



Taste genomics and type 2 diabetes mellitus: a systematic qualitative meta-synthesis

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Abstract

Background: Emerging evidence suggests that genetic variations in taste receptor genes may influence dietary behaviors, energy homeostasis, and metabolic risk, contributing to type 2 diabetes mellitus (T2DM) pathogenesis. The objective of this study is to evaluate the association between single nucleotide polymorphisms (SNPs) in taste receptor genes and T2DM.

Methods: This systematic review followed the Preferred Reporting Items for Systematic reviews and Meta-Analyses (PRISMA) 2020 guidelines and was registered with the International Prospective Register of Systematic Reviews (PROSPERO; CRD42022351880). A comprehensive literature search was conducted across PubMed, ScienceDirect, Cochrane Library, and Google Scholar through June 2025. Original studies examining SNPs in taste receptor genes among individuals with T2DM were included. Quality assessment was performed independently by using the Newcastle-Ottawa scale.

Results: Sixteen studies involving diverse populations met the inclusion criteria. Significant associations with T2DM were observed for SNPs in type 2 taste receptor gene family R member 3 (*TAS2R3*; rs11763979), *TAS2R4* (rs2233998), *TAS2R7*, *TAS2R9*, *TAS2R38*, *TAS2R50*, cluster determinant 36 (*CD36*; rs1761667, rs3211956, rs7755), carbonic anhydrase VI gene (*CA6*; rs2274327), transient receptor potential vanilloid-1 (*TRPV1*; rs161364, rs8065080), transient receptor potential cation channel subfamily M gene member 5 (*TRPM5*; rs4929982), and *TRPM8* (rs12472151). These polymorphisms may alter taste perception and gut hormone responses [e.g., glucagon-like peptide 1 (GLP-1)], affecting dietary intake, satiety, insulin secretion, and glucose regulation.

Discussion: The findings suggest that genetic variations in taste receptor genes may contribute to T2DM through behavioral and metabolic mechanisms. Incorporating gustatory phenotyping with genotypic profiling could enable personalized dietary strategies and inform novel therapeutic approaches targeting taste-mediated gut signaling. Further large-scale, multi-ethnic, and mechanistic studies are warranted to confirm these associations and elucidate their clinical implications.



Keywords

Type 2 diabetes mellitus, taste receptors, taste gene polymorphism, genetic risk, dietary behavior

Introduction

Over the past few decades, the global prevalence of type 2 diabetes mellitus (T2DM) has increased markedly, with projections estimating up to 7,862 cases per 100,000 individuals [1]. This upward trend may be attributed to a complex interplay of genetic and environmental risk factors, including obesity, insulin resistance, metabolic dysfunction, dietary habits, and epigenetic modifications [1]. In addition to these well-established risk factors, emerging evidence suggests that behavioral psychology and impaired satiety signaling also play a significant role in the pathogenesis of T2DM [2–4].

Recent studies have identified that single nucleotide polymorphisms (SNPs) in taste genes are significantly associated with elevated risk of metabolic syndrome, diabetes mellitus, obesity, carcinogenesis, Alzheimer's disease, Parkinson's disease, thyroid dysfunction, and substance use disorders [3–5]. Taste perception and signal transduction across the six taste modalities—sweet, salt, sour, bitter, umami, and fat taste—are mediated by various taste receptor genes such as type 1 taste receptor gene family R (*TAS1R*), *TAS2R*, sodium channel epithelial 1 (*SCNN1*), cluster determinant 36 (*CD36*), transient receptor potential cation channel subfamily M gene (*TRPM*), guanine nucleotide binding protein subunit alpha transducing 3 (*GNAT3*), carbonic anhydrase VI gene (*CA6*), *IZUMO* sperm-egg fusion 1 gene (*IZUMO1*), metabotropic glutamate receptor 1 gene (*GRM1*), and polycystic kidney disease (PKD)-like genes (e.g., *PKD1L3*, *PKD2L1*, and *PKD2L3*) [4–7]. Each taste modality is regulated by specific taste receptor genes. For instance: sweet taste is primarily mediated by *TAS1Rs*, *TRPMs*, and *GNAT3*; bitter taste by *TAS2Rs*, *TRPMs*, and *CA6*; salt taste by transient receptor potential vanilloid-1 (*TRPV1*; nerve endings) and *SCNN1s* (subunits alpha, beta, gamma, and delta); sour taste by *TAS1Rs* and PKD-like genes; umami taste by *TAS1Rs*, *TRPMs*, *GNAT3*, and *GRM1*; and fat taste by *CD36* and *IZUMO1* [5–7]. Taste perception begins with the interaction of food containing particular taste stimuli with oral and extra-oral taste receptors, triggering intracellular calcium release into the cytoplasm, depolarization of afferent nerve fibers and signal transduction via cranial nerves (facial nerve, glossopharyngeal nerve, sensory vagal afferents, trigeminal nerve and the trigeminal ganglion) to the central processing centers (nucleus of solitary tract, ventral posteromedial thalamic nucleus, the operculum, insular and the somatosensory cortex) [8]. These brain regions also influence gut hormone secretion [e.g., ghrelin, glucagon-like peptide 1 (GLP-1), glucose-dependent insulin-releasing peptide], thereby regulating satiety and energy homeostasis [8]. Genetic variations in taste receptors may impair this signaling cascade, leading to altered taste perception, eating behavior, impaired energy homeostasis, and increased susceptibility to T2DM [5]. Notably, *TAS1R* expression in the gut is upregulated in response to hyperglycemia in individuals with T2DM [9]. This suggests that modulating taste receptor pathways may have therapeutic potential, particularly through agents that mimic GLP-1 receptor agonists [2, 8].

