





Deciphering the liver enigma: distinguishing drug-induced liver injury and metabolic dysfunction-associated steatotic liver disease—a comprehensive narrative review

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Abstract

Drug-induced liver injury (DILI) poses a complex and heterogeneous clinical challenge, which often resembles non-drug related acute or chronic liver diseases, such as metabolic dysfunction-associated steatotic liver disease (MASLD). Furthermore, certain drugs can induce hepatic steatosis, which is considered a rare variant of hepatotoxicity. Additionally, the detection and diagnosis of DILI in patients with non-alcoholic liver disease present additional challenges that require attention. The importance of achieving an accurate diagnosis is highlighted by the different therapeutic approaches needed for each of these diseases. Nonetheless, as definitive diagnostic tests and distinct biomarkers often remain elusive, the differential diagnosis must rely on a combination of clinical, biochemical, histological, and immunophenotypic profiling. The diagnosis of hepatotoxicity is predicated upon the temporal nexus between the administration of a potentially hepatotoxic drug and the onset of hepatic injury, concomitantly excluding alternative hepatic pathologies. More frequently, this condition presents an acute course, with a more pronounced elevation of cytolytic and cholestatic parameters as compared to fatty liver disease. Advances in elucidating the underlying mechanisms hold promise for bolstering the diagnosis and management of these conditions. This article aims to thoroughly examine and emphasize the currently available scientific evidence to provide valuable insights into the diagnostic strategies for DILI, metabolicassociated liver disease, and drug-induced steatosis (DIS).

Keywords

Drug-induced liver injury, metabolic dysfunction-associated steatotic liver disease, non-alcoholic fatty liver disease, drug-induced steatosis

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Introduction

Drug-induced liver injury (DILI) and herb-induced liver injury (HILI) is a growing health problem, often difficult to differentiate from other forms of liver disease [1]. Liver toxicity is classically divided into intrinsic (predictable, dose-related), idiosyncratic (unpredictable, non-dose-related), and more recently an indirect type of liver damage has been described [2]. Although not fully understood, the pathogenesis of idiosyncratic DILI (iDILI) is based on the interplay between drug characteristics and environmental and host factors, where the genetic background and the immune system have a significant role to play [3]. The severity and outcome of the liver damage are highly variable, ranging from an asymptomatic rise of liver enzymes to a more severe injury that can evolve to acute liver failure [4-6].

Diagnosis of iDILI is based on establishing a temporal association between drug exposure and the onset of signs and symptoms of liver disease, and ruling out alternative etiologies of liver injury. The capacity of drugs to induce different phenotypes of liver damage, makes it frequently indistinguishable from other liver injuries such as metabolic dysfunction-associated steatotic liver disease (MASLD), formerly non-alcoholic fatty liver disease (NAFLD) [7].

Up to date, MASLD is a major cause of liver disease worldwide, with an estimated prevalence of 25% [8]. It is characterized by triglyceride deposition in hepatocytes and defined as the presence of macrovesicular steatosis after the exclusion of significant alcohol consumption and other secondary causes as steatogenic drugs [9]. MASLD can lead to metabolic dysfunction-associated steatohepatitis (MASH), former non-alcoholic steatohepatitis (NASH), cirrhosis, and hepatocellular carcinoma, and diagnosis is based on imaging, serologic, and liver biopsy findings.

The relationship between MASLD and DILI is hard to discern (Figure 1). First, although robust evidence is lacking, MASLD may increase the risk of DILI. What is certain is that a toxic event may induce a more severe liver injury in patients with underlying liver disease, especially in patients with cirrhosis [10]. Second, a variable proportion of patients with iDILI have data of steatosis in combination with other types of histologic findings in liver biopsy. In the US drug-induced liver injury network (DILIN), up to 26% of cases with available histological information showed some degree of steatosis, while this condition was found in less than 2% of the Spanish DILI Registry patients [11, 12]. Whether steatosis in DILI cases is induced by the drug or was previously present is difficult to elucidate if no liver studies were available before DILI onset.

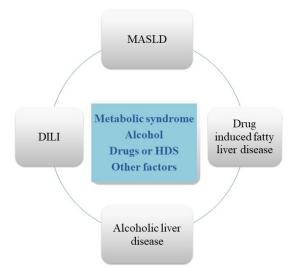


Figure 1. Multidirectional relationship between NAFLD, drug-induced liver disease, drug induced steatosis and alcohol. First, liver steatosis may increase the risk of DILI induced by certain drugs, a variable prevalence of steatosis is present in patients with DILI, and hepatotoxicity may induce a more severe liver injury in patients with MASLD. Besides, some drugs as tamoxifen or methotrexate, can be the cause of drug-induced steatosis/steatohepatitis (DIS/DISH). Finally, alcohol can be another source of confusion as a cause of fatty liver disease, aggravating metabolic steatosis, and as a contributing factor to DILI associated with specific drugs. HDS: herbal and dietary supplements

On the other hand, DIS/DISH is considered as a rare type of DILI. It has been estimated that only 2% of MASLD cases are caused by drugs [13]. This adverse event can induce an acute severe injury or can evolve into a chronic liver disease which can be confused with MASLD [14]. While DIS/DISH may exhibit histological and imaging similarities to MASLD, their natural history, pathophysiology, and prognosis often differ and are influenced by factors such as the specific drug, length of exposure, and individual susceptibility. Nevertheless, the differential diagnosis between iDILI and MASLD is sometimes troublesome [15].

Alcohol can be another source of confusion as a cause of fatty liver disease, aggravating metabolic steatosis, and as a contributing factor to DILI associated with specific drugs such as halothane, methotrexate and isoniazid [15].

Finally, the detection of DILI in patients with MASLD plays an important role not only in routine clinical practice, but also in the setting of clinical trials, especially in MASLD trials, where baseline abnormal liver tests may confound the detection and risk assessment of DILI [16].

The aim of this article is to review the available evidence and provide guidance for the diagnosis of iDILI, MASLD, and DIS.

DILI

Drugs and HDS can induce a number of changes in the functionality and structure of the hepatobiliary system, most of which are non-specific. Moreover, the clinical presentation of DILI is very heterogeneous and may mirror acute or chronic liver damage due to other aetiologies [3]. As a result, the diagnosis of DILI is particularly challenging, as it requires a comprehensive assessment to rule out alternative etiologies of liver injury (Figure 2) [15].

Liver injury: clinical and pharmacological history

Temporal relationship

- · Positive temporal relationship between drug or HDS and liver injury
- Dechallenge: improvement of liver injury after withdrawal of the drug or HDS

Exclusion of other causes of liver disease

- Viral hepatitis
- Autoimmune hepatitis
- Metabolic liver diseases
- Alcohol associated liver disease
- Cholestatic diseases
- · Sepsis, ischemic hepatitis, cancer, biliary obstruction

Phenotypic characterization

• LiverTox (http://livertox.nlm.nih.gov)

Rechallenge

· Liver injury after readministration of the drug/HDS

RUCAM and/or RECAM scales

Liver biopsy

- Uncertain cases
- Prognostic evaluation

Figure 2. DILI diagnostic algorithm with a point-by-point assessment strategy. RECAM: revised electronic causality assessment method; RUCAM: Roussel Uclaf Causality Assessment Method

Clinical manifestations

Symptoms and signs of DILI, when present, include fatigue, weakness, right upper quadrant pain, nausea, dark urine, pruritus, fever, and rash. Severe cases may manifest with jaundice, encephalopathy, ascites, or bleeding [17, 18]. An exhaustive examination and clinical assessment are necessary to rule out other conditions that may lead to liver damage as alcohol consumption, total parenteral nutrition, heart failure, hypotension, sepsis, or cancer [18].

Medication history

When DILI is suspected, a detailed pharmacological history must be taken of all drugs and/or HDS consumed in the last 6 months; although some drugs have longer latency as nitrofurantoin, methotrexate, minocycline, or statins [18, 19]. In addition, medical history should include the start and stop dates of suspected agents, possible dose changes, and previous exposure. Besides, it is essential to collect the clinical-analytical course after discontinuation of the drug (dechallenge) and possible re-exposure effect (rechallenge). An improvement after dechallenge or worsening after rechallenge would support the diagnosis of DILI [20].

