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Rheumatoid arthritis: a complex tale of autoimmune hypersensitivity

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Abstract

Rheumatoid arthritis (RA) is a complex autoimmune disorder characterized by a spectrum of hypersensitivity reactions, encompassing Type II, Type III, and Type IV responses. Firstly, RA is marked by Type II hypersensitivity reactions driven by autoantibodies, such as rheumatoid factor (RF) and anti-(cyclic) citrullinated protein antibodies (ACPAs). These autoantibodies serve not only as serological markers for RA but also actively participate in inflammation, bone erosion, and clinical outcomes, with concurrent activation of the complement system involving C1q, C3, and C5 components specifically linked to RA progression and bone damage. Secondly, RA exhibits traits of Type III hypersensitivity, marked by the formation of immune complexes inciting inflammatory reactions. Immunoglobulin G (IgG) autoantibodies like RF and ACPA play pivotal roles in immune complex formation and the ensuing inflammatory responses. RA also demonstrates Type IV hypersensitivity propelled by CD4⁺ T cells, encompassing T helper 1 (Th1) and Th17 subsets. Th1 cells release interferon (IFN)-γ, promoting proinflammatory cytokines, while Th17 cells secrete IL-17, IL-22, and granulocyte-macrophage colony-stimulating factor (GM-CSF), contributing to synovial inflammation, bone and cartilage damage, and angiogenesis. RA concurrently exhibits features of Type II, Type III, and Type IV hypersensitivity. It is crucial to comprehend the presence and complex interplay of hypersensitivity responses and specific immune cell subsets in RA to create precise and efficient therapeutic approaches for the management of this incapacitating autoimmune condition. Thus, in this review, we aim to provide a comprehensive overview of the hypersensitivity features of RA.

Keywords

Rheumatoid arthritis, hypersensitivity, lymphocytes, autoantibodies, immune complexes

Introduction

Rheumatoid arthritis (RA) is a complex autoimmune disease characterized by chronic joint inflammation caused by the immune system aberrantly targeting synovial cells in the joints. The hallmark of RA is joint-

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related symptoms, including joint swelling and stiffness. Of note, RA is not limited to joint issues alone; it can lead to systemic inflammation affecting various parts of the body. This systemic inflammation can increase the risk of other conditions such as stroke, heart disease, diabetes, and osteoporosis [1, 2].

While the exact cause of RA remains unclear, it is believed to involve a combination of genetic and environmental factors. RA displays a higher prevalence in women compared to men, as is often observed in other autoimmune diseases. This suggests that hormonal factors [3] and gender-specific genetic variables [4] likely contribute to the disease onset. Furthermore, RA tends to be more common among older individuals, underscoring the role of aging in its development [5, 6].

Genetic factors are thought to play a crucial role in the development of RA. For instance, the human leukocyte antigen (*HLA*)-*DRB1* gene has been associated with the production of rheumatoid factor (RF), which is an antibody produced as part of an autoimmune response [7]. Additionally, abnormal activity of peptidyl arginine deiminase (PAD) enzymes can trigger a process known as citrullination, leading to the formation of cyclic citrullinated peptides (CCP). These CCPs can act as autoantigens, provoking an immune response and the production of anti-(cyclic) citrullinated protein antibodies (ACPAs) (Figure 1) [8].

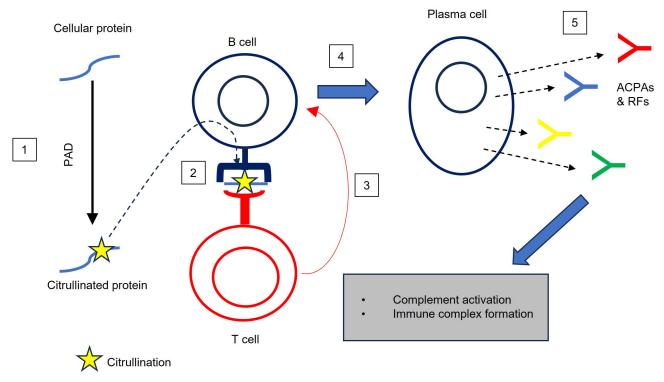


Figure 1. B cell-dependent hypersensitivity. 1: Citrullination of cellular proteins; 2: processing and presentation of citrullinated protein; 3: activation of B cells by T cells; 4: differentiation of naive B cells into plasma cells; 5: production of ACPAs and RFs. ACPAs: anti-(cyclic) citrullinated protein antibodies; PAD: peptidyl arginine deiminase; RFs: rheumatoid factors

In synovium, the infiltration of leukocytes leads to inflammation of the synovial membrane, a condition known as rheumatoid synovitis [9]. This infiltration involves both innate and adaptive immune cells [10], fostering an environment, in which fibroblast-like synoviocytes (FLSs) exhibit invasive and inflammatory behaviors, ultimately resulting in synovial lining hyperplasia [11]. FLSs release enzymes like collagenase and matrix metalloproteinases (MMPs), which contribute to cartilage destruction and the formation of pannus, which can obliterate the joint [12, 13]. The degradation of bone is facilitated by the maturation of pre-osteoclasts into osteoclasts, a CXCR4-dependent process favored by leukocyte infiltration and the release of pro-inflammatory cytokines [14]. Synovial hyperplasia, combined with hypoxia and the cytokines and growth factors generated by infiltrating cells, promotes angiogenesis [15], establishing a feedback loop that sustains ongoing leukocyte infiltration and chronic inflammation.

The distinctive feature of RA-associated inflammation is the recruitment of various immune cells, including monocytes, macrophages, neutrophils, B cells, and CD4⁺ and CD8⁺ T cells, to the synovial compartment [16]. CD4⁺ and CD8⁺ T cells within the synovium produce pro-inflammatory cytokines and chemokines that significantly contribute to the pathogenesis of RA [17–19]. It is widely recognized that aberrant inflammatory CD4⁺ T cell activity fundamentally influences the development and progression of RA [20]. Among the CD4⁺ T cell subsets involved in RA, T helper 1 (Th1) and Th17 cells have been identified as prominent contributors [21, 22]. The inflamed synovium is also noted for its enrichment in memory CD4⁺ T cells, which support B cell activity and immunoglobulin (Ig) synthesis [23].

Significantly, RA displays characteristics of both Type II, Type III, and Type IV hypersensitivity reactions simultaneously. It is crucial to comprehend the presence and complex interplay of hypersensitivity responses and specific immune cell subsets in RA to create precise and efficient therapeutic approaches for the management of this incapacitating autoimmune condition. Thus, in this review, we aim to provide a comprehensive overview of the hypersensitivity features of RA.

Type II hypersensitivity in RA

The impact of Type II hypersensitivity in RA

IgG and IgM antibodies are key to humoral immunity. They bind to microbes, such as bacteria, and facilitate their direct elimination through various antibody-dependent mechanisms. Unfortunately, the humoral response can turn against the host when these antibodies recognize self-antigens, potentially causing serious harm. Type II hypersensitivity reactions can be categorized into two subtypes: Type IIa and Type IIb hypersensitivity. Firstly, Type IIa hypersensitivity represents antibody-mediated cytotoxic reactions, where antigen-antibody complexes trigger the complement pathway and generate the membrane attack complex (MAC), leading to the lysis of target cells. Both IgG and IgM antibodies can also bind to Fc receptors (FcRs) on B lymphocytes, macrophages, and dendritic cells, thereby activating antibody-dependent cellular cytotoxicity (ADCC). Secondly, Type IIb hypersensitivity involves antibody-mediated cell-stimulating reactions driven by autoantibodies. For instance, in Graves' disease, antibodies bind to thyroid cells, stimulating the excessive production of thyroid hormone.