Several studies have reported significant taste impairments in individuals with diabetes mellitus [10, 11]. Chamoun et al. [12] demonstrated associations between psychophysical measures of taste and 94 SNPs across 11 taste receptor genes, particularly those related to sweet, salty, umami, and fat taste perception. A recent review further highlighted the critical role of taste receptor function in the pathophysiology of T2DM and energy homeostasis [13].

Despite the growing body of evidence, no qualitative meta-synthesis has systematically evaluated the association between SNPs in taste receptor genes and T2DM. This study aims to evaluate the evidence on the association of taste gene polymorphisms and T2DM.

Materials and methods

This qualitative meta-synthesis was registered with the International Prospective Register of Systematic Reviews (PROSPERO; Registration No. CRD42022351880) [14] and conducted in accordance with the Preferred Reporting Items for Systematic reviews and Meta-Analyses (PRISMA) 2020 guidelines.

Search strategy and selection criteria

A comprehensive literature search was performed across PubMed, ScienceDirect, Cochrane Library databases, and Google Scholar using the keywords “Diabetes mellitus” AND (“taste receptor gene” OR “taste gene” OR “taste gene polymorphisms” OR “taste gene mutations”). Manual screening of reference lists and citation tracking of relevant articles was also conducted to ensure thorough coverage of the literature up to June 2025. Inclusion criteria: Original research articles investigating SNPs in taste receptor genes among patients with T2DM were included. Exclusion criteria: Articles not relevant to the study, including review articles, case reports, editorials, consensus statements, clinical guidelines, conference abstracts, and book chapters, were excluded. Titles and abstracts were screened to identify potentially relevant records. Full-text screening of the identified records was assessed for inclusion by one reviewer and independently reassessed by the reviewer. The PICO framework for this study are as follows: (1) population: individuals diagnosed with T2DM, (2) intervention/exposure: presence of SNPs or variant genotypes in taste receptor genes, (3) comparison: comparison between individuals with variant versus wild-type genotypes, and (4) outcome: association between taste receptor polymorphisms and T2DM.

Data extraction and quality assessment

Relevant study characteristics were systematically extracted, including first author, year of publication, study design, country of origin, sample size, age of participants, diagnostic criteria for T2DM, taste receptor genotyping, and other characteristic features, from studies that met the inclusion criteria. Methodological quality assessment of the included studies was carried out with the Newcastle-Ottawa scale (Table S1) [3, 4]. Data extraction was performed by one reviewer and cross-verified by another reviewer.

Results

The PRISMA flowchart (Figure 1) illustrates the selection process. A total of 5,712 records were identified through database searches and manual screening. After title and abstract screening, followed by full-text review, sixteen studies met the eligibility criteria and were included in the qualitative meta-synthesis. Four studies were excluded due to the inclusion of populations other than T2DM (one study was conducted in individuals with prediabetes [15], and three studies were conducted in patients with gestational diabetes mellitus (GDM) [16–18]).

The characteristics of the included studies were summarized in Table 1 and Table S1. Diagnostic criteria for T2DM were based on the definitions used within each included study. Methodological quality assessment of the included studies by the Newcastle-Ottawa scale indicated that all sixteen studies were of good quality (Table S1).

The results revealed that SNPs in taste genes including *TAS2R3* gene (SNP rs11763979), *TAS2R4* gene (SNP rs2233998), *TAS2R7* gene (SNPs rs2588350 and rs619381), *TAS2R9* gene (SNP rs3741845), *TAS2R38*, *TAS2R50* gene (SNP rs6488334), *TRPV1* gene (SNPs rs161364 and rs8065080), *CD36* gene (SNPs rs1761667, rs3211956, rs7755, rs1049673, and rs1527479), and *CA6* gene (SNP rs2274327) were significantly associated with patients with T2DM. Additionally, *TRPM5* gene (SNP rs4929982) and *TRPM8* gene (SNP rs12472151) polymorphisms were significantly associated with metabolic syndrome, including T2DM.