The LiverTox website (https://livertox.nih.gov/) is a useful tool for DILI assessment, providing an updated review of over 1,000 drugs and HDS. In addition, it collects information on latency, the pattern of liver injury, and the LiverTox likelihood scale of liver injury for a particular agent; highly useful data when multiple aetiological agents are involved [21].

Initial laboratory assessment

DILI is generally detected by the presence of abnormal liver biochemical blood tests including alanine aminotransferase (ALT), aspartate aminotransferase (AST), alkaline phosphatase (ALP), total bilirubin levels (TBL), and direct bilirubin levels. Additionally, serum albumin levels and the international normalized ratio (INR) serve as indicators of disease severity [20].

For the iDILI case definition, at least one of the following criteria is necessary: ALT elevation exceeding 5 times the upper limit of normal (ULN), ALP levels surpassing 2 times the ULN, or ALT increase exceeding 3 times the ULN in conjunction with total TBL levels exceeding two times the ULN. In cases where ALT values are unavailable during the recognition of iDILI, AST values can be employed as a reliable substitute. In patients with underlying liver injury, ULN must be replaced by the baseline mean values prior to the initiation of the hepatotoxic agent [15].

The biochemical pattern of liver injury may be of help in the characterization and differential diagnosis of DILI. For this aim, the ratio value (R-value) between ALT and ALP is used, which is defined as the result of dividing ALT/ULN-ALT by ALP/ULN-ALP, using the first serum values available during the event. Liver injury is considered "hepatocellular" if there is an ALT elevation ≥ 5 ULN or if the R-value is ≥ 5 . Alternatively, when there is an ALP elevation ≥ 2 ULN or the R-value is ≤ 2 , the liver injury is termed "cholestatic". The term "mixed" liver injury is used for R values between 2 and 5 [15]. While delineating the specific pattern of DILI aids in differential diagnosis, it is important to note that certain drugs have been linked to multiple clinical profiles [20].

On the other hand, apart from the initial approach, additional laboratory investigations, and imaging studies are needed to exclude alternative etiologies of hepatic injury. This work-up should include the determination of hepatitis B surface antigen, anti-hepatitis B core antibody immunoglobulin M (IgM), hepatitis C ribonucleic acid (RNA), hepatitis A IgM antibody, and hepatitis E IgM antibody (and/or RNA). In addition, the evaluation of ceruloplasmin levels plus 24-hour urine copper (Wilson's disease), autoantibodies and immunoglobulin G (IgG, autoimmune hepatitis), and antimitochondrial antibody levels (primary biliary cholangitis) is recommended [15].

Imaging

Abdominal imaging, such as ultrasonography, may exclude bile duct obstruction, evidence of chronic liver disease or other parenchymal changes (e.g., hepatic steatosis), vascular thrombosis or tumors. In most

cases of iDILI, the abdominal ultrasound is normal. In certain clinical situations, such as persistent abdominal pain or jaundice, further imaging tests, such as computerized tomography (CT) or magnetic resonance cholangiography, should be recommended despite normal abdominal ultrasonography [3].

Liver biopsy

Given the clinical similarity between iDILI and other liver diseases, coupled with the lack of definitive diagnostic tests, liver biopsy serves as a valuable tool in discerning certain cases of hepatotoxicity. This is relevant in the presence of autoimmune features [22, 23] and when no improvement or worsening of the liver injury is observed despite the withdrawal of the culprit drug or HDS. Histological findings may support the diagnosis and/or rule out other possibilities [15]. Furthermore, the biopsy is useful in establishing prognosis, as certain histopathological features, such as ductular reaction, microvesicular steatosis and advanced stages of necrosis, and fibrosis, have been linked to an elevated risk of liver failure and mortality [11].

Causality assessment tools

An objective validated and structured approach would be of help to confidently attribute liver injury to a particular drug. Several causality assessment scales and methods have been developed for this purpose [24]. The DILIN structured expert opinion causality scale categorizes DILI likelihood as "unlikely", "possible", "probable", "very likely" or "definitive", which limitations are poor reproducibility and the limited number of DILI experts [3, 21].

On the other hand, the RUCAM is a well-validated scale that is widely applicable in clinical practice and provides a probability category based on the scoring of different items. Therefore, this scale can be used by experts as well as practicing clinicians [21]. Subsequently, the updated RUCAM was developed and it has been enhanced through the provision of a refined definition of the elements to be considered and greater precision in data elements, thereby facilitating the exclusion of alternative causes [25].

In addition, a revised electronic version of the RUCAM scale, the RECAM scale, has recently been described (http://gihep.com/dili-recam/) [26]. This scale might offer certain advantages over RUCAM, as its computerized format, risk factors elimination, simplifies the latency and dechallenge fields, and incorporates the possibility of including data from liver biopsy, rechallenge, and other diagnostic tests. However, in contrast to the RUCAM scale, this scale has not yet been validated. Pending future studies demonstrating its efficacy, it may serve as an additional tool in the assessment of causality in DILI [21].

Genetic testing

In recent years, due to genome-wide association studies (GWAS) and transcriptome-wide association studies (TWAS), several genetic variants related to iDILI have been identified, mainly haplotypes and genotypes of human leucocyte antigen (HLA) (Figure 3) [27–33]. HLA genotyping is available and can be highly helpful in specific clinical situations, since certain HLA alleles have a high negative predictive value (> 95%), allowing a particular drug to be ruled out when multiple agents are involved [3]. Furthermore, thanks to the international collaborative work, a 5-locus genetic risk score has been developed and tested in amoxicillin-clavulanic acid-related DILI patients with a high specificity, supporting a future role for genetic testing in DILI causality assessment and potential risk management [28].

MASLD

MASLD, former NAFLD, represents the prevailing liver disorder on a global scale, impacting 17–46% of adults in Western populations, and its incidence is on the rise [34, 35]. MASLD encompasses a wide spectrum of histopathological features spanning from simple steatosis to varying degrees of inflammation (MASH) and fibrosis [34]. Obesity stands as the primary risk factor for developing MASLD, and it is associated with a multitude of comorbidities including cardiovascular disease, type 2 diabetes mellitus, and malignancy [36].

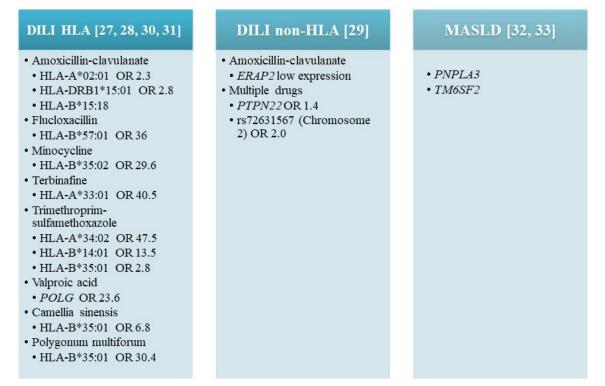


Figure 3. Genetic polymorphisms associated with DILI and MASLD development [27–33]. DILI induced by specific drugs, has been associated with several genetic variants, mainly haplotypes, and genotypes of HLA with different odds ratios. Non-HLA related with DILI. OR: odds ratio; *PNPLA3*: patatin-like phospholipase domain-containing 3; *PTPN22*: protein tyrosine phosphatase non-receptor type 22; *TM6SF2*: transmembrane 6 superfamily member 2; *POLG*: ploymerase gamma gene; *ERAP2*: endoplasmic reticulum aminopeptidase 2

In the presence of environmental and genetic factors, the interaction between the liver, adipose tissue, and the gastrointestinal system gives rise to systemic inflammation and insulin resistance. Consequently, there is an augmented supply of fatty acids to the liver and an increased activation of *de novo* lipogenesis. Criteria for NAFLD diagnosis are imaging or biopsy evidence of hepatic steatosis, the exclusion of substantial alcohol consumption (more than 20 g in women and 30 g in men per day) and alternative causes of hepatic steatosis (Figure 4).