Type II hypersensitivity has been associated with RA. Studies have consistently shown elevated levels of complement components, such as C2, C3, C3a, C5a, and the terminal complement complexes C5b-9, in synovial fluid and synovial tissue [24]. This suggests that complement activation plays a pivotal role in the early stages of RA and contributes to the amplification of inflammation as the disease progresses [25].

Autoantibody production in RA

Following the differentiation of plasma cells, autoreactive B lymphocytes, driven by Toll-like receptors (TLR), primarily secrete autoantibodies [26]. The expansion of autoreactive B cells in RA may result from interactions between specific proteins with post-translational modifications and external antigens (Figure 1). Key autoantibodies in RA include RFs, ACPAs, and anti-PAD-4 antibodies [27]. The presence of RF is one of the diagnostic criteria for RA, but it is not definitive on its own and is used in conjunction with other clinical and laboratory findings for diagnosis and disease monitoring, as RF is not present in every RA patient. Among autoantibodies, ACPA holds significant prognostic value for the onset of RA in at-risk patients. ACPAs have emerged as a more specific serological biomarker than RF, which has higher sensitivity but lower specificity. This has led to the development of diagnostic tests based on ACPA detection. Approximately 60 percent to 70 percent of RA patients exhibit ACPAs, which target epitopes modified through citrullination by PAD [28]. Overexpression of PAD in the RA synovium leads to the production of ACPAs against citrullinated antigens. ACPAs are closely associated with inflammatory markers and often appear in serum years before the onset of RA symptoms [29]. They can activate immune cells and upregulate the production of pro-inflammatory cytokines. Studies have observed the involvement of ACPAs in bone erosion, arthralgia (joint pain), and arthritis induction, all common symptoms of RA. Bone erosion is linked to early joint inflammation and correlates with high ACPA levels. However, debates continue regarding whether ACPAs are the primary antibodies responsible for these effects, or if other antibodies contribute. Some studies suggest that ACPAs have an osteoclast-activating effect, enhancing inflammation. Arthralgia, an early symptom of RA, has been examined in models like collagen antibody-induced arthritis (CAIA), highlighting the relationship between antibodies and pain associated with immune complex formation [30]. Notably, ACPA-positive RA patients tend to experience worse clinical outcomes than ACPA-negative individuals, emphasizing the importance of early diagnosis and therapy guided by ACPA markers [31].

The complement system in RA

The complement system, a component of the innate immune system, is responsible for recognizing foreign antigens and clearing dead cells. Complement proteins, primarily synthesized in the liver, can rapidly activate when danger is detected, acting as opsonins, proinflammatory mediators, and lytic agents. The complement system can be activated through three major pathways: classical, lectin, and alternative. Although these pathways differ in their triggering mechanisms, they all lead to C3 cleavage, the release of anaphylatoxins, C3a and C5a, and the formation of the MAC. Studies have indicated a strong connection between the complement system and RA pathogenesis, particularly the pathogenic role of C1q in RA and the involvement of C3 and C5 in disease progression [32–35]. The interplay between the complement system and RA is evident through the induction of osteoporosis due to ACPA detection, complement activation, and subsequent bone damage. The role of complement cascades in RA has been confirmed through observations of C3-deficient and C5-deficient mice, which were found to be resistant to animal models of collagen-induced arthritis (CIA) [36, 37]. As synovial inflammation progresses, both the complement system and TNF- α secretion are activated. Anti-TNF- α treatment results in reduced levels of C3 and C4 complement components and shows potential in reducing cardiovascular disease associated with RA [38]. These findings support the use of anti-TNF- α therapies in the treatment of RA. Using complement receptor 1-related gene/protein y (Crry) targeted by means of CR 2 (CR2-Crry), a complement inhibitor that targets C3 breakdown products deposited in RA-affected joints, has shown promise in reducing inflammation, cartilage degradation, and bone damage [39]. It also significantly reduced the production of anti-collagen IgG and inflammatory cytokines TNF- α and IL-1 β [39]. This approach may offer a therapeutic avenue for RA, although clinical trials of such inhibitors in RA patients have remained without success so far.

Type III hypersensitivity in RA

The impact of Type III hypersensitivity in RA

The formation of immune complexes is a natural part of the humoral response to an antigen. In the body, antigen-antibody complexes are typically removed via phagocytosis, with CRs and FcRs recognizing these complexes. These immune complexes often contain denatured proteins, antigens from gut bacteria, or dead cells and are cleared by macrophages. However, when the immune complex concentration exceeds a certain threshold, inflammatory reactions occur, leading to what is known as Type III hypersensitivity. The pathogenic effects of these immune complexes have been studied to understand their inflammatory consequences.

Type III hypersensitivity, which leads to the formation of immune complexes, is associated with the development of RA [40]. Studies have consistently shown that these immune complexes play a crucial role in the pathogenesis of autoimmune diseases [41]. Autoantibodies, such as RF and ACPAs, are found in RA patients and contribute to inflammation in the joints. These autoantibodies, specifically IgG antibodies, are known to be involved in further inflammatory responses, making them widely present in RA synovial fluids (Figure 1) [42].

Type III hypersensitivity arises from the deposition of immune complexes due to overproduced IgG and IgM reacting against foreign or self-antigens. When an excessive number of immune complexes are formed and cannot be efficiently cleared from tissues, it also triggers the activation of the complement

system (Type II hypersensitivity) and leads to the recruitment of inflammatory mediators, ultimately activating immune cells like basophils and neutrophils in the tissue. These recruitments can persist for hours, weeks, or even years. Immune complexes are a normal occurrence but become problematic when their reaction becomes hypersensitive, resulting in Type III hypersensitivity. This condition results from an overproduction of antigen-antibody immune complexes, a process that often requires complement system activation for phagocyte recruitment to the site of the immune complex. This can occur both via complement-dependent factors, such as C1q and C5a, and complement-independent factors, including P-selectin, Mac-1, substance P, FcRs, and phagocytes [40].

FcRs

The formation and accumulation of immune complexes, specifically those involving the interaction of IgG and IgM, play a crucial role in autoimmune responses. These immune complexes may cause inflammation in the joints, contributing to autoimmune antibody responses [43].

IgG is recognized through $Fc\gamma Rs$ present on various immune cells in the body, which are key components of both the humoral and cellular immune systems. The activation and downregulation of the immune system are mediated through $Fc\gamma Rs$. These receptors are diverse, each having different specificities for isotypes [40].