Discussion

The findings suggest that SNPs in several taste-related genes, including *TAS2R3*, *TAS2R4*, *TAS2R7*, *TAS2R9*, *TAS2R38*, *TAS2R50*, *TRPV1*, *CD36*, *CA6*, *TRPM5*, and *TRPM8*, may contribute to the etiology of T2DM and its associated metabolic complications. Although the precise mechanisms remain to be elucidated, current evidence supports the hypothesis that genetic variations in taste genes influence taste stimuli perception, individual food preferences, nutrient intake, and eating behavior, thereby increasing the risk of T2DM [22].

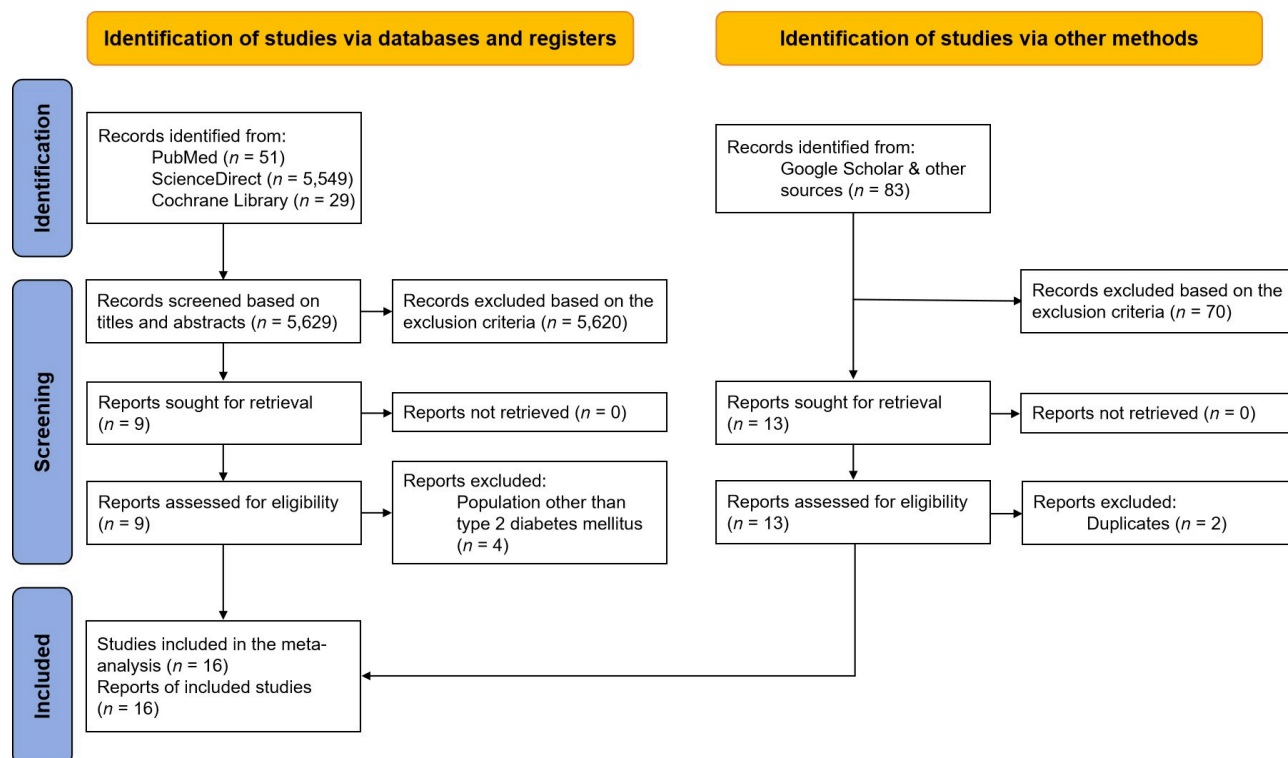


Figure 1. Flow chart summarizing the selection process. Adapted from [19], © 2021, The Author(s) (CC BY 4.0)

Table 1. General characteristics of the studies included in the qualitative meta-synthesis