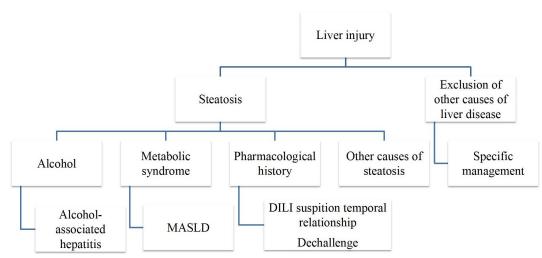


Figure 4. Causes of liver steatosis. Causes of liver steatosis, other than MASLD and alcohol include drugs, autoimmune hepatitis, haemochromatosis, Wilson's disease, coeliac disease, hepatitis C virus, starvation, A/hypo-betalipoproteinaemia, lipoatrophy, hypopituitarism, hypothyroidism, parenteral nutrition or inborn errors of metabolism

In a recent pan-national consensus of hepatology societies, the terms NAFLD and NASH have been redefined. They have been replaced, respectively, by MASLD and MASH to avoid using confusingly exclusive terms such as "non-alcoholic" and potentially stigmatizing terms as "fatty".

Thus, the presence of MASLD should be considered in a patient with established hepatic steatosis and at least one of these five cardiometabolic criteria: (1) body mass index $\ge 25 \text{ kg/m}^2$ (23 Asia) or waist circumference > 94 cm (male), 80 cm (female) or ethnicity adjusted; (2) fasting serum glucose $\ge 5.6 \text{ mmol/L}$ (100 mg/dL) or 2-hour post-load glucose levels $\ge 7.8 \text{ mmol/L}$ ($\ge 140 \text{ mg/dL}$) or HbA1c $\ge 5.7\%$ (39 mmol/L) or type 2 diabetes or treatment for type 2 diabetes; (3) blood pressure $\ge 130/85 \text{ mmHg}$ or specific antihypertensive drug treatment; (4) plasma triglycerides $\ge 1.70 \text{ mmol/L}$ (150 mg/dL) or lipid-lowering treatment; (5) plasma HDL-cholesterol $\le 1.0 \text{ mmol/L}$ (40 mg/dL) (male) and $\le 1.3 \text{ mmol/L}$ (50 mg/dL) (female) or lipid-lowering treatment. In the event that an additional cause of hepatic steatosis was identified, a combined aetiology would then be considered. A new category, outside of pure MASLD, called MetALD (MASLD and alcohol liver disease) was selected to describe people with MASLD who consume larger amounts of alcohol per week (> 140 g per week in females and > 210 g per week in males) [7].

Less frequent causes of liver steatosis, other than MASLD, alcohol or drugs, include autoimmune hepatitis, haemochromatosis, Wilson's disease, coeliac disease, hepatitis C virus-associated fatty liver (genotype 3), starvation, A/hypo-betalipoproteinaemia, lipoatrophy, hypopituitarism, hypothyroidism, parenteral nutrition or inborn errors of metabolism (such as lysosomal acid lipase deficiency) [15]. A liver biopsy is necessary for the definitive diagnosis of MASH [37].

Clinical manifestations

Most patients with MASLD are asymptomatic, although occasionally may refer to asthenia, malaise, and discomfort in the right upper quadrant of the abdomen [38]. In the cases where MASH has progressed to cirrhosis, patients may debut or suffer from any of its complications including hepatocellular carcinoma (HCC). Most often, diagnosis is made after detection of elevated hepatic aminotransferase levels or hepatic steatosis on imaging.

Initial laboratory assessment

Patients with fatty liver disease usually exhibit mild to moderate elevations in ALT or AST levels [38]. When aminotransferases are increased, they are usually two to five times the ULN, with an AST to ALT ratio of less than one [39]. It should be noted that a normal ALT level does not rule out clinically significant histologic injury and the magnitude of aminotransferase elevation does not serve as a predictor of hepatic inflammation or fibrosis severity [40]. ALP levels may be increased less than two to three times the ULN. Ferritin levels or transferrin saturation may be also increased in MASLD patients, and the former has been associated with a higher nonalcoholic fatty liver disease activity score, and with liver fibrosis [41].

Some of the most used serological tests and scoring systems to evaluate MASLD include the Fatty Liver Index (FLI) for evaluation of steatosis, the NAFLD Fibrosis Score (NFS), Fibrosis-4 (FIB-4) Index, and the Enhanced Liver Fibrosis (ELF) test for the assessment of the presence and severity of liver fibrosis in MASLD patients [42].

Imaging

Steatosis must be identified by imaging modalities such as ultrasound, CT or magnetic resonance imaging (MRI).

Qualitative assessment

A meta-analysis comprising 49 studies and 4,720 patients reported a sensitivity of 85% and specificity of 94% for ultrasound in diagnosing steatosis, with liver biopsy serving as the gold standard [43]. However, it has limited sensitivity and does not reliably detect fatty liver disease when less than 20% of steatosis is present or in obese patients [44]. While CT and MRI can effectively identify steatosis, they lack sufficient sensitivity in detecting inflammation or fibrosis.

Quantitative assessment

1H magnetic resonance spectroscopy (1H-MRS) and MRI-proton density fat fraction (PDFF) are the primary techniques for hepatic steatosis quantification and monitoring changes over time [45]. Another imaging

technique that allows the evaluation of steatosis is the controlled attenuation parameter (CAP). Although it has limited ability to discriminate histological grades, it has demonstrated a very good accuracy in assessing steatosis compared with liver biopsies [46]. Ultrasound attenuation imaging (ATI) is an emerging quantitative approach for evaluating hepatic steatosis. In comparison to CAP, this novel technology enables the simultaneous display of B-mode ultrasound images and ATI images on a single screen. This combined visualization facilitates the assessment of liver structure and assists the operator in identifying the optimal measurement zone, thereby enhancing accuracy in steatosis measurement [47].

Liver biopsy

Liver biopsy is still the gold standard for MASLD and MASH diagnosis, and severity prediction. Diagnosis of fatty liver disease is based on the presence of steatosis in more than 5% of hepatocytes, while steatohepatitis is defined by the appearance of hepatocyte ballooning and lobular inflammation. Different histologic patterns have been described in patients with MASLD, zone 3 injury of adult steatohepatitis, zone 1 steatosis with fibrosis observed in children, and steatosis without steatohepatitis criteria [48]. The best histologic predictors of outcome are the fibrosis stage and the presence of ballooning and portal inflammation [49, 50]. To standardize the histologic evaluation of MASLD, the "NAFLD Activity Score" and the "Steatosis Activity Fibrosis" classification methods are recommended to assess disease activity [51, 52]. Albeit this is important information, liver biopsy is an invasive and not risk-free technique. Thus it is indicated for patients with uncertain diagnosis of MASLD or in the setting of clinical trials [53].

Genetic testing

Numerous genetic modifiers of MASLD have been identified [54], yet only a fraction have undergone robust validation. The most well-established genetic association is with *PNPLA3*, the gene responsible for encoding PNPLA3. This association was initially discovered through genome-wide association studies and subsequently confirmed in diverse cohorts, and ethnicities as a modulator of MASLD severity across the entire histological spectrum [32]. Another gene, TM6SF2, has been reported as an additional disease modifier [33], offering potential utility in risk stratification for liver-related and cardiovascular morbidity (Figure 3). However, genetic variants only account for 10–20% of overall heritability [55].

DIS

Definition

DIS refers to the abnormal accumulation of fat in the liver as a result of drug use. It is characterized by an excessive deposition of triglycerides within hepatocytes, leading to hepatocellular lipidosis [56].

DIS is considered a rare DILI phenotype whose incidence and prevalence rates are uncertain. It is estimated that up to 2% of cases diagnosed as MASLD are actually caused by drugs. However, this phenotype may be underdiagnosed, given that differentiation between the two entities can only be made by identifying exposure to a steatogenic drug [13, 15]. Indeed, within the Spanish DILI Registry, the predominant occurrence of steatosis was observed in a limited number of cases with histological data [12], whereas the US DILIN reported steatosis in as many as 26% of cases, varying in severity [11].