Activation signals are associated with a common FcR γ chain, containing an immunoreceptor tyrosinebased activation motif (ITAM). Activation signals induce phosphorylation of tyrosine residues in the ITAM and initiate a cascade of responses, including the release of cytokines by macrophages and ADCC mediated by natural killer (NK) cells. The binding of IgG to FcγRIIB, which harbors an intracellular immunoreceptor tyrosine-based inhibitory motif (ITIM), regulates immune cell activity. The co-expression of B-cell receptors (BCRs) and FcγRIIB on B cells enables them to bind to IgG and antigens forming immune complexes. Dysregulation of FcγRs is associated with RA, making these receptors crucial in the protection against IgGcomplexed antigens and the development of RA [44].

The immune complexes are also formed when an $Fc\gamma R$ binds with IgG and complexes with antigens, as evidenced by the development of animal models for arthritis [45]. In autoimmune arthritis, $Fc\gamma Rs$ play a critical role, with some receptors inducing arthritis (e.g., $Fc\gamma RI$ and $Fc\gamma RIII$) and others suppressing it (e.g., $Fc\gamma RIIB$). The expression level of the $Fc\gamma RIIA$ gene has been observed to be abnormal in RA patients, and it can affect RA activity, whereas the increase in expression of $Fc\gamma RIIB$ can affect stabilizing $Fc\gamma R$ function in patients [46]. This receptor also influences the uptake of immune complexes and may be used in the therapeutic progress of RA. Therefore, molecules targeting FcRs have become a potential approach in RA treatment [47].

Immune complexes are internalized through the action of $Fc\gamma Rs$, and immune cells, particularly macrophages, are crucial for the uptake of antigen-antibody complexes. The binding of immune complexes by $Fc\gamma Rs$ leads to the activation of phagocytes and the release of inflammatory cytokines. These antigen-antibody complexes can promote cellular immune responses, often having a stronger effect than responses initiated by antigens alone. The involvement of $Fc\gamma Rs$ extends to influencing bone erosion in RA, a common consequence of the disease [48]. Different types of $Fc\gamma Rs$ are associated with various aspects of RA. The absence of $Fc\gamma Rs$ in models has been used to study their role, with the FcR γ chain being associated with disease development [45].

Type IV hypersensitivity in RA

The impact of Type IV hypersensitivity in RA

Type IV hypersensitivity, also known as delayed-type hypersensitivity, manifests slowly, typically taking a day or more to develop [49]. Unlike other (I-III) hypersensitivity types, it is a cell-mediated, not an antibody-mediated reaction. Key players in this response include T lymphocytes, monocytes, and macrophages. This reaction is initiated when antigens are presented on class II MHC by antigen-presenting cells, leading CD4⁺ T cells to recognize the complex and trigger a hypersensitivity response. Macrophages

release the cytokine IL-12, promoting the expansion of additional CD4⁺ T cells [50]. In cases of intracellular pathogens, activated macrophages produce hydrolytic enzymes and transform into multinucleated giant cells [51]. However, an exaggerated response by helper T cells, along with excessive cytokine production, can damage tissues, leading to inflammation, and cell death. Genetic (e.g., HLA haplotype) and environmental factors may cause enzyme modification, failure of tolerance, and upregulated lymphocyte activation, inducing Th1 and Th17 responses to self-antigens. This contributes to the influx of T lymphocytes into the joints [52].

Effector function of Th1 cells in RA

RA was identified as a Th1-driven disease, as it was found that CD4⁺ T cells inclined to release interferon (IFN)- γ are present in the synovial fluids of RA patients [51]. This subset of CD4⁺ T cells expresses CXCR3, which binds the chemokines CXCL9 and CXCL10 induced by IFN- γ . In RA, synovial fluids exhibit higher concentrations of CXCL9 and CXCL10, and CD4⁺ T cells in these fluids express CXCR3 [53]. Notably, most CD4⁺ T cells in synovial joints are of the memory phenotype (CD45RO⁺), but their antigen specificity remains uncertain. Th1 cells typically stimulate macrophage activation, characterized by an increased capacity to release pro-inflammatory cytokines such as TNF- α , especially in the synovial joint [54]. Investigating whether Th1 cells can alter the characteristics of local macrophages is crucial, as it might contribute to disease maintenance (Figure 2) [55].

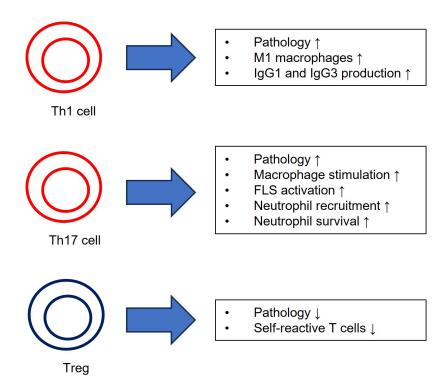


Figure 2. Effector functions of RA-specific T cell subsets. ↑: promoting effect; ↓: inhibitory effect. FLS: fibroblast-like synoviocyte; IgG1: immunoglobulin G1; Th1: T helper 1; Treg: regulatory T cell

Additionally, Th1 cells may influence class switching toward IgG1 and IgG3 antibodies, potentially impacting the antibody profile in RA [56]. Polyclonal antibodies to type II collagen are primarily of the IgG1 and IgG3 subclasses in RA while autoantibodies to citrullinated fibrin are primarily of the IgG1 subclass, indicating a prior interaction with IFN- γ -producing cells [57]. However, this process is influenced by a variety of factors throughout inflammation, not solely by a specific CD4⁺ T-cell subset. While CD8⁺ T cells have been identified in synovial joints, their interaction with CD4⁺ T cells and their activation mechanisms are not yet fully understood [58].

Effector function of Th17 cells in RA

Th17 cells, a distinct subset of T cells, secrete IL-17 independently of cytokines from Th1 or Th2 cells [59]. IL-17 bridges innate and adaptive immunity, exerting both positive and negative effects on immune responses. In addition to IL-17, Th17 cells produce TNF- α , IL-21, IL-22, and granulocyte-macrophage colony-stimulating factor (GM-CSF), making them significant contributors to immune responses [60, 61]. Th17 cells are implicated in the maintenance of immunological homeostasis, such as at mucosal membranes in the intestine, and are increasingly linked to persistent inflammation in RA and other autoimmune diseases [62, 63]. Notably, Th17 cells have been found to play a critical role in the early stages of RA due to immune dysfunction [64]. IL-17 is substantially expressed in the synovial fluid of RA patients, and there is a strong correlation between the proportion of Th17 cells in the synovial fluid and the severity of arthritis inflammation [65, 66]. Furthermore, plasma levels of IL-21 and IL-23, cytokines associated with Th17 cells, strongly correlate with RA severity [67]. Th17 cells release various cytokines, including IL-17A, IL-17F, and IL-22, which stimulate synovial fibroblasts and macrophages to produce proinflammatory substances such as IL-1, IL-6, TNF- α , and prostaglandin E2 (PGE2), exacerbating synovial inflammation [68]. Th17 cells also induce the release of GM-CSF, promoting joint inflammation [69]. IL-17 directly stimulates osteoclastogenesis, leading to bone degradation and cartilage damage [70]. Therefore, Th17 cells contribute to RA pathogenesis by inducing synovial inflammation, pannus development, cartilage and bone degradation, and angiogenesis [71–75]. Additional research into the interactions between Th17 cells and other T-cell subsets can provide insights into the immunological processes underlying RA.