Study	Study design (country)	Population	Genotyping	Inference
Leprêtre et al. [20], 2004	Cohort (France)	454	<i>CD36</i> gene (entire sequence)	No significant association was found between <i>CD36</i> gene and T2DM ($P > 0.1$)
Corpeleijn et al. [21], 2006	Cohort (Netherlands)	151	<i>CD36</i> gene SNP rs1527479 and 478 C/T substitution	<i>CD36</i> gene SNP rs1527479 TT haplotype was significantly associated with T2DM ($P = 0.035$), fasting glucose concentration ($P < 0.05$) and insulin resistance ($P < 0.05$)
Dotson et al. [22], 2008	Case-control (USA)	503	70 SNPs in <i>TAS1Rs</i> and <i>TAS2Rs</i> gene subtypes	<i>TAS2R3</i> gene SNP rs11763979 ($P = 0.03$), <i>TAS2R7</i> gene SNPs rs2588350 ($P = 0.0007$) and rs619381 ($P = 0.009$), <i>TAS2R9</i> gene SNP rs3741845 ($P = 0.005$), and <i>TAS2R50</i> gene SNP rs6488334 ($P = 0.04$) were significantly associated with patients with T2DM
Banerjee et al. [23], 2010	Case-control (India)	400	Two SNPs (rs1527483 and rs1761667) in the <i>CD36</i> gene	<i>CD36</i> SNP (G>A) rs1761667 GA was significantly associated with patients with T2DM ($P < 0.001$)
Wang et al. [24], 2012	Case-control (China)	113	<i>CD36</i> gene SNPs (rs1527483 and rs1049673)	<i>CD36</i> SNP rs1049673 CG & GG haplotypes were significantly associated with impaired glucose tolerance ($P = 0.023$) and T2DM ($P = 0.011$) in patients with essential hypertension, respectively
Gautam et al. [25], 2015	Case-control (India)	100	Six <i>CD36</i> SNPs (rs1984112, rs1761667, rs1527479, rs3211938, rs1527483, and rs3212018)	<i>CD36</i> SNPs rs1761667 (G>A) and rs3211938 (T>G) showed significant association with T2DM (GAATTC1, $P < 0.001$)
Tabur et al. [26], 2015	Case-control (Turkey)	308	25 <i>TRPM1–8</i> gene SNPs rs28441327, rs11070811, rs2241493, rs111649153, rs1618355, rs1328142, rs3760663, rs34364959, rs4929982, rs886277, rs34551253, rs3986599, rs3750425, rs62569677, rs55924090, rs1016062, rs2362294, rs2362295, rs10490018, rs2052029, rs6431648, rs10803666, rs12472151, rs2215173, and rs6740118	<i>TRPM5</i> gene SNP rs4929982 A allele ($P = 0.0019$) and <i>TRPM8</i> gene SNP rs12472151 C allele ($P < 0.001$) polymorphisms might be related to the individual susceptibility to metabolic syndrome (including T2DM)

Table 1. General characteristics of the studies included in the qualitative meta-synthesis (continued)

Study	Study design (country)	Population	Genotyping	Inference
Park et al. [27], 2016	Cohort (South Korea)	8,842	7 SNPs in <i>TRPV1</i> gene such as SNPs rs161364, rs8065080, rs150908, rs222745, rs7217945, rs222741, and rs2737141	<i>TRPV1</i> gene SNPs rs161364 C allele ($P = 0.0487$) and rs8065080 C allele ($P = 0.0378$) were significantly associated with the prevalence of T2DM in dominant genetic models
Zhang et al. [28], 2018	Case-control (China)	546	Four <i>CD36</i> SNPs rs1194197, rs2151916, rs3211956, and rs7755	Overweight/obesity individuals carrying SNP variant alleles of rs3211956 (GG+GT, $P = 0.024$) and rs7755 (AA+AG, $P = 0.007$) were associated with increased risk of T2DM compared to normal weight individuals carrying wild-type homozygous alleles
Fujii et al. [29], 2019	Cross-sectional (Japan)	495	Two <i>CD36</i> gene SNPs (rs1761667 and rs1527483)	<i>CD36</i> gene SNP rs1761667 AA haplotype was associated with higher intake of total fat ($P = 0.01$) and monounsaturated fatty acids ($P = 0.05$) when compared to GG and GA haplotypes. In addition, the frequency of <i>CD36</i> gene SNP rs1761667 GG haplotype was higher in T2DM
Mrag et al. [30], 2020	Cohort (Tunisia)	300	CA6 gene SNP rs2274327	The CA6 gene SNP rs2274327 T allele in its dominant model (TT+CT vs. CC, 67.7% vs. 32.3%) was increasingly associated with T2DM. Similarly, taste impairment in T2DM was significantly associated with CA6 gene SNP rs2274327 T allele in its dominant model (OR = 1.97 [95% CI = 1.21 to 3.23], $P = 0.006$)
Hatmal et al. [31], 2021	Case-control (Jordan)	350	<i>CD36</i> gene rs1761667 (G>A) and rs1527483 (C>T) were genotyped	No significant association was observed between <i>CD36</i> polymorphisms and patients with T2DM or dyslipidemia ($P > 0.1$)
Touré et al. [32], 2022	Cross-sectional (Senegal)	100	2 tag SNPs in <i>CD36</i> (rs3211867 and rs1761667)	No significant difference was observed between controls and T2DM subjects ($P = 0.9$)
Franzago et al. [33], 2023	Cohort (Italy)	23	<i>CD36</i> gene SNPs rs1984112 (A>G) and rs1761667 (G>A), <i>BMAL1</i> gene SNP rs7950226 (G>A), and <i>CLOCK</i> gene SNPs rs1801260 (A>G), rs4864548 (A>G), and rs3736544 (G>A)	<i>CD36</i> gene SNP rs1761667 (G>A) A allele in its dominant form (AA+GG genotype) was significantly associated with patients with T2DM ($P = 0.001$)
Lee and Shin [34], 2023	Cohort (Korea)	4,552	<i>TAS2R4</i> SNP rs2233998	<i>TAS2R4</i> SNP rs2233998 TT haplotype was significantly associated with the incidence of T2DM in women (HR [95% CI] = 1.48 [1.13–1.93], $P = 0.0182$)
Husami et al. [35], 2025	Case-control (India)	680	2,658 gene variants	<i>TAS2R38</i> genetic variants were associated with an increased risk of T2DM ($P < 0.05$)