Pathogenesis

Hepatocytes have a key role in lipid metabolism. In steatosis liver disease there is an increase in free fatty acids within the hepatocyte. This may be caused by increased uptake (mainly from peripheral tissue, especially adipose tissue, and to a lesser extent from dietary sources), increased *de novo* lipogenesis within the hepatocyte or reduced utilization (either through β -oxidation or secretion) [56]. Although it has been poorly investigated, it is believed that drugs can activate fat accumulation in the liver via different pathways, which include inhibition of mitochondrial function (β -oxidation), increasing insulin resistance, promoting hepatic *de novo* lipogenesis or increasing free fatty acid uptake [57].

Causative agents and patterns of damage

DIS includes pure microvesicular steatosis, mixed microvesicular and macrovesicular, pure macrovesicular steatosis, and steatohepatitis [58].

Microvesicular fatty liver is related to mitochondrial injury mainly associated with drugs such as acetylsalicylic acid (AAS, Reye syndrome), valproic acid, glucocorticoids, NSAIDs, tetracycline, NRTI, and cocaine.

Glucocorticoids, methotrexate, NSAIDs, metoprolol, chlorinated hydrocarbons, 5-fluorouracil, cisplatin, irinotecan, and tamoxifen can induce macrovesicular steatosis by altering lipid metabolism and promoting lipid accumulation in the liver. Besides, amiodarone, valproic acid, and methotrexate can induce mixed macro and microvesicular steatosis. Finally, DISH has been associated with amiodarone, methotrexate, 5-floururacil, cisplatin, irinotecan, and tamoxifen [20].

Metabolic syndrome and its components have been considered as risk factors for the development and severity of DIS in patients treated with methotrexate and tamoxifen [15]. Preventing DIS involves careful monitoring of patients upon treatment with the above-mentioned drugs and treatment is mainly based on the withdrawal of the offending medication, which can lead to the resolution of the liver disease.

Differential diagnosis between DILI, DIS and MASLD

The absence of definite diagnostic methods and biomarkers together with the limits of the causality scales in the assessment of idiosyncratic hepatotoxicity makes diagnosis frequently troublesome and is mainly based on the exclusion of alternative causes of liver disease [11]. This makes differential diagnosis between this entity and nonalcoholic steatohepatitis sometimes extremely difficult, since the intake of drugs is widespread among the general population and the prevalence of metabolic syndrome causing fatty liver in developed countries is increasing. Thus, both processes may overlap. Furthermore, it is not known exactly to what extent the presence of metabolic syndrome with or without nonalcoholic fatty liver disease may increase the risk or influence the prognosis and evolution of an episode of hepatotoxicity. On the other hand, certain agents have the potential to induce or exacerbate fatty liver disease, further contributing to the development or progression of steatosis [58]. Clinical, biochemical, and histological features of DILI, DIS, MASLD, and alcohol-associated hepatitis are described in Table 1 [7, 15, 20, 38, 58–61].

Clinical manifestations

DILI and MASLD can be asymptomatic or paucisymptomatic, thus differential diagnosis based on clinical signs is difficult to make. MASLD is less likely to produce jaundice, choluria or acholia than DILI. In addition, patients with cholestatic DILI may have pruritus, which can be severe, leading to excoriations from scratching. In severe DILI cases, hepatic encephalopathy may develop, indicating acute liver failure [62]. DILI patients may develop hypersensitivity signs and symptoms, such as a fever and rash, Stevens-Johnson syndrome, drug reaction with eosinophilia and systemic symptoms (DRESS) or a mononucleosis-like illness (pseudo mononucleosis). Finally, both MASH and chronic DILI may go on to develop advanced fibrosis or cirrhosis and have signs, and symptoms associated with advanced liver disease or hepatic decompensation.

Initial laboratory assessment

Regarding biochemical blood tests, serum aminotransferase increase in acute DILI can be marked (≥ 25 times the ULN), while elevation of AST and ALT is generally mild or moderate (two to five times the ULN) in MASLD patients. Even more, in general, MASLD does not present as flares of raised liver enzymes over 5 times the ULN or as an acute hepatitis. The study published by Shamseddeen et al. [63] showed a high prevalence of spontaneous liver enzyme abnormalities among patients with cirrhosis due to MASH participating in clinical trials. However, these abnormalities rarely fulfilled the criteria for suspicion of DILI. Besides, total bilirubin may be increased in both hepatocellular and cholestatic hepatotoxic DILI. In contrast, in patients with MASLD, bilirubin is not increased until advanced liver disease stages. Cholestatic DILI can produce an increase of more than two times the ULN of ALP in the absence of other causes (e.g.,

Differential diagnosis	DILI [15, 59]	DIS/DISH [20, 58]	NAFLD/MAFLD [15, 34, 38]	Alcohol-associated hepatitis [61]
Clinical presentation	Acute or chronic	Acute or chronic	Chronic	Acute or chronic
	Multiple phenotypes			
Associated signs and symptoms	Fatigue, weakness, right upper quadrant pain, nausea, jaundice, dark urine, pruritus, fever, and rash	Fatigue, weakness, right upper quadrant pain, nausea, jaundice, dark urine, pruritus, fever, and rash	Asymptomatic	Jaundice
			Asthenia, malaise, and right upper quadrant pain	Ascites, edema, malaise, fever, nepatomegaly, confusion
		AAS		
Drugs, HDS, and alcohol	More than 1,000 drugs and HDS	Acute fatty liver: amiodarone, didanosine, stavudine, valproate, and zalcitabine Drug-associated fatty liver disease: methotrexate, 5- fluorouracil, irinotecan, tamoxifen, corticosteroids, lomitapide, and mipomerson	NAFLD: < 20 g/d (F), < 30 g/d (M) MASLD: < 140 g/w (F), < 210 g/w (M)	Alcohol consumption of more than 40 g/d in women and 50–60 g/d for men, with less than 60 days of abstinence before the onset
	More frequent antibiotics (amoxicillin-clavulanic acid)			
	Check-in LiverTox (https://livertox.nih.gov/) [60]			
Biochemical parameters	ALT > 5 × ULN	Not established	AST and ALT < 5 × ULN	TB > 3 mg/dL (> 50 µmol/L)
	ALT > 3 × ULN + TB > 2		AST/ALT < 1	
	× ULN		Normal ALP and TB	AST/ALT > 1.5
	$ALP > 2 \times ULN$			AST, and ALT > 400 UUI/L
				GGT > 100 U/L
Radiological	No specific findings	Liver steatosis	Liver steatosis	Liver steatosis
findings				Hepatomegaly
Liver biopsy	Acute hepatocellular liver injury: lobular inflammation, portal inflammation, interface hepatitis, apoptosis, granulomas, coagulative necrosis, and confluent or bridging necrosis	Steatosis, steatohepatitis	Steatosis > 5% of hepatocytes, ballooned hepatocytes, lobular inflammation, apoptotic bodies, portal inflammation, perisinusoidal collagen, portal fibrosis, Mallory-Denk, megamitochondria, glycogenated nuclei in periportal hepatocytes, lobular lipogranulomas, PAS-diastase-resistant	Ballooned hepatocytes, Mallory-Denk bodies, neutrophil infiltration, ductular reaction, bilirubinostasis, and pericellular, and sinusoidal fibrosis t
		Microvesicular (mitochondrial injury)		
		Macrovesciular Mixed		
	Cholestatic DILI: acute cholestasis, chronic cholestasis, and acute cholestatic hepatitis		Kupffer cells, and hepatic siderosis	

Table 1. Differences between NAFLD/MAFLD, iDILI, DIS, and alcohol associated hepatitis [7, 15, 20, 34, 38, 58–61]

d: day; F: female; GGT: gamma-glutamyl transpeptidase; M: male; TB: total bilirubin; w: week; PAS: periodic acid-schiff; ×: times

bone pathology), as occurs in patients with MASLD. Once both entities develop cirrhosis, the analytical alterations resulting from cirrhosis are indistinguishable.