Effector function of regulatory T cells in RA

Regulatory T cells (Tregs), a subset of CD4⁺CD25⁺ lymphocytes expressing FOXP3, play a vital role in maintaining immunological tolerance [76]. Tregs release TGF- β and IL-10 to suppress the immune system, and they express markers like CTLA-4, GITR, and CD39, with changing expressions based on circumstances [77]. Tregs inhibit self-reactive T cells through cell-to-cell contact mediated by substances like CTLA-4, which suppress T-cell activation [78]. In peripheral blood, Tregs can be categorized into naive and activated forms based on CD45RA expression [79]. While FOXP3 is expressed in human effector T cells, its transient upregulation does not restrict it to Tregs. Therefore, assessing Tregs for their ability to inhibit immune responses and demethylation of the FOXP3 Treg-specific demethylated region (TSDR) is essential for their characterization [80]. FOXP3⁺CD25⁺CD4⁺ T cells accumulate in the synovial fluid and tissue of RA patients and exhibit suppressive capacity in vitro, indicating their functionality [81]. However, effector T-cell proliferation and proinflammatory cytokines can reduce Treg function in vitro, a phenomenon expected to occur in situ. It has been observed that introducing TNF- α during co-culture experiments inhibits Treg functions. However, it is not clear whether this is due to direct interactions with Tregs or effector T cells [82, 83]. There have been contradictory reports about the frequency of Tregs in RA patients. More stringent characterization of Tregs has revealed that RA patients have decreased numbers of Treg cells in peripheral blood but increased numbers in synovial fluid, raising the question about the functionality of Tregs in RA [84]. The origin of FOXP3⁺ Tregs in synovial tissues is not well understood; they may develop in situ from conventional T cells (induced Tregs) or originate from the thymus (thymic Tregs) [85]. There is evidence that Tregs with a distinct T cell receptor V gene segment are present in juvenile arthritis patients, indicating that Tregs develop from a separate lineage from conventional T cells [86]. The synovial fluid of RA patients contains cytokines like IL-2 and TGF- β , essential in facilitating the development of Tregs. Treatments that reduce inflammation may improve Treg function, as they have been shown to increase the frequency of Tregs in RA patients. Targeting molecules like IL-2R or IL-6R can shift the balance between Treg and Th17 cells in favor of Tregs [87, 88]. Recent treatment approaches involve administering modest doses of IL-2 to stimulate Treg growth in autoimmune patients, as low IL-2R signaling is crucial for Treg proliferation and development [89, 90]. Clinical trials exploring these treatments for RA are currently underway [91].

Hypersensitivity-associated innate immune cells in RA

Mononuclear phagocytes

The initial phase of differentiation from recently migrated monocytes to mature macrophages takes place within the RA synovial membrane. These subgroups colonize the superficial and deep layers of the lining and the synovial sub-lining and lining layer, respectively. The expression of various activation markers and adhesion molecules highlights the potential functional variations in these regions, which may contribute differentially to the progression of the disease [92].

The synovial joint is a complex structure that connects skeletal elements, facilitating movement. Articular bones are separated by synovial fluid, with articular cartilage covering the corresponding bones. The joint is externally supported by a joint capsule, while the inner synovial membrane encloses the synovial cavity, separating its fluid from exterior joint structures. The synovial membrane consists of a lining layer enriched with macrophages (type A synoviocytes) and synovial fibroblasts (type B synoviocytes). In addition to these, the sub-lining interstitial synovial tissue houses mast cells, adipocytes, blood vessels, lymphocytes, and diverse populations of interstitial macrophages, and fibroblasts [93]. Cytokines produced by macrophages and T cells stimulate synovial fibroblasts, creating feedback loops that polarize and activate macrophages. Synovial fibroblasts respond to TNF- α and IL-1 by producing TNF ligand superfamily member 11 (RANKL) and GM-CSF, crucial for the development of osteoclasts, the bone-resident macrophages [94]. Synovial fibroblasts produce GM-CSF, which possibly enhances the survival and activity of neutrophils and macrophages, promoting arthritis.

Synovial tissue macrophages

Macrophages are among the most abundant cell types in sites of inflammation in RA. Human and mouse synovial tissue macrophages express macrophage markers such as CD64, CD11b, CD68 (human), or F4/80 (mouse). Based on the cell membrane expression of MerTK, a tyrosine-protein kinase receptor involved in efferocytosis and inflammation resolution, and CD206, a C-type lectin mannose receptor functioning as a pattern recognition receptor for pathogens, two primary macrophage populations within the human synovium can be distinguished [16]. These populations include tissue-resident MerTK⁺CD206⁺ macrophages and tissue-infiltrating MerTK⁻CD206⁻ macrophages. In response to arthritis, MerTK⁺CD206⁺ synovial tissue macrophages express CD163. Single-cell RNA sequencing (scRNA-seq) and multiparameter flow cytometry studies reveal numerous transcriptionally unique phenotypic clusters within these two synovial tissue macrophage populations, each with varying functions and distribution across different stages of arthritis progression [95].

Several significant clinical observations have provided initial evidence for the pivotal contribution of macrophages to RA pathogenesis. These include the positive correlation between the abundance of synovial tissue macrophages and the extent of joint damage and the association between a high disease activity score in local joints and the abundance of macrophages in the synovial sub-lining layer [96, 97]. Macrophages play an active role in disease, not only by presenting antigens but also by producing reactive oxygen species (ROS), nitric oxide intermediates, and matrix-degrading enzymes. Macrophages contribute to the maintenance of the inflammatory response by attracting innate immune cells like neutrophils and monocytes and promoting T-cell differentiation [98, 99]. In contrast, RA sustains chronic osteoclast activity, resulting in widespread bone erosion. Macrophages isolated from synovial tissue in RA secrete monocyte chemoattractant protein 1 (MCP-1, also known as CCL2) and CXCL8 (also known as IL-8), essential for recruiting monocytes and neutrophils [100]. They also secrete IL-23 stimulating the growth and activation of Th17 cells [101]. Immunohistology and the analysis of dissociated synovium have indicated that macrophages are the primary producers of TNF- α in inflamed RA joints. The therapeutic efficacy of TNF- α inhibitors in treating RA underlines the significance of macrophages in this condition. The precise mechanisms leading to the overproduction of $TNF-\alpha$ by synovial macrophages remain unclear but may involve various factors such as cytokines, T cells, immune complexes, TLRs, or neutrophil extracellular traps (NETs) [95].

Interestingly, in RA, anti-TNF- α medication has proven more effective than blocking IL-17 signaling, suggesting the involvement of other effector pathways. Clinically effective treatments often reduce the number of macrophages in the affected joints, and complete macrophage elimination has been advantageous in RA animal models. The data suggest that macrophages play a crucial role in RA due to their control over the cytokine environment, promoting inflammation and contributing to bone and cartilage deterioration [100]. Macrophage-produced TNF- α , IL-6, and IL-1 can further enhance osteoclast activities, either directly or indirectly. In healthy individuals, osteoclast activity is episodically stimulated, leading to localized bone erosion, typically at cartilage-pannus junctions [102].