case: patients with T2DM as defined in the respective studies; control: normal healthy individuals as defined in the respective studies; *CD36*: cluster determinant 36; T2DM: type 2 diabetes mellitus; SNP: single nucleotide polymorphism; *TAS1R*: type 1 taste receptor gene family R; *TRPM1*: transient receptor potential cation channel subfamily M gene member 1; *TRPV1*: transient receptor potential vanilloid-1; CA6: carbonic anhydrase VI gene; OR: odds ratio; CI: confidence intervals; *BMAL1*: brain and muscle aryl hydrocarbon receptor nuclear translocator-like protein-1; *CLOCK*: circadian locomotor output cycles kaput; HR: hazard ratio

For instance, *TAS2R9* is expressed in gut entero-endocrine L-cells and mediates GLP-1 secretion in response to sweet taste stimuli [22]. Canivenc-Lavier et al. [13] recently described local glucose-dependent GLP-1 secretion by taste bud cells and proposed taste receptors as potential targets for T2DM treatment [3]. Similarly, reduced *CD36* expression due to genetic polymorphisms may influence fat taste sensitivity and intake behaviors, possibly exerting a protective effect due to reduced fat taste sensitivity [4, 33]. Variations in the *TRPM5* gene are associated with reduced GLP-1 levels, impaired insulin release, and altered glucose tolerance, highlighting the metabolic relevance of taste receptor polymorphisms [3, 22].

This is in line with previous observations that the taste-dependent manner of GLP-1 secretion, glucose-stimulated insulin secretion, and insulin sensitivity in patients with T2DM might be associated with the polymorphisms in taste receptor genes [3, 22, 26, 33]. These genetic variations may affect hunger signaling, gut motility, and taste-driven satiety hormone release [3, 4, 13]. For example, individuals carrying the *TAS2R38* proline-alanine-valine (PAV) allele [a 6-n-propylthiouracil (PROP) taster genotype] exhibit heightened sensitivity to bitter compounds, which may impact food choices and contribute to dietary avoidance of bitter but nutritionally beneficial foods [4, 36–38]. Such phenotypic taste differences can now be identified using validated clinical gustatory tests (e.g., taste strips, solutions), which could help screen individuals at risk of metabolic diseases, including obesity and diabetes [3, 4, 36]. Moreover, pharmacologic agents that modulate the taste receptor expression based on an individual's genotype may offer a novel therapeutic avenue [37]. Nevertheless, further large-scale molecular and clinical studies are warranted to validate these associations and uncover the underlying mechanisms driving gene-diet interactions in T2DM.

Strengths and limitations

Overall, this study provides the first systematic synthesis of the association between taste receptor gene polymorphisms and T2DM. However, several limitations exist: (1) high methodological heterogeneity among included studies; (2) potential selection and publication biases; (3) lack of randomized controlled trials; and (4) insufficient data for meta-analysis. Allele frequency variations across ethnicities further limit generalizability. Due to insufficient data to conduct a statistical analysis for all the genes included in the present study, the meta-analysis was not feasible. To address the potential heterogeneity in genotyping methods, variability in study designs, population characteristics, and definitions of T2DM in the included studies, Table 1 and Table S1 summarize the comparison of the characteristics of the included studies, including study design, sample size, country or location of study, and main findings. The risk of bias was assessed using the Newcastle-Ottawa scale, and narrative synthesis and reporting followed the PRISMA 2020 guidelines. Further, citation searches and gray literature searches were performed and a predetermined inclusion and exclusion criteria based on the PROSPERO protocol was used to select the studies for inclusion.