Liver biopsy

As mentioned above, histologic diagnosis of MASLD is based on the presence of more than 5% of steatosis in liver tissue, while MASH necessitates the presence of hepatic steatosis accompanied by hepatocyte ballooning degeneration and hepatic lobular inflammation (typically in acinar zone 3). Fibrosis is not an obligatory diagnostic characteristic but may be observed. The morphological appearance of MASH may be histologically indistinguishable from alcoholic steatohepatitis. Additional histological findings in MASH encompass apoptotic (acidophilic) bodies, mild chronic portal inflammation, perisinusoidal collagen, portal fibrosis devoid of perisinusoidal or pericellular fibrosis, Mallory-Denk bodies, megamitochondria, glycogenated (vacuolated) nuclei in periportal hepatocytes (rarely observed in alcoholic steatohepatitis), lobular lipo granulomas, PAS-diastase-resistant Kupffer cells, and hepatic siderosis [64]. Histologic findings

in patients with DILI include acute or chronic hepatocellular injury, acute or chronic cholestasis, steatosis, and steatohepatitis, granulomas, zonal necrosis, signs of hepatic venous outflow obstruction, sinusoidal obstruction syndrome, nodular regenerative hyperplasia, phospholipidosis or peliosis hepatis [20].

New biomarkers

Genetic, epigenetics, and mechanistic biomarkers

Different studies have been conducted to identify new biomarkers for the improvement of DILI and MAFLD detection, and diagnosis [15]. Bonkovsky et al. [65] evaluated non-HLA-related genetic variants associated with common liver diseases as MAFLD (*PNPLA3* and *TM6SF2*) in iDILI patients, finding that none of the genetic polymorphisms tested were significantly associated with the risk of development, severity, or outcome of DILI.

The consortium known as the International Safer and Faster Evidence-based Translation Consortium conducted an evaluation of various biomarkers, leading to the discovery of several potentially valuable candidates for assessing DILI. These include osteopontin (OPN), cytokeratin-18 [total keratin-18 (K18) and caspase cleaved K18 (ccK18)], α -glutathione-S-transferase (α -GST), and miRNA-122 (miR-122), among others. These promising biomarkers show great potential for DILI assessment [66]. Among epigenetic markers, miRNAs are single-stranded RNAs of 21-23 nucleotides that regulate gene expression at epigenetic, transcriptional, and post-transcriptional levels. They in turn influence cell proliferation, differentiation, metabolism, infectious, and non-infectious disease progression, apoptosis, and metastasis. Among these miRNAs, miR-122 is specific to vertebrate species and is highly expressed in the liver, accounting for 70% of all miRNAs, and is important for hepatocyte function. It is also minimally expressed in other organs [67]. miR-122 has been implicated in fatty acid metabolism in mouse studies and its increase in plasma has been linked to the degree of fibrosis in fatty liver disease, viral hepatitis B and C virus hepatitis, and as a marker of liver damage in mice, and in humans with hepatotoxicity [68]. miR-122 has been described as one of the most promising biomarkers in the detection of DILI, whose levels appear to be more specific for liver damage than AST or ALT, with a large intra-individual and inter-individual variability [56]. On the other hand, only miR-122 has been accepted as a circulating biomarker for the diagnosis and prognosis of MASLD. Other studies have found a set of candidate miRNAs to distinguish DIS from MAFLD in animal and in vitro studies, that could be used in drug screening during preclinical development [69–71]. In this way, López-Riera et al. [71] showed that 10 miRNAs [miR-22-5p, -3929, -24-2-5p, -663a, -29a3p, -21 (5p and 3p), -27a-5p, -1260, and -202-3p] were induced in human HepG2 cells and secreted to the culture medium upon incubation with model steatogenic drugs (valproate, doxycycline, cyclosporine A and tamoxifen). Although promising, these findings need to be confirmed in prospective clinical studies.

K18, a structural protein of the cytoskeleton that has a full-length form, and ccK18 are considered as mechanistic biomarkers of liver injury. During hepatocellular necrosis, K18 is passively released from necrotic cells into the blood, while when apoptosis occurs, caspases cleave K18, and release it into the blood. Therefore, early hepatocyte damage could be detected by measuring the levels of K18 (an indicator of necrosis) and ccK18 (an indicator of apoptosis) [72]. Both forms are elevated in the circulation after DILI, showing potential for diagnosis and prognostic use. Although proportional serum levels of K18 and ccK18 may be useful indicators of DILI, elevated levels of these proteins have also been found in patients with hypoxic hepatitis, alcoholic steatohepatitis, MASLD, and other related disorders [73]. In MASLD patients, ccK18 levels have been found to be significantly higher in MASH than in patients with simple steatosis [74] thus ccK18 has been extensively validated as a marker for MASH, with a pooled receiver operating characteristic curve (AUROC) of 0.82 [95% confidence interval (CI), 0.76–0.88], and has been recognized by MASH guidelines as the most promising non-invasive test for the diagnosis and treatment of MASH [75, 76].

Cueto-Sanchez et al. [77] compared the performance of OPN, total K18 and ccK18, α -GST and miR-122 among patients with DILI and other forms of acute liver injury, finding limited specificity of these biomarkers for DILI, except ccK18, that presented the highest potential in distinguishing DILI from autoimmune hepatitis.

Other potential biomarkers for DILI are glutamate dehydrogenase, high mobility group protein B1, and macrophage colony-stimulating factor receptor; however further studies are needed for the validation of these markers [3]. Finally, a recent prospective cohort study has identified several protein biomarkers that could be useful to diagnose and distinguish DILI from any other acute liver injury in clinical practice, with fructose-1,6-bisphosphatase 1 being of particular interest [78].

Immunological biomarkers

Caballano-Infantes et al. [79] analyzed the activation profile [cluster of differentiation 69 (CD69), CD25, and HLA-DR] and natural killer group 2 member D (NKG2D) on iNKT cells and CD4/CD8 T cells in peripheral blood from prospectively collected MASLD and DILI patients compared to controls. This study showed an increase in iNKT cells in MASLD patients compared to DILI subjects. Besides, a positive correlation between CD69⁺iNKT, insulin resistance, AST level, FIB4, and AST to Platelet Ratio Index (APRI) was detected in these patients. DILI cases showed an increase in CD69⁺ and HLA-DR⁺ in both CD4⁺ and CD8⁺ T cells, detecting the most relevant difference in CD69⁺CD8⁺ T cells, concluding that this could be a potential distinctive biomarker for differentiating DILI from MASLD [79].

Microbiota

The potential impact of gut microbiota on the progression of MASLD remains a subject of considerable interest, yet its relevance in DILI is still poorly comprehended. A recent investigation undertook a metagenomic analysis of gut microbiota in MASLD, DILI, and control cohorts, unveiling the key bacterial metabolic pathways associated with these conditions [80]. Notably, MAFLD patients (referring to those recruited with the old NAFLD criteria) exhibited reduced abundances of *Alistipes, Barnesiella, Eisenbergiella, Flavonifractor, Fusicatenibacter, Gemminger, Intestinimonas, Oscillibacter, Parasutterella, Saccharofermentans*, and *Subdoligranulum* compared to DILI patients. Conversely, DILI patients demonstrated diminished abundances of *Acetobacteroides, Blautia, Caloramator, Coprococcus, Flavobacterium, Lachnospira, Natronincola, Oscillospira, Pseudobutyrivibrio, Shuttleworthia, Themicanus, and <i>Turicibacter* in comparison to the MASLD group. Furthermore, the study identified seven bacterial metabolic pathways that were exclusively impaired in DILI, with the majority being associated with metabolic biosynthesis. In the MASLD group, variations in bacterial metabolic pathways in relation to DILI and control groups primarily pertained to fatty acid and lipid biosynthesis. Further confirmation of these findings in larger cohorts of patients could be the basis for the development of individual or panels of microbiome biomarkers to distinguish MASLD and DILI [80].

Detection of DILI on MASLD patients

DILI can occur with lower than stipulated levels of laboratory abnormalities, the reason for the current higher transaminase level threshold for hepatotoxicity diagnosis, is that up to 20% of the general population has mildly elevated liver biochemistries due to alcohol consumption, MASLD or other common disorders [20, 81].

The following criteria are used to make a DILI diagnosis in patients with MASLD: time from exposure to the beginning of the first signs of hepatic impairment, biochemical, and histological markers of hepatic impairment and information on improvements in hepatic function upon therapy discontinuation [82]. Among all the scoring systems, RUCAM is the most accurate to establish the diagnosis of DILI in MASLD patients [83].

