLVVRVKNKT <del>DYTGPESYVAOMIKKNKNLDWFPRMRA</del>		2600	

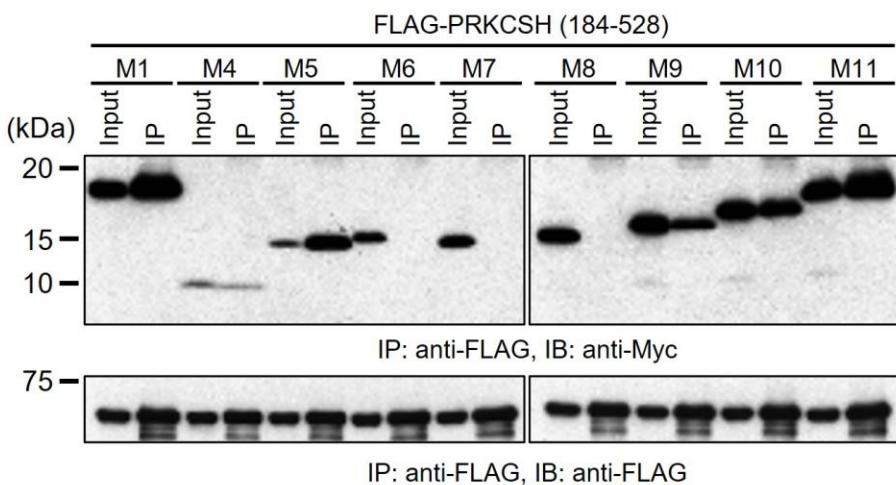
**B**

**Figure S2. Myoclonin1 interacts with C-termini of all three IP<sub>3</sub>R subtypes. (A)**  
 Binding regions of all three IP<sub>3</sub>R subtypes (IP<sub>3</sub>R1, 2, 3) to myoclonin1 are highly conserved. **(B)** IP<sub>3</sub>R2 and IP<sub>3</sub>R3 C-termini also interacted with myoclonin1 but not with Endophilin (negative control). IP, immunoprecipitation; IB, immunoblot; Input, 5% of cell lysate; /C, C-terminal.