Future perspectives

Further molecular and clinical studies, particularly randomized controlled trials and large, multi-ethnic cohort studies, are warranted to validate the role of taste receptor gene polymorphisms in the pathophysiology of T2DM. Gustatory testing to assess taste phenotypes, when paired with genotypic profiling, may offer a cost-effective approach to identify at-risk individuals and guide personalized dietary interventions. Exploration of taste modulators and GLP-1 analogs targeting specific taste receptors may open new therapeutic avenues in obesity and T2DM management. Moreover, SNPs in taste genes could serve as genetic markers for early detection and risk stratification.

Conclusion

In summary, this review provides evidence that SNPs in taste receptor genes are associated with T2DM. Altered taste perception and signal transduction may influence eating behavior, energy homeostasis, and glucose metabolism. Identifying taste phenotypes and targeting taste receptor gene expression may represent promising strategies for the prevention and treatment of T2DM. However, larger, well-designed studies are needed to confirm these associations and facilitate their clinical translation.

Abbreviations

CA6: carbonic anhydrase VI gene

CD36: cluster determinant 36

GLP-1: glucagon-like peptide 1

GNAT3: guanine nucleotide binding protein subunit alpha transducing 3

GRM1: metabotropic glutamate receptor 1 gene

IZUMO1: *IZUMO* sperm-egg fusion 1 gene

PKD: polycystic kidney disease

PRISMA: Preferred Reporting Items for Systematic reviews and Meta-Analyses

PROSPERO: International Prospective Register of Systematic Reviews

SCNN1: sodium channel epithelial 1

SNPs: single nucleotide polymorphisms

T2DM: type 2 diabetes mellitus

TAS1R: type 1 taste receptor gene family R

TRPM: transient receptor potential cation channel subfamily M gene

TRPV1: transient receptor potential vanilloid-1

Supplementary materials

The supplementary materials for this article are available at: https://www.explorationpub.com/uploads/Article/file/101439_sup_1.pdf.

Declarations

Author contributions

VS: Conceptualization, Visualization, Methodology, Investigation, Data curation, Writing—original draft, Writing—review & editing. SK: Writing—review & editing. VH: Writing—review & editing.

Conflicts of interest

Part of the manuscript has been presented as a poster presentation in the International Diabetes Federation Congress 2025 and the preparation of this manuscript has not been influenced by the presentation or any other factors.

Ethical approval

Not applicable.

Consent to participate

Not applicable.

Consent to publication

Not applicable.

Availability of data and materials

The raw data supporting the conclusions of this manuscript will be made available by the authors, without undue reservation, to any qualified researcher.