The increase of MASLD clinical trials and the higher inclusion of patients with MASLD in trials of therapies for other diseases as diabetes mellitus, has advocated to the development of consensus guidelines for detecting DILI in these patients [84].

Treem et al. [16] published a consensus guideline for the assessment and management of DILI during clinical trials in adults with MASLD and other chronic liver diseases. These authors recommend the establishment of laboratory criteria for DILI detection, knowledge of the underlying disease behavior, to

use more sensitive liver function tests and being aware of potential confounders related to complications of the disease. Additionally, they advocate for the establishment of pretreatment laboratory values as a basis for increased monitoring and for discontinuing drugs if potential DILI arises, using multiples of baseline liver test values and/or a specific threshold value, rather than multiples of ULN.

Conclusions

The distinction between DILI, MASLD, and DIS is crucial for establishing the correct diagnosis and providing the appropriate treatment. Misdiagnosis can lead to inappropriate management, unnecessary tests, and delays in the treatment of these conditions. Actual limitations in the differentiation between these three entities call for studies of clinical, biochemical, and histological data, along with biomarkers and immunophenotyping profiles for a comprehensive phenotyping of DILI, MASLD, and DIS. Nowadays, using clinical evaluation along with the available tests mentioned above, an accurate diagnosis can be reached in most cases. Further comprehension of insights into the underlying mechanistic pathways will lead to the discovery and validation of novel biomarkers that will contribute to improving the diagnosis of these diseases.

Abbreviations

ALP: alkaline phosphatase ALT: alanine aminotransferase AST: aspartate aminotransferase CCK18: caspase cleaved keratin 18 CD69: cluster of differentiation 69 **CT: computerized tomography** DILI: drug-induced liver injury DILIN: drug-induced liver injury network DIS: drug-induced steatosis DISH: drug-induced steatohepatitis HDS: herbal and dietary supplements HLA: human leucocyte antigen iDILI: idiosyncratic drug-induced liver injury IgM: immunoglobulin M K18: keratin-18 MASH: metabolic dysfunction-associated steatohepatitis MASLD: metabolic dysfunction-associated steatotic liver disease MRI: magnetic resonance imaging NAFLD: non-alcoholic fatty liver disease OR: odds ratio PNPLA3: patatin-like phospholipase domain-containing 3 RECAM: revised electronic causality assessment method RNA: ribonucleic acid RUCAM: Roussel Uclaf Causality Assessment Method

TB: total bilirubin *TM6SF2*: transmembrane 6 superfamily member 2 ULN: upper limit of normal

Declarations

Author contributions

MGC: Conceptualization, Writing—original draft, Writing—review & editing. JPTO and AGG: Conceptualization, Writing—original draft.

Conflicts of interest

The authors declare that they have no conflicts of interest.

Ethical approval

Not applicable.

Consent to participate

Not applicable.

Consent to publication

Not applicable.

Availability of data and materials

Not applicable.

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References

- Stephens C, Robles-Diaz M, Medina-Caliz I, Garcia-Cortes M, Ortega-Alonso A, Sanabria-Cabrera J, et al. Comprehensive analysis and insights gained from long-term experience of the Spanish DILI registry. J Hepatol. 2021;75:86–97.
- 2. Hoofnagle JH, Björnsson ES. Drug-induced liver injury types and phenotypes. N Engl J Med. 2019; 381:264–73.
- 3. Andrade RJ, Chalasani N, Björnsson ES, Suzuki A, Kullak-Ublick GA, Watkins PB, et al. Drug-induced liver injury. Nat Rev Dis Primers. 2019;5:58.
- 4. Reuben A, Koch DG, Lee WM; Acute Liver Failure Study Group. Drug-induced acute liver failure: results of a U.S. multicenter, prospective study. Hepatology. 2010;52:2065–76.
- 5. Devarbhavi H, Patil M, Reddy VV, Singh R, Joseph T, Ganga D. Drug-induced acute liver failure in children and adults: results of a single-centre study of 128 patients. Liver Int. 2018;38:1322–9.
- 6. Björnsson E, Jerlstad P, Bergqvist A, Olsson R. Fulminant drug-induced hepatic failure leading to death or liver transplantation in Sweden. Scand J Gastroenterol. 2005;40:1095–101.

- 7. Rinella ME, Lazarus JV, Ratziu V, Francque SM, Sanyal AJ, Kanwal F, et al.; NAFLD Nomenclature consensus group. A multisociety Delphi consensus statement on new fatty liver disease nomenclature. J Hepatol. 2023;79:1542–56.
- 8. Younossi ZM, Koenig AB, Abdelatif D, Fazel Y, Henry L, Wymer M. Global epidemiology of nonalcoholic fatty liver disease—meta-analytic assessment of prevalence, incidence, and outcomes. Hepatology. 2016;64:73–84.
- 9. Ratziu V, Bellentani S, Cortez-Pinto H, Day C, Marchesini G. A position statement on NAFLD/NASH based on the EASL 2009 special conference. J Hepatol. 2010;53:372–84.
- Aller R, Fernández-Rodríguez C, Lo Iacono O, Bañares R, Abad J, Carrión JA, et al. Consensus document. Management of non-alcoholic fatty liver disease (NAFLD). Clinical practice guideline. Gastroenterol Hepatol. 2018;41:328–49.
- 11. Kleiner DE, Chalasani NP, Lee WM, Fontana RJ, Bonkovsky HL, Watkins PB, et al. Hepatic histological findings in suspected drug-induced liver injury: systematic evaluation and clinical associations. Hepatology. 2014;59:661–70.
- 12. Andrade RJ, Lucena MI, Fernández MC, Pelaez G, Pachkoria K, García-Ruiz E, et al. Drug-induced liver injury: an analysis of 461 incidences submitted to the Spanish registry over a 10-year period. Gastroenterology. 2005;129:512–21.
- 13. Grieco A, Forgione A, Miele L, Vero V, Greco AV, Gasbarrini A, et al. Fatty liver and drugs. Eur Rev Med Pharmacol Sci. 2005;9:261–3.
- 14. Allende DS, Kleiner DE. Fatty liver disease that is neither metabolic nor alcoholic. Hum Pathol. 2023; 141:212–21.
- 15. European Association for the Study of the Liver; Clinical Practice Guideline Panel. EASL clinical practice guidelines: drug-induced liver injury. J Hepatol. 2019;70:1222–61.
- 16. Treem WR, Palmer M, Lonjon-Domanec I, Seekins D, Dimick-Santos L, Avigan MI, et al. Consensus guidelines: best practices for detection, assessment and management of suspected acute drug-induced liver injury during clinical trials in adults with chronic viral hepatitis and adults with cirrhosis secondary to hepatitis B, C and nonalcoholic steatohepatitis. Drug Saf. 2021;44:133–65.
- 17. Agarwal VK, McHutchison JG, Hoofnagle JH; Drug-Induced Liver Injury Network. Important elements for the diagnosis of drug-induced liver injury. Clin Gastroenterol Hepatol. 2010;8:463–70.
- 18. Chalasani NP, Maddur H, Russo MW, Wong RJ, Reddy KR; Practice Parameters Committee of the American College of Gastroenterology. ACG clinical guideline: diagnosis and management of idiosyncratic drug-induced liver injury. Am J Gastroenterol. 2021;116:878–98.
- 19. Bessone F, Ferrari A, Hernandez N, Mendizabal M, Ridruejo E, Zerega A, et al. Nitrofurantoin-induced liver injury: long-term follow-up in two prospective DILI registries. Arch Toxicol. 2023;97:593–602.
- 20. Fontana RJ, Liou I, Reuben A, Suzuki A, Fiel MI, Lee W, et al. AASLD practice guidance on drug, herbal, and dietary supplement-induced liver injury. Hepatology. 2023;77:1036–65.
- 21. Fontana RJ, Bjornsson ES, Reddy R, Andrade RJ. The evolving profile of idiosyncratic drug-induced liver injury. Clin Gastroenterol Hepatol. 2023;21:2088–99.
- 22. Suzuki A, Brunt EM, Kleiner DE, Miquel R, Smyrk TC, Andrade RJ, et al. The use of liver biopsy evaluation in discrimination of idiopathic autoimmune hepatitis *versus* drug-induced liver injury. Hepatology. 2011;54:931–9.
- 23. Foureau DM, Walling TL, Maddukuri V, Anderson W, Culbreath K, Kleiner DE, et al. Comparative analysis of portal hepatic infiltrating leucocytes in acute drug-induced liver injury, idiopathic autoimmune and viral hepatitis. Clin Exp Immunol. 2015;180:40–51.
- 24. García-Cortés M, Stephens C, Lucena MI, Fernández-Castañer A, Andrade RJ. Causality assessment methods in drug induced liver injury: strengths and weaknesses. J Hepatol. 2011;55:683–91.
- Danan G, Teschke R. RUCAM in drug and herb induced liver injury: the update. Int J Mol Sci. 2016;17: 14.