Neutrophils

Th17-associated recruitment of neutrophils

Neutrophils are the most prominent cell type in the synovial fluid of RA patients. In RA joints with inflammation, extensive recruitment of neutrophils occurs. Various cell types, including Th17 cells, neutrophils, and epithelial cells, produce the chemokine CXCL8, which is crucial for neutrophil migration and recruitment. Particularly in RA patients, cytokines produced by Th17 cells significantly enhance neutrophil migration. In the synovial fluid of RA patients, both Th17 cells and neutrophils are abundant. Th17 cells activate neutrophils directly by releasing IL-8, and they also produce IL-17, IL-23, TNF- α , various chemokines, and leukotriene B4 (LTB4), all of which facilitate neutrophil migration [103].

Additionally, the cytokine IL-22, related to Th17, is highly expressed in the synovial tissue of mice and humans with RA induced by antigens. This emphasizes the significant role of Th17-induced production of IL-17, -22, and -23 in the migration and recruitment of neutrophils, thereby contributing to the pathogenesis of RA by intensifying joint inflammation. Recent research has revealed that serum complement activation induces Th17 cell-dependent autoimmune arthritis, primarily through the activation of C5a. This finding underscores the role of Th17 cells in the pathogenesis of RA [104].

Although less prevalent in synovial tissue, they are abundant in the cartilage-pannus junction, where synovial tissue invades cartilage. In areas of inflammation, neutrophils exhibit prolonged activity and can inflict substantial damage on RA joints. Neutrophils in RA synovial fluid secrete two critical cytokines and chemokines, RANKL and TNFSF13B (also known as BLyS or BAFF). These substances are associated with the activation of osteoclasts and B lymphocytes, respectively. Moreover, RA synovial fluid neutrophils exhibit immunological activity similar to that of macrophages and dendritic cells, as they can present antigens to T lymphocytes and express MHC class II molecules [105].

Survival of neutrophils in the synovial joint is promoted by hypoxia and the presence of anti-apoptotic cytokines, such as TNF- α , GM-CSF, IL-6, IL-15, and CXCL8, which can extend their lifespan for several days [106]. Th17 cells, prevalent in RA, produce IL-17, doubling the neutrophil lifespan [107]. IL-1 further delays neutrophil apoptosis, intensifying inflammation [108, 109]. Activated neutrophils pose a threat to host tissues by ROS and degranulating, either into the synovial fluid or directly onto the joint surface. This process is known as frustrated phagocytosis, arising from immune complex deposition on the joint surface, creating an environment where neutrophils release high concentrations of oxidants and cytotoxic chemicals directly onto the articular cartilage surface [110].

Neutrophils promoting inflammation in RA joints

RA is characterized by synovial inflammation and the formation of pannus tissue, largely attributed to the infiltration and activation of neutrophils. Neutrophils in the synovial fluid, when exposed to immune complexes like RF and ACPAs, release ROS and proteases, which degrade hyaluronic acid and activate cytokines. Additionally, neutrophils near cartilage sites encounter immune complex deposition, leading to frustrated phagocytosis, resulting in the release of harmful substances onto the cartilage. Cathepsin G, a serine protease in neutrophil granules, is involved in RA pathogenesis [111]. Notably, RA synovial fluid shows elevated cathepsin G levels and activity, linked to cartilage degradation [112]. Of note, stimulated neutrophils release PAD4. As previously described, PAD4 stimulates and activates adaptive immune cells, resulting in APCA formation which contributes to RA pathogenesis [113].

One remarkable feature of neutrophils in host defense is their ability to combat pathogens through a process called NETosis, which involves the formation of NETs [114]. In RA, NETs play a critical role in pathogenesis. The NET formation is triggered by danger signals and can involve citrullination [115]. Of note, ACPA-targeted citrullinated peptides are present in NETs [116, 117]. On the other hand, researchers discovered that NET formation is increased by ACPA depending on its concentration [116].

Neutrophils interact with various immune cells to induce inflammation. B cells, stimulated by citrullinated proteins in the presence of ACPAs, activate neutrophil migration by producing CXCL8 [118]. RA synovial fibroblasts internalize NET peptides and present them to T cells, promoting T-cell immune responses [119]. NET formation primes macrophage activation, leading to cytokine production that activates Th17 cells, increasing inflammation [120]. Interestingly, neutrophils were found to express MHC class II, promoting antigen presentation and CD4⁺ T cell growth, suggesting a role in regulating adaptive immune responses [105, 121].

Conventional therapies for RA include anti-TNF- α and anti-IL-6 receptor treatments [122]. These therapies reduce inflammation and inhibit neutrophil-related functions.

Targeting molecules derived from neutrophils is an emerging approach. Modulating CCR2 [123] and CXCR1/CXCR2 chemokine receptors [124, 125] can inhibit neutrophil migration and function. PAD4 inhibitors have shown some promise in preventing RA severity [126].

Conclusions

RA is a complex disease with a multifaceted etiology, manifesting features of Type II, Type III, and Type IV hypersensitivity reactions simultaneously, engaging both adaptive and innate immune cells. Although the precise cause remains elusive, a combination of genetic and environmental factors is suspected to be involved. Unfortunately, RA cannot be cured yet, but ongoing research strives to formulate treatments targeting the autoimmune aspects of the disease. Despite a myriad of proposed approaches to support individuals grappling with RA, these solutions may fall short of fully addressing the intricacies of the condition. The pursuit of innovative strategies, such as comprehensive combination therapies addressing various facets of RA, holds the potential to significantly improve the prospects of conquering this challenging ailment.

Abbreviations

Tregs: regulatory T cells

ACPAs: anti-(cyclic) citrullinated protein antibodies CR: complement receptor FcRs: Fc receptors GM-CSF: granulocyte-macrophage colony-stimulating factor IFN: interferon Ig: immunoglobulin NETs: neutrophil extracellular traps PAD: peptidyl arginine deiminase RA: rheumatoid arthritis RF: rheumatoid factor ROS: reactive oxygen species Th1: T helper 1

Declarations

Author contributions

JH, SH, JRK, JK, and YL equally contributed to: Investigation, Writing—original draft. JHB: Conceptualization, Validation, Writing—review & editing, Supervision. All authors read and approved the submitted version.

Conflicts of interest

The authors declare that they have no conflicts of interest.

Ethical approval

Not applicable.

Consent to participate

Not applicable.