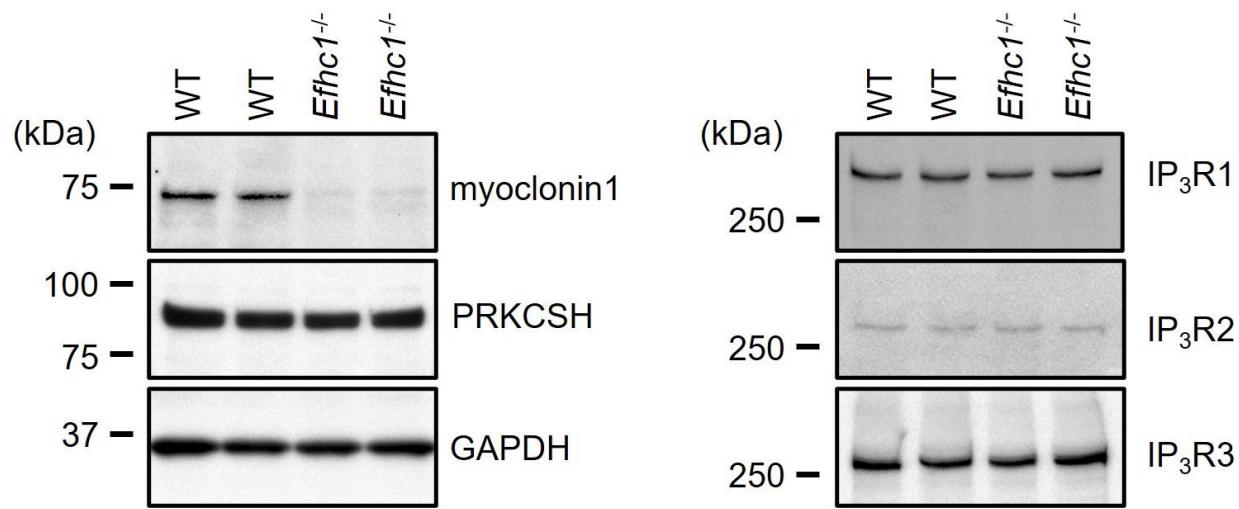
**A**  
**IP<sub>3</sub>R1/C**



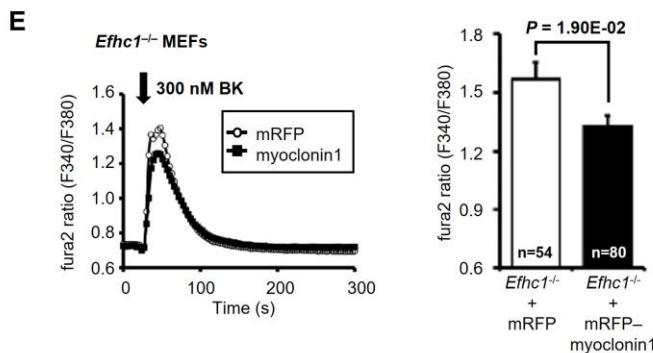
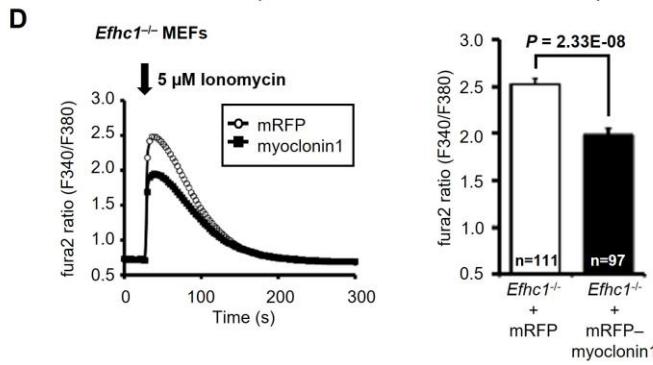
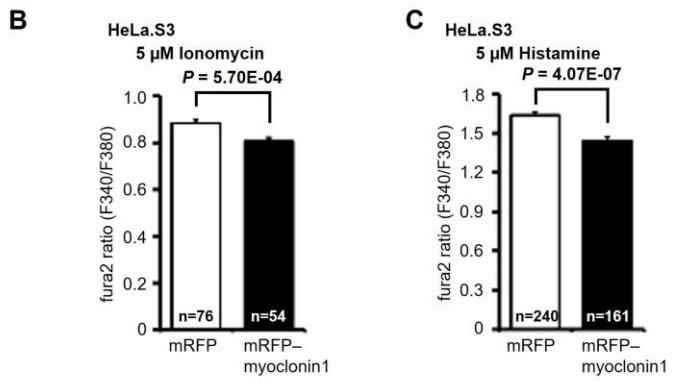
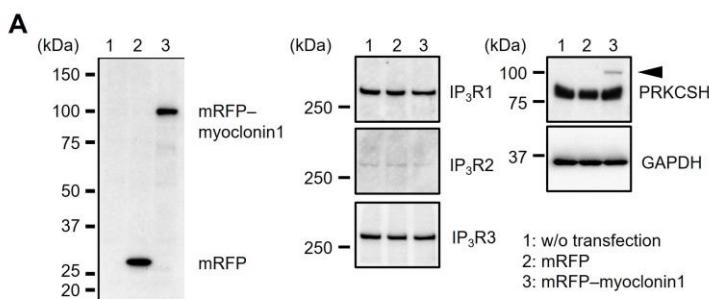
**B**



**Figure S3. PRKCSH interacts with IP<sub>3</sub>R1 C-terminus at its interaction site for myoclonin1.** (A) Schematic diagram of C-terminal IP<sub>3</sub>R1 deletion constructs and their binding activities to PRKCSH. A bold black bar (top) indicates a binding site to PRKCSH. Degrees of interaction are indicated by +, ± or -. (B) Western blots of co-IP showing that PRKCSH bound to a.a. 2555–2594 of C-terminal IP<sub>3</sub>R1. IP, immunoprecipitation; IB, immunoblot; Input, 5% of cell lysate; /C, C-terminal.



**Figure S4. Expressions of IP<sub>3</sub>Rs and PRKCSH do not change in *Efhc1*<sup>-/-</sup> MEFs.**  
 Western blot analyses revealed that myoclonin1 expression was abrogated in *Efhc1*<sup>-/-</sup> MEFs, whereas expressions of endogenous IP<sub>3</sub>Rs and PRKCSH do not change (n=2 independent embryos per genotypes). An antibody to GAPDH was used as a control.



**Figure S5. Myoclonin1 reduces  $[Ca^{2+}]_{ER}$  and IICR.** (A) Western blot analyses [n=1 without (w/o) transfection, 1 mRFP, 1 mRFP–myoclonin1] revealed that exogenous of mRFP or mRFP–myoclonin1 proteins were detected at expected size (~30 kDa and ~100 kDa, respectively). Expression levels of endogenous IP<sub>3</sub>R<sub>s</sub> and PRKCSH were not affected by over-expression of myoclonin1 in the cells. An mRFP–myoclonin1 band at ~100 kDa remained weakly in blot for PRKCSH (arrowhead) even after stripping of blot. An antibody to GAPDH was used as a control. (B, C) Ionomycin releasable  $[Ca^{2+}]_{ER}$  (B; n=76 mRFP, 54 mRFP–myoclonin1 expressing cells) and histamine evoked IICR (C; n=240 mRFP, 161 mRFP–myoclonin1 expressing cells) were significantly lower in mRFP–myoclonin1 expressing HeLa.S3 cells than mRFP expressing one. Ionomycin releasable  $[Ca^{2+}]_{ER}$  (D, n=111 mRFP, 97 mRFP–myoclonin1) and bradykinin (BK) evoked IICR (E, n=54 mRFP, 80 mRFP–myoclonin1) were significantly lower in mRFP–myoclonin1 expressing *Efhc1<sup>-/-</sup>* MEFs than mRFP expressing one. Arrows indicate the time point of addition of ionomycin or BK. n, total number of cells measured.