- 26. Hayashi PH, Lucena MI, Fontana RJ, Bjornsson ES, Aithal GP, Barnhart H, et al. A revised electronic version of RUCAM for the diagnosis of DILI. Hepatology. 2022;76:18–31.
- 27. Nicoletti P, Aithal GP, Bjornsson ES, Andrade RJ, Sawle A, Arrese M, et al. Association of liver injury from specific drugs, or groups of drugs, with polymorphisms in HLA and other genes in a genome-wide association study. Gastroenterology. 2017;152:1078–89.
- 28. Nicoletti P, Dellinger A, Li YJ, Barnhart HX, Chalasani N, Fontana RJ, et al.; Drug-Induced Liver Injury Network (DILIN); International Drug-Induced Liver Injury Consortium (iDILIC); Prospective European Drug-Induced Liver Injury (Pro-Euro DILI) Investigators. Identification of reduced ERAP2 expression and a novel HLA allele as components of a risk score for susceptibility to liver injury due to amoxicillin-clavulanate. Gastroenterology. 2023;164:454–66.
- 29. Cirulli ET, Nicoletti P, Abramson K, Andrade RJ, Bjornsson ES, Chalasani N, et al. A missense variant in PTPN22 is a risk factor for drug-induced liver injury. Gastroenterology. 2019;156:1707–16.e2.
- 30. Hoofnagle JH, Bonkovsky HL, Phillips EJ, Li YJ, Ahmad J, Barnhart H, et al.; Drug-Induced Liver Injury Network. HLA-B*35:01 and green tea-induced liver injury. Hepatology. 2021;73:2484–93.
- 31. Li C, Rao T, Chen X, Zou Z, Wei A, Tang J, et al. HLA-B*35:01 allele is a potential biomarker for predicting polygonum multiflorum-induced liver injury in humans. Hepatology. 2019;70:346–57.
- 32. Liu YL, Patman GL, Leathart JB, Piguet AC, Burt AD, Dufour JF, et al. Carriage of the *PNPLA3* rs738409 C >G polymorphism confers an increased risk of non-alcoholic fatty liver disease associated hepatocellular carcinoma. J Hepatol. 2014;61:75–81.
- 33. Dongiovanni P, Petta S, Maglio C, Fracanzani AL, Pipitone R, Mozzi E, et al. Transmembrane 6 superfamily member 2 gene variant disentangles nonalcoholic steatohepatitis from cardiovascular disease. Hepatology. 2015;61:506–14.
- 34. Tiniakos DG, Vos MB, Brunt EM. Nonalcoholic fatty liver disease: pathology and pathogenesis. Annu Rev Pathol. 2010;5:145–71.
- 35. Milić S, Stimac D. Nonalcoholic fatty liver disease/steatohepatitis: epidemiology, pathogenesis, clinical presentation and treatment. Dig Dis. 2012;30:158–62.
- 36. Polyzos SA, Kountouras J, Mantzoros CS. Adipose tissue, obesity and non-alcoholic fatty liver disease. Minerva Endocrinol. 2017;42:92–108.
- 37. Ratziu V, Charlotte F, Heurtier A, Gombert S, Giral P, Bruckert E, et al. Sampling variability of liver biopsy in nonalcoholic fatty liver disease. Gastroenterology. 2005;128:1898–906.
- 38. Bacon BR, Farahvash MJ, Janney CG, Neuschwander-Tetri BA. Nonalcoholic steatohepatitis: an expanded clinical entity. Gastroenterology. 1994;107:1103–9.
- 39. McCullough AJ. The clinical features, diagnosis and natural history of nonalcoholic fatty liver disease. Clin Liver Dis. 2004;8:521–33, viii.
- 40. Fracanzani AL, Valenti L, Bugianesi E, Andreoletti M, Colli A, Vanni E, et al. Risk of severe liver disease in nonalcoholic fatty liver disease with normal aminotransferase levels: a role for insulin resistance and diabetes. Hepatology. 2008;48:792–8.
- 41. Kowdley KV, Belt P, Wilson LA, Yeh MM, Neuschwander-Tetri BA, Chalasani N, et al.; NASH Clinical Research Network. Serum ferritin is an independent predictor of histologic severity and advanced fibrosis in patients with nonalcoholic fatty liver disease. Hepatology. 2012;55:77–85.
- 42. Rinella ME, Lominadze Z, Loomba R, Charlton M, Neuschwander-Tetri BA, Caldwell SH, et al. Practice patterns in NAFLD and NASH: real life differs from published guidelines. Therap Adv Gastroenterol. 2016;9:4–12.
- 43. Hernaez R, Lazo M, Bonekamp S, Kamel I, Brancati FL, Guallar E, et al. Diagnostic accuracy and reliability of ultrasonography for the detection of fatty liver: a meta-analysis. Hepatology. 2011;54: 1082–90.
- 44. Fishbein M, Castro F, Cheruku S, Jain S, Webb B, Gleason T, et al. Hepatic MRI for fat quantitation: its relationship to fat morphology, diagnosis, and ultrasound. J Clin Gastroenterol. 2005;39:619–25.