Consent to publication

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Availability of data and materials

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References

- 1. Bullock J, Rizvi SAA, Saleh AM, Ahmed SS, Do DP, Ansari RA, et al. Rheumatoid Arthritis: A Brief Overview of the Treatment. Med Princ Pract. 2018;27:501–7.
- 2. Cojocaru M, Cojocaru IM, Silosi I, Vrabie CD, Tanasescu R. Extra-articular Manifestations in Rheumatoid Arthritis. Maedica (Bucur). 2010;5:286–91.
- 3. Capellino S, Cosentino M, Wolff C, Schmidt M, Grifka J, Straub RH. Catecholamine-producing cells in the synovial tissue during arthritis: modulation of sympathetic neurotransmitters as new therapeutic target. Ann Rheum Dis. 2010;69:1853–60.
- 4. Martin GV, Kanaan SB, Hemon MF, Azzouz DF, El Haddad M, Balandraud N, et al. Mosaicism of XX and XXY cells accounts for high copy number of *Toll like Receptor 7 and 8* genes in peripheral blood of men with Rheumatoid Arthritis. Sci Rep. 2019;9:12880.
- 5. Li Y, Shen Y, Hohensinner P, Ju J, Wen Z, Goodman SB, et al. Deficient Activity of the Nuclease MRE11A Induces T Cell Aging and Promotes Arthritogenic Effector Functions in Patients with Rheumatoid Arthritis. Immunity. 2016;45:903–16.
- 6. Fujii H, Shao L, Colmegna I, Goronzy JJ, Weyand CM. Telomerase insufficiency in rheumatoid arthritis. Proc Natl Acad Sci U S A. 2009;106:4360–5.
- Chen YC, Huang CM, Liu TY, Wu N, Chan CJ, Shih PY, et al. Effects of Human Leukocyte Antigen DRB1 Genetic Polymorphism on Anti-Cyclic Citrullinated Peptide (ANTI-CCP) and Rheumatoid Factor (RF) Expression in Rheumatoid Arthritis (RA) Patients. Int J Mol Sci. 2023;24:12036.
- 8. Puszczewicz M, Iwaszkiewicz C. Role of anti-citrullinated protein antibodies in diagnosis and prognosis of rheumatoid arthritis. Arch Med Sci. 2011;7:189–94.

- 9. Jonsson AH. Synovial Tissue Insights into Heterogeneity of Rheumatoid Arthritis. Curr Rheumatol Rep. 2024;26:81–8.
- 10. Kraan MC, Reece RJ, Smeets TJ, Veale DJ, Emery P, Tak PP. Comparison of synovial tissues from the knee joints and the small joints of rheumatoid arthritis patients: Implications for pathogenesis and evaluation of treatment. Arthritis Rheum. 2002;46:2034–8.
- 11. Bottini N, Firestein GS. Duality of fibroblast-like synoviocytes in RA: passive responders and imprinted aggressors. Nat Rev Rheumatol. 2013;9:24–33.
- 12. Konttinen YT, Ceponis A, Takagi M, Ainola M, Sorsa T, Sutinen M, et al. New collagenolytic enzymes/ cascade identified at the pannus-hard tissue junction in rheumatoid arthritis: destruction from above. Matrix Biol. 1998;17:585–601.
- 13. Mukherjee A, Das B. The role of inflammatory mediators and matrix metalloproteinases (MMPs) in the progression of osteoarthritis. Biomater Biosyst. 2024;13:100090.
- 14. Axmann R, Böhm C, Krönke G, Zwerina J, Smolen J, Schett G. Inhibition of interleukin-6 receptor directly blocks osteoclast formation in vitro and in vivo. Arthritis Rheum. 2009;60:2747–56.
- 15. Honorati MC, Neri S, Cattini L, Facchini A. Interleukin-17, a regulator of angiogenic factor release by synovial fibroblasts. Osteoarthritis Cartilage. 2006;14:345–52.
- 16. Alivernini S, MacDonald L, Elmesmari A, Finlay S, Tolusso B, Gigante MR, et al. Distinct synovial tissue macrophage subsets regulate inflammation and remission in rheumatoid arthritis. Nat Med. 2020;26:1295–306.
- Yap HY, Tee SZ, Wong MM, Chow SK, Peh SC, Teow SY. Pathogenic Role of Immune Cells in Rheumatoid Arthritis: Implications in Clinical Treatment and Biomarker Development. Cells. 2018;7: 161.
- 18. Choy E. Understanding the dynamics: pathways involved in the pathogenesis of rheumatoid arthritis. Rheumatology (Oxford). 2012;51:v3–11.
- 19. Guo Q, Wang Y, Xu D, Nossent J, Pavlos NJ, Xu J. Rheumatoid arthritis: pathological mechanisms and modern pharmacologic therapies. Bone Res. 2018;6:15.
- 20. Chemin K, Gerstner C, Malmström V. Effector Functions of CD4+ T Cells at the Site of Local Autoimmune Inflammation—Lessons From Rheumatoid Arthritis. Front Immunol. 2019;10:353.
- 21. James EA, Rieck M, Pieper J, Gebe JA, Yue BB, Tatum M, et al. Citrulline-specific Th1 cells are increased in rheumatoid arthritis and their frequency is influenced by disease duration and therapy. Arthritis Rheumatol. 2014;66:1712–22.
- 22. Kotake S, Yago T, Kobashigawa T, Nanke Y. The Plasticity of Th17 Cells in the Pathogenesis of Rheumatoid Arthritis. J Clin Med. 2017;6:67.
- 23. Rao DA, Gurish MF, Marshall JL, Slowikowski K, Fonseka CY, Liu Y, et al. Pathologically expanded peripheral T helper cell subset drives B cells in rheumatoid arthritis. Nature. 2017;542:110–4.
- 24. Galindo-Izquierdo M, Pablos Alvarez JL. Complement as a Therapeutic Target in Systemic Autoimmune Diseases. Cells. 2021;10:148.
- 25. Holers VM, Banda NK. Complement in the Initiation and Evolution of Rheumatoid Arthritis. Front Immunol. 2018;9:1057.
- 26. Rubtsov AV, Rubtsova K, Fischer A, Meehan RT, Gillis JZ, Kappler JW, et al. Toll-like receptor 7 (TLR7)-driven accumulation of a novel CD11c⁺ B-cell population is important for the development of autoimmunity. Blood. 2011;118:1305–15.
- 27. Wu CY, Yang HY, Luo SF, Lai JH. From Rheumatoid Factor to Anti-Citrullinated Protein Antibodies and Anti-Carbamylated Protein Antibodies for Diagnosis and Prognosis Prediction in Patients with Rheumatoid Arthritis. Int J Mol Sci. 2021;22:686.
- Trela M, Perera S, Sheeran T, Rylance P, Nelson PN, Attridge K. Citrullination facilitates crossreactivity of rheumatoid factor with non-IgG1 Fc epitopes in rheumatoid arthritis. Sci Rep. 2019;9: 12068.