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References

1. Khan MAB, Hashim MJ, King JK, Govender RD, Mustafa H, Al Kaabi J. Epidemiology of Type 2 Diabetes — Global Burden of Disease and Forecasted Trends. *J Epidemiol Glob Health*. 2020;10:107–11. [DOI] [PubMed] [PMC]
2. Schwingshackl L, Hoffmann G, Lampousi AM, Knüppel S, Iqbal K, Schwedhelm C, et al. Food groups and risk of type 2 diabetes mellitus: a systematic review and meta-analysis of prospective studies. *Eur J Epidemiol*. 2017;32:363–75. [DOI] [PubMed] [PMC]
3. Shivam V, Gillies CL, Goff LM, Zaccardi F, Khunti K. Taste perception genomics in gestational diabetes mellitus: A systematic review. *Diabetes Obes Metab*. 2024;26:1544–7. [DOI] [PubMed]
4. Shivam V. A meta-analysis on polymorphic trait of taste perception mediated by TAS2R38 genotype. *Exp Clin Psychopharmacol*. 2024;32:497–505. [DOI] [PubMed]
5. Murovets VO, Sozontov EA, Zachepilo TG. The Effect of the Taste Receptor Protein T1R3 on the Development of Islet Tissue of the Murine Pancreas. *Dokl Biol Sci*. 2019;484:1–4. [DOI] [PubMed]
6. Diószegi J, Llanaj E, Ádány R. Genetic Background of Taste Perception, Taste Preferences, and Its Nutritional Implications: A Systematic Review. *Front Genet*. 2019;10:1272. [DOI] [PubMed] [PMC]
7. Fujikura K. Multiple loss-of-function variants of taste receptors in modern humans. *Sci Rep*. 2015;5:12349. [DOI] [PubMed] [PMC]
8. Li C, Li Y, Sun Q, Abdurehim A, Xu J, Xie J, et al. Taste and its receptors in human physiology: A comprehensive look. *Food Front*. 2024;5:1512–33. [DOI]
9. Young RL, Chia B, Isaacs NJ, Ma J, Khoo J, Wu T, et al. Disordered control of intestinal sweet taste receptor expression and glucose absorption in type 2 diabetes. *Diabetes*. 2013;62:3532–41. [DOI] [PubMed] [PMC]
10. Hartman-Petrycka M, Knefel G, Lebedowska A, Nowak M, Błońska-Fajfrowska B. Taste perception and food preferences in patients with diabetic foot ulcers before and after hyperbaric oxygen therapy. *Nutr Diabetes*. 2022;12:41. [DOI] [PubMed] [PMC]
11. Gondivkar SM, Indurkar A, Degwekar S, Bhowate R. Evaluation of gustatory function in patients with diabetes mellitus type 2. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod*. 2009;108:876–80. [DOI] [PubMed]
12. Chamoun E, Liu AS, Duizer LM, Feng Z, Darlington G, Duncan AM, et al. Single nucleotide polymorphisms in sweet, fat, umami, salt, bitter and sour taste receptor genes are associated with gustatory function and taste preferences in young adults. *Nutr Res*. 2021;85:40–6. [DOI] [PubMed]
13. Canivenc-Lavier MC, Kouidhi W, Boudalia S, Folia M. Taste and endocrine disruption. *Ann Endocrinol (Paris)*. 2025;86:101768. [DOI] [PubMed]
14. Shivam V. Diabetes Mellitus and Taste Receptors Expression: A Systematic Review [Internet]. [cited 2025 Jul 7]. Available from: <https://www.crd.york.ac.uk/PROSPERO/view/CRD42022351880>
15. Ketterer C, Müssig K, Heni M, Dudziak K, Randrianarisoa E, Wagner R, et al. Genetic variation within the *TRPM5* locus associates with prediabetic phenotypes in subjects at increased risk for type 2 diabetes. *Metabolism*. 2011;60:1325–33. [DOI] [PubMed]
16. Bartáková V, Kuricová K, Zlámál F, Bělobrádková J, Kaňková K. Differences in food intake and genetic variability in taste receptors between Czech pregnant women with and without gestational diabetes mellitus. *Eur J Nutr*. 2018;57:513–21. [DOI] [PubMed]