- 45. Runge JH, Smits LP, Verheij J, Depla A, Kuiken SD, Baak BC, et al. MR spectroscopy-derived proton density fat fraction is superior to controlled attenuation parameter for detecting and grading hepatic steatosis. Radiology. 2018;286:547–56.
- 46. Eddowes PJ, Sasso M, Allison M, Tsochatzis E, Anstee QM, Sheridan D, et al. Accuracy of FibroScan controlled attenuation parameter and liver stiffness measurement in assessing steatosis and fibrosis in patients with nonalcoholic fatty liver disease. Gastroenterology. 2019;156:1717–30.
- 47. Tada T, Iijima H, Kobayashi N, Yoshida M, Nishimura T, Kumada T, et al. Usefulness of attenuation imaging with an ultrasound scanner for the evaluation of hepatic steatosis. Ultrasound Med Biol. 2019;45:2679–87.
- 48. Brunt EM, Kleiner DE, Carpenter DH, Rinella M, Harrison SA, Loomba R, et al. NAFLD: reporting histologic findings in clinical practice. Hepatology. 2021;73:2028–38.
- 49. Singh S, Allen AM, Wang Z, Prokop LJ, Murad MH, Loomba R. Fibrosis progression in nonalcoholic fatty liver *vs* nonalcoholic steatohepatitis: a systematic review and meta-analysis of paired-biopsy studies. Clin Gastroenterol Hepatol. 2015;13:643–54.e9.
- 50. Kleiner DE, Brunt EM, Wilson LA, Behling C, Guy C, Contos M, et al.; Nonalcoholic Steatohepatitis Clinical Research Network. Association of histologic disease activity with progression of nonalcoholic fatty liver disease. JAMA Netw Open. 2019;2:e1912565.
- 51. Bedossa P, Poitou C, Veyrie N, Bouillot JL, Basdevant A, Paradis V, et al. Histopathological algorithm and scoring system for evaluation of liver lesions in morbidly obese patients. Hepatology. 2012;56: 1751–9.
- 52. Bedossa P; FLIP Pathology Consortium. Utility and appropriateness of the fatty liver inhibition of progression (FLIP) algorithm and steatosis, activity, and fibrosis (SAF) score in the evaluation of biopsies of nonalcoholic fatty liver disease. Hepatology. 2014;60:565–75.
- 53. Neuschwander-Tetri BA, Clark JM, Bass NM, Van Natta ML, Unalp-Arida A, Tonascia J, et al.; NASH Clinical Research Network. Clinical, laboratory and histological associations in adults with nonalcoholic fatty liver disease. Hepatology. 2010;52:913–24.
- 54. Anstee QM, Targher G, Day CP. Progression of NAFLD to diabetes mellitus, cardiovascular disease or cirrhosis. Nat Rev Gastroenterol Hepatol. 2013;10:330–44.
- 55. Eslam M, George J. Genetic contributions to NAFLD: leveraging shared genetics to uncover systems biology. Nat Rev Gastroenterol Hepatol. 2020;17:40–52.
- 56. Rabinowich L, Shibolet O. Drug induced steatohepatitis: an uncommon culprit of a common disease. Biomed Res Int. 2015;2015:168905.
- 57. Bessone F, Dirchwolf M, Rodil MA, Razori MV, Roma MG. Review article: drug-induced liver injury in the context of nonalcoholic fatty liver disease a physiopathological and clinical integrated view. Aliment Pharmacol Ther. 2018;48:892–913.
- Lammert C, Imler T, Teal E, Chalasani N. Patients with chronic liver disease suggestive of nonalcoholic fatty liver disease may be at higher risk for drug-induced liver injury. Clin Gastroenterol Hepatol. 2019;17:2814–5.
- 59. Aithal GP, Watkins PB, Andrade RJ, Larrey D, Molokhia M, Takikawa H, et al. Case definition and phenotype standardization in drug-induced liver injury. Clin Pharmacol Ther. 2011;89:806–15.
- 60. LiverTox: clinical and research information on drug-induced liver injury [Internet]. Bethesda (MD): National Institute of Diabetes and Digestive and Kidney Diseases; 2012–. [cited 2023 Jan 6]. Available from: https://www.ncbi.nlm.nih.gov/books/NBK547852/
- 61. Bataller R, Arab JP, Shah VH. Alcohol-associated hepatitis. N Engl J Med. 2022;387:2436–48.
- 62. Davern TJ. Drug-induced liver disease. Clin Liver Dis. 2012;16:231–45.
- 63. Shamseddeen H, Vilar-Gomez E, Chalasani N, Myers RP, Subramanian GM, Shlevin HH, et al. Spontaneous fluctuations in liver biochemistries in patients with compensated NASH cirrhosis: implications for drug hepatotoxicity monitoring. Drug Saf. 2020;43:281–90.

- 64. Brunt EM, Tiniakos DG. Histopathology of nonalcoholic fatty liver disease. World J Gastroenterol. 2010;16:5286–96.
- 65. Bonkovsky HL, Severson T, Nicoletti P, Barnhart H, Serrano J, Chalasani N, et al. Genetic polymorphisms implicated in nonalcoholic liver disease or selected other disorders have no influence on drug-induced liver injury. Hepatol Commun. 2019;3:1032–5.
- 66. Church RJ, Kullak-Ublick GA, Aubrecht J, Bonkovsky HL, Chalasani N, Fontana RJ, et al. Candidate biomarkers for the diagnosis and prognosis of drug-induced liver injury: an international collaborative effort. Hepatology. 2019;69:760–73.
- 67. Baffy G. MicroRNAs in nonalcoholic fatty liver disease. J Clin Med. 2015;4:1977–88.
- 68. Kasztelan-Szczerbińska B, Surdacka A, Celiński K, Roliński J, Zwolak A, Miącz S, et al. Prognostic significance of the systemic inflammatory and immune balance in alcoholic liver disease with a focus on gender-related differences. PLoS One. 2015;10:e0128347.
- 69. Russo MW, Steuerwald N, Norton HJ, Anderson WE, Foureau D, Chalasani N, et al. Profiles of miRNAs in serum in severe acute drug induced liver injury and their prognostic significance. Liver Int. 2017; 37:757–64.
- 70. Liu Z, Wang Y, Borlak J, Tong W. Mechanistically linked serum miRNAs distinguish between drug induced and fatty liver disease of different grades. Sci Rep. 2016;6:23709.
- 71. López-Riera M, Conde I, Tolosa L, Zaragoza Á, Castell JV, Gómez-Lechón MJ, et al. New microRNA biomarkers for drug-induced steatosis and their potential to predict the contribution of drugs to non-alcoholic fatty liver disease. Front Pharmacol. 2017;8:3.
- 72. Antoine DJ, Williams DP, Kipar A, Jenkins RE, Regan SL, Sathish JG, et al. High-mobility group box-1 protein and keratin-18, circulating serum proteins informative of acetaminophen-induced necrosis and apoptosis *in vivo*. Toxicol Sci. 2009;112:521–31.
- 73. McGill MR, Jaeschke H. Biomarkers of drug-induced liver injury: progress and utility in research, medicine, and regulation. Expert Rev Mol Diagn. 2018;18:797–807.
- 74. Feldstein AE, Wieckowska A, Lopez AR, Liu YC, Zein NN, McCullough AJ. Cytokeratin-18 fragment levels as noninvasive biomarkers for nonalcoholic steatohepatitis: a multicenter validation study. Hepatology. 2009;50:1072–8.
- 75. Musso G, Gambino R, Cassader M, Pagano G. Meta-analysis: natural history of non-alcoholic fatty liver disease (NAFLD) and diagnostic accuracy of non-invasive tests for liver disease severity. Ann Med. 2011;43:617–49.
- 76. Chalasani N, Younossi Z, Lavine JE, Diehl AM, Brunt EM, Cusi K, et al. The diagnosis and management of non-alcoholic fatty liver disease: practice guideline by the American Association for the Study of Liver Diseases, American College of Gastroenterology, and the American Gastroenterological Association. Hepatology. 2012;55:2005–23.
- 77. Cueto-Sánchez A, Niu H, Álvarez-Álvarez I, López-Longarela B, Del Campo-Herrera E, Ortega-Alonso A, et al. Evaluation of diagnostic and prognostic candidate biomarkers in drug-induced liver injury *vs.* other forms of acute liver damage. Br J Clin Pharmacol. 2023;89:2497–507.
- 78. Ravindra KC, Vaidya VS, Wang Z, Federspiel JD, Virgen-Slane R, Everley RA, et al. Tandem mass tag-based quantitative proteomic profiling identifies candidate serum biomarkers of drug-induced liver injury in humans. Nat Commun. 2023;14:1215.
- 79. Caballano-Infantes E, García-García A, Lopez-Gomez C, Cueto A, Robles-Diaz M, Ortega-Alonso A, et al. Differential iNKT and T cells activation in non-alcoholic fatty liver disease and drug-induced liver injury. Biomedicines. 2022;10:55.
- 80. Rodriguez-Diaz C, Taminiau B, García-García A, Cueto A, Robles-Díaz M, Ortega-Alonso A, et al. Microbiota diversity in nonalcoholic fatty liver disease and in drug-induced liver injury. Pharmacol Res. 2022;182:106348.

- Fontana RJ, Watkins PB, Bonkovsky HL, Chalasani N, Davern T, Serrano J; DILIN Study Group, et al. Drug-induced liver injury network (DILIN) prospective study: rationale, design and conduct. Drug Saf. 2009;32:55–68.
- 82. Marchesini G, Bugianesi E, Forlani G, Cerrelli F, Lenzi M, Manini R, et al. Nonalcoholic fatty liver, steatohepatitis, and the metabolic syndrome. Hepatology. 2003;37:917–23.
- 83. Benichou C, Danan G, Flahault A. Causality assessment of adverse reactions to drugs--II. An original model for validation of drug causality assessment methods: case reports with positive rechallenge. J Clin Epidemiol. 1993;46:1331–6.
- Regev A, Palmer M, Avigan MI, Dimick-Santos L, Treem WR, Marcinak JF, et al. Consensus: guidelines: best practices for detection, assessment and management of suspected acute drug-induced liver injury during clinical trials in patients with nonalcoholic steatohepatitis. Aliment Pharmacol Ther. 2019;49:702–13.