- 29. Nielen MM, van Schaardenburg D, Reesink HW, van de Stadt RJ, van der Horst-Bruinsma IE, de Koning MH, et al. Specific autoantibodies precede the symptoms of rheumatoid arthritis: a study of serial measurements in blood donors. Arthritis Rheum. 2004;50:380–6.
- 30. Agrawal S, Misra R, Aggarwal A. Autoantibodies in rheumatoid arthritis: association with severity of disease in established RA. Clin Rheumatol. 2007;26:201–4.
- 31. Kroot EJ, de Jong BA, van Leeuwen MA, Swinkels H, van den Hoogen FH, van't Hof M, et al. The prognostic value of anti-cyclic citrullinated peptide antibody in patients with recent-onset rheumatoid arthritis. Arthritis Rheum. 2000;43:1831–5.
- 32. Nakagawa K, Sakiyama H, Tsuchida T, Yamaguchi K, Toyoguchi T, Masuda R, et al. Complement C1s activation in degenerating articular cartilage of rheumatoid arthritis patients: immunohistochemical studies with an active form specific antibody. Ann Rheum Dis. 1999;58:175–81.
- 33. Wu XY, Li KT, Yang HX, Yang B, Lu X, Zhao LD, et al. Complement C1q synergizes with PTX3 in promoting NLRP3 inflammasome over-activation and pyroptosis in rheumatoid arthritis. J Autoimmun. 2020;106:102336.
- 34. Monach PA, Verschoor A, Jacobs JP, Carroll MC, Wagers AJ, Benoist C, et al. Circulating C3 is necessary and sufficient for induction of autoantibody-mediated arthritis in a mouse model. Arthritis Rheum. 2007;56:2968–74.
- 35. Vergunst CE, Gerlag DM, Dinant H, Schulz L, Vinkenoog M, Smeets TJ, et al. Blocking the receptor for C5a in patients with rheumatoid arthritis does not reduce synovial inflammation. Rheumatology (Oxford). 2007;46:1773–8.
- 36. Hietala MA, Jonsson IM, Tarkowski A, Kleinau S, Pekna M. Complement deficiency ameliorates collagen-induced arthritis in mice. J Immunol. 2002;169:454–9.
- 37. Barrington R, Zhang M, Fischer M, Carroll MC. The role of complement in inflammation and adaptive immunity. Immunol Rev. 2001;180:5–15.
- 38. Avouac J, Allanore Y. Cardiovascular risk in rheumatoid arthritis: effects of anti-TNF drugs. Expert Opin Pharmacother. 2008;9:1121–8.
- 39. Song H, Qiao F, Atkinson C, Holers VM, Tomlinson S. A complement C3 inhibitor specifically targeted to sites of complement activation effectively ameliorates collagen-induced arthritis in DBA/1J mice. J Immunol. 2007;179:7860–7.
- 40. Takai T. Roles of Fc receptors in autoimmunity. Nat Rev Immunol. 2002;2:580–92.
- 41. Uzzaman A, Cho SH. Chapter 28: Classification of hypersensitivity reactions. Allergy Asthma Proc. 2012;33:96–9.
- 42. el Bannoudi H, Ioan-Facsinay A, Toes RE. Bridging autoantibodies and arthritis: the role of Fc receptors. Curr Top Microbiol Immunol. 2014;382:303–19.
- 43. Nicolò A, Amendt T, El Ayoubi O, Young M, Finzel S, Senel M, et al. Rheumatoid factor IgM autoantibodies control IgG homeostasis. Front Immunol. 2022;13:1016263. Erratum in: Front Immunol. 2022;13:1123117.
- 44. Magnusson SE, Wennerberg E, Matt P, Lindqvist U, Kleinau S. Dysregulated Fc receptor function in active rheumatoid arthritis. Immunol Lett. 2014;162:200–6.
- 45. Wu Y, Pan W, Hu X, Zhang A, Wei W. The prospects for targeting FcR as a novel therapeutic strategy in rheumatoid arthritis. Biochem Pharmacol. 2021;183:114360.
- 46. Schellekens GA, Visser H, de Jong BA, van den Hoogen FH, Hazes JM, Breedveld FC, et al. The diagnostic properties of rheumatoid arthritis antibodies recognizing a cyclic citrullinated peptide. Arthritis Rheum. 2000;43:155–63.
- 47. Araujo GR, Vaz ER, Fujimura PT, Fonseca JE, de Lima LM, Canhão H, et al. Improved serological detection of rheumatoid arthritis: a highly antigenic mimotope of carbonic anhydrase III selected in a murine model by phage display. Arthritis Res Ther. 2015;17:168.

- 48. Zuo Y, Deng GM. Fc Gamma Receptors as Regulators of Bone Destruction in Inflammatory Arthritis. Front Immunol. 2021;12:688201.
- 49. Warrington R, Watson W, Kim HL, Antonetti FR. An introduction to immunology and immunopathology. Allergy Asthma Clin Immunol. 2011;7:S1.
- 50. Arango Duque G, Descoteaux A. Macrophage cytokines: involvement in immunity and infectious diseases. Front Immunol. 2014;5:491.
- 51. Dolhain RJ, van der Heiden AN, ter Haar NT, Breedveld FC, Miltenburg AM. Shift toward T lymphocytes with a T helper 1 cytokine-secretion profile in the joints of patients with rheumatoid arthritis. Arthritis Rheum. 1996;39:1961–9.
- 52. Schulze-Koops H, Kalden JR. The balance of Th1/Th2 cytokines in rheumatoid arthritis. Best Pract Res Clin Rheumatol. 2001;15:677–91.
- 53. Patel DD, Zachariah JP, Whichard LP. CXCR3 and CCR5 ligands in rheumatoid arthritis synovium. Clin Immunol. 2001;98:39–45.
- 54. Maruotti N, Cantatore FP, Crivellato E, Vacca A, Ribatti D. Macrophages in rheumatoid arthritis. Histol Histopathol. 2007;22:581–6.
- 55. Kurowska-Stolarska M, Alivernini S. Synovial tissue macrophages: friend or foe? RMD Open. 2017;3: e000527.
- 56. Borst J, Ahrends T, Bąbała N, Melief CJM, Kastenmüller W. CD4⁺ T cell help in cancer immunology and immunotherapy. Nat Rev Immunol. 2018;18:635–47.
- Chapuy-Regaud S, Nogueira L, Clavel C, Sebbag M, Vincent C, Serre G. IgG subclass distribution of the rheumatoid arthritis-specific autoantibodies to citrullinated fibrin. Clin Exp Immunol. 2005;139: 542–50.
- 58. Cho BA, Sim JH, Park JA, Kim HW, Yoo WH, Lee SH, et al. Characterization of effector memory CD8⁺ T cells in the synovial fluid of rheumatoid arthritis. J Clin Immunol. 2012;32:709–20.
- 59. Harrington LE, Hatton RD, Mangan PR, Turner H, Murphy TL, Murphy KM, et al. Interleukin 17producing CD4⁺ effector T cells develop via a lineage distinct from the T helper type 1 and 2 lineages. Nat Immunol. 2005;6:1123–32.
- 60. Ip B, Cilfone NA, Belkina AC, DeFuria J, Jagannathan-Bogdan M, Zhu M, et al. Th17 cytokines differentiate obesity from obesity-associated type 2 diabetes and promote TNFα production. Obesity (Silver Spring). 2016;24:102–12.
- 61. Codarri L, Gyülvészi G, Tosevski V, Hesske L, Fontana A, Magnenat L, et al. RORγt drives production of the cytokine GM-CSF in helper T cells, which is essential for the effector phase of autoimmune neuroinflammation. Nat Immunol. 2011;12:560–7.
- 62. Stockinger B, Omenetti S. The dichotomous nature of T helper 17 cells. Nat Rev Immunol. 2017;17: 535–44.
- Hu D, Notarbartolo S, Croonenborghs T, Patel B, Cialic R, Yang TH, et al. Transcriptional signature of human pro-inflammatory T_H17 cells identifies reduced *IL10* gene expression in multiple sclerosis. Nat Commun. 2017;8:1600.
- 64. Kotake S, Nanke Y, Yago T, Kawamoto M, Kobashigawa T, Yamanaka H. Ratio of Circulating IFNγ⁺ "Th17 Cells" in Memory Th Cells Is Inversely Correlated with the Titer of Anti-CCP Antibodies in Early-Onset Rheumatoid Arthritis Patients Based on Flow Cytometry Methods of the Human Immunology Project. Biomed Res Int. 2016;2016:9694289.
- 65. Raza K, Falciani F, Curnow SJ, Ross EJ, Lee CY, Akbar AN, et al. Early rheumatoid arthritis is characterized by a distinct and transient synovial fluid cytokine profile of T cell and stromal cell origin. Arthritis Res Ther. 2005;7:R784–95.
- 66. Zizzo G, De Santis M, Bosello SL, Fedele AL, Peluso G, Gremese E, et al. Synovial fluid-derived T helper
 17 cells correlate with inflammatory activity in arthritis, irrespectively of diagnosis. Clin Immunol.
 2011;138:107–16.