17. Yang Y, Luo BR, Hu M, Zhao DM, Jing WJ. Association of CD36 gene single nucleotide polymorphism with gestational diabetes mellitus in Chinese Han population. *Clin Exp Obstet Gynecol*. 2018;45: 266–71. [\[DOI\]](#)
18. Li X, Lai L, Su J, Chen S, Lin S, Wang B, et al. Novel association between a transient receptor potential cation channel subfamily M member 5 expression quantitative trait locus rs35197079 and decreased susceptibility of gestational diabetes mellitus in a Chinese population. *J Diabetes Investig*. 2021;12: 2062–70. [\[DOI\]](#) [\[PubMed\]](#) [\[PMC\]](#)
19. Page MJ, McKenzie JE, Bossuyt PM, Boutron I, Hoffmann TC, Mulrow CD, et al. The PRISMA 2020 statement: an updated guideline for reporting systematic reviews. *BMJ*. 2021;372:n71. [\[DOI\]](#) [\[PubMed\]](#) [\[PMC\]](#)
20. Leprêtre F, Linton KJ, Lacquemant C, Vatin V, Samson C, Dina C, et al. Genetic study of the CD36 gene in a French diabetic population. *Diabetes Metab*. 2004;30:459–63. [\[DOI\]](#) [\[PubMed\]](#)
21. Corpeleijn E, van der Kallen CJ, Kruijschoop M, Magagnin MG, de Bruin TW, Feskens EJ, et al. Direct association of a promoter polymorphism in the CD36/FAT fatty acid transporter gene with Type 2 diabetes mellitus and insulin resistance. *Diabet Med*. 2006;23:907–11. [\[DOI\]](#) [\[PubMed\]](#)
22. Dotson CD, Zhang L, Xu H, Shin YK, Vigues S, Ott SH, et al. Bitter taste receptors influence glucose homeostasis. *PLoS One*. 2008;3:e3974. [\[DOI\]](#) [\[PubMed\]](#) [\[PMC\]](#)
23. Banerjee M, Gautam S, Saxena M, Bid HK, Agrawal CG. Association of *CD36* gene variants rs1761667 (G>A) and rs1527483 (C>T) with Type 2 diabetes in North Indian population. *Int J Diabetes Mellitus*. 2010;2:179–83. [\[DOI\]](#)
24. Wang Y, Zhou XO, Zhang Y, Gao PJ, Zhu DL. Association of the *CD36* gene with impaired glucose tolerance, impaired fasting glucose, type-2 diabetes, and lipid metabolism in essential hypertensive patients. *Genet Mol Res*. 2012;11:2163–70. [\[DOI\]](#) [\[PubMed\]](#)
25. Gautam S, Agrawal CG, Banerjee M. *CD36* gene variants in early prediction of type 2 diabetes mellitus. *Genet Test Mol Biomarkers*. 2015;19:144–9. [\[DOI\]](#) [\[PubMed\]](#)
26. Tabur S, Oztuzcu S, Duzen IV, Eraydin A, Eroglu S, Ozkaya M, et al. Role of the transient receptor potential (TRP) channel gene expressions and TRP melastatin (TRPM) channel gene polymorphisms in obesity-related metabolic syndrome. *Eur Rev Med Pharmacol Sci*. 2015;19:1388–97. [\[PubMed\]](#)
27. Park S, Zhang X, Lee NR, Jin HS. *TRPV1* Gene Polymorphisms Are Associated with Type 2 Diabetes by Their Interaction with Fat Consumption in the Korean Genome Epidemiology Study. *J Nutrigenet Nutrigenomics*. 2016;9:47–61. [\[DOI\]](#) [\[PubMed\]](#)
28. Zhang D, Zhang R, Liu Y, Sun X, Yin Z, Li H, et al. *CD36* gene variants is associated with type 2 diabetes mellitus through the interaction of obesity in rural Chinese adults. *Gene*. 2018;659:155–9. [\[DOI\]](#) [\[PubMed\]](#)
29. Fujii R, Hishida A, Suzuki K, Imaeda N, Goto C, Hamajima N, et al. Cluster of differentiation 36 gene polymorphism (rs1761667) is associated with dietary MUFA intake and hypertension in a Japanese population. *Br J Nutr*. 2019;121:1215–22. [\[DOI\]](#) [\[PubMed\]](#)
30. Mrag M, Hamdouni H, Gouiaa A, Omezzine A, Ben Amor F, Kassab A. Investigation of carbonic anhydrase 6 gene polymorphism rs2274327 in relation to the oral health status and salivary composition in type 2 diabetic patients. *Acta Odontol Scand*. 2020;78:560–4. [\[DOI\]](#) [\[PubMed\]](#)
31. Hatmal MM, Alshaer W, Mahmoud IS, Al-Hatamleh MAI, Al-Ameer HJ, Abuyaman O, et al. Investigating the association of *CD36* gene polymorphisms (rs1761667 and rs1527483) with T2DM and dyslipidemia: Statistical analysis, machine learning based prediction, and meta-analysis. *PLoS One*. 2021;16:e0257857. [\[DOI\]](#) [\[PubMed\]](#) [\[PMC\]](#)
32. Touré M, Samb A, Sène M, Thiam S, Mané CAB, Sow AK, et al. Impact of the interaction between the polymorphisms and hypermethylation of the *CD36* gene on a new biomarker of type 2 diabetes mellitus: circulating soluble CD36 (sCD36) in Senegalese females. *BMC Med Genomics*. 2022;15:186. [\[DOI\]](#) [\[PubMed\]](#) [\[PMC\]](#)

33. Franzago M, Borrelli P, Di Nicola M, Stuppia L, Vitacolonna E. Genetic Variants in *CD36* Involved in Fat Taste Perception: Association with Anthropometric and Clinical Parameters in Overweight and Obese Subjects Affected by Type 2 Diabetes or Dysglycemia—A Pilot Study. *Nutrients*. 2023;15:4656. [DOI] [PubMed] [PMC]
34. Lee KW, Shin D. Interactions between Bitter Taste Receptor Gene Variants and Dietary Intake Are Associated with the Incidence of Type 2 Diabetes Mellitus in Middle-Aged and Older Korean Adults. *Int J Mol Sci*. 2023;24:2199. [DOI] [PubMed] [PMC]
35. Husami SF, Kaur T, Gupta L, Rastogi G, Singh L, Meena P, et al. Corporate genome screening India (CoGsI) identified genetic variants association with T2D in young Indian professionals. *Sci Rep*. 2025; 15:506. [DOI] [PubMed] [PMC]
36. Ebba S, Abarintos RA, Kim DG, Tiyouh M, Stull JC, Movalia A, et al. The examination of fatty acid taste with edible strips. *Physiol Behav*. 2012;106:579–86. [DOI] [PubMed] [PMC]
37. Yoo M, Shin J, Kim J, Ryall KA, Lee K, Lee S, et al. DSigDB: drug signatures database for gene set analysis. *Bioinformatics*. 2015;31:3069–71. [DOI] [PubMed] [PMC]
38. Abrol R, Tan J, Hui H, Goddard WA III, Pandol SJ. Structural basis for bitter taste receptor activation and its potential role in targeting diabetes. *Funct Foods Health Dis*. 2015;5:117–25. [DOI]