- 67. Rasmussen TK, Andersen T, Hvid M, Hetland ML, Hørslev-Petersen K, Stengaard-Pedersen K, et al. Increased interleukin 21 (IL-21) and IL-23 are associated with increased disease activity and with radiographic status in patients with early rheumatoid arthritis. J Rheumatol. 2010;37:2014–20.
- 68. Neumann E, Lefèvre S, Zimmermann B, Gay S, Müller-Ladner U. Rheumatoid arthritis progression mediated by activated synovial fibroblasts. Trends Mol Med. 2010;16:458–68. Erratum in: Trends Mol Med. 2011;17:118.
- 69. Hirota K, Hashimoto M, Ito Y, Matsuura M, Ito H, Tanaka M, et al. Autoimmune Th17 Cells Induced Synovial Stromal and Innate Lymphoid Cell Secretion of the Cytokine GM-CSF to Initiate and Augment Autoimmune Arthritis. Immunity. 2018;48:1220–32.e5.
- 70. Koenders MI, Marijnissen RJ, Devesa I, Lubberts E, Joosten LA, Roth J, et al. Tumor necrosis factor-interleukin-17 interplay induces S100A8, interleukin-1β, and matrix metalloproteinases, and drives irreversible cartilage destruction in murine arthritis: rationale for combination treatment during arthritis. Arthritis Rheum. 2011;63:2329–39.
- 71. Pickens SR, Volin MV, Mandelin AM 2nd, Kolls JK, Pope RM, Shahrara S. IL-17 contributes to angiogenesis in rheumatoid arthritis. J Immunol. 2010;184:3233–41.
- 72. van Hamburg JP, Asmawidjaja PS, Davelaar N, Mus AM, Colin EM, Hazes JM, et al. Th17 cells, but not Th1 cells, from patients with early rheumatoid arthritis are potent inducers of matrix metalloproteinases and proinflammatory cytokines upon synovial fibroblast interaction, including autocrine interleukin-17A production. Arthritis Rheum. 2011;63:73–83.
- 73. Kim HJ, Kang WY, Seong SJ, Kim SY, Lim MS, Yoon YR. Follistatin-like 1 promotes osteoclast formation via RANKL-mediated NF-κB activation and M-CSF-induced precursor proliferation. Cell Signal. 2016;28:1137–44.
- 74. Löfvall H, Newbould H, Karsdal MA, Dziegiel MH, Richter J, Henriksen K, et al. Osteoclasts degrade bone and cartilage knee joint compartments through different resorption processes. Arthritis Res Ther. 2018;20:67.
- 75. Ke D, Fu X, Xue Y, Wu H, Zhang Y, Chen X, et al. IL-17A regulates the autophagic activity of osteoclast precursors through RANKL-JNK1 signaling during osteoclastogenesis in vitro. Biochem Biophys Res Commun. 2018;497:890–6.
- 76. Tao JH, Cheng M, Tang JP, Liu Q, Pan F, Li XP. Foxp3, Regulatory T Cell, and Autoimmune Diseases. Inflammation. 2017;40:328–39.
- 77. Chen X, Oppenheim JJ. Resolving the identity myth: key markers of functional CD4⁺FoxP3⁺ regulatory T cells. Int Immunopharmacol. 2011;11:1489–96.
- 78. Langdon K, Haleagrahara N. Regulatory T-cell dynamics with abatacept treatment in rheumatoid arthritis. Int Rev Immunol. 2018;37:206–14.
- 79. Miyara M, Yoshioka Y, Kitoh A, Shima T, Wing K, Niwa A, et al. Functional delineation and differentiation dynamics of human CD4⁺ T cells expressing the FoxP3 transcription factor. Immunity. 2009;30:899–911.
- 80. Polansky JK, Kretschmer K, Freyer J, Floess S, Garbe A, Baron U, et al. DNA methylation controls *Foxp3* gene expression. Eur J Immunol. 2008;38:1654–63.
- 81. Herrath J, Müller M, Amoudruz P, Janson P, Michaëlsson J, Larsson PT, et al. The inflammatory milieu in the rheumatic joint reduces regulatory T-cell function. Eur J Immunol. 2011;41:2279–90.
- Zanin-Zhorov A, Ding Y, Kumari S, Attur M, Hippen KL, Brown M, et al. Protein Kinase C-θ Mediates Negative Feedback on Regulatory T Cell Function. Science. 2010;328:372–6. Erratum in: Science. 2010;328:974.
- 83. Zaragoza B, Chen X, Oppenheim JJ, Baeyens A, Gregoire S, Chader D, et al. Suppressive activity of human regulatory T cells is maintained in the presence of TNF. Nat Med. 2016;22:16–7.
- 84. Jiang Q, Yang G, Liu Q, Wang S, Cui D. Function and Role of Regulatory T Cells in Rheumatoid Arthritis. Front Immunol. 2021;12:626193.

- 85. Curotto de Lafaille MA, Lafaille JJ. Natural and adaptive foxp3⁺ regulatory T cells: more of the same or a division of labor? Immunity. 2009;30:626–35.
- 86. Bending D, Giannakopoulou E, Lom H, Wedderburn LR. Synovial Regulatory T Cells Occupy a Discrete TCR Niche in Human Arthritis and Require Local Signals To Stabilize FOXP3 Protein Expression. J Immunol. 2015;195:5616–24.
- 87. Yan S, Golumba-Nagy V, Kotschenreuther K, Thiele J, Refaian N, Shuya D, et al. Membrane-bound IL-6R is upregulated on Th17 cells and inhibits Treg cell migration by regulating post-translational modification of VASP in autoimmune arthritis. Cell Mol Life Sci. 2021;79:3.
- 88. Lykhopiy V, Malviya V, Humblet-Baron S, Schlenner SM. IL-2 immunotherapy for targeting regulatory T cells in autoimmunity. Genes Immun. 2023;24:248–62.
- 89. Zhang R, Zhao Y, Chen X, Zhuang Z, Li X, Shen E. Low-dose IL-2 therapy in autoimmune diseases: An update review. Int Rev Immunol. 2024;43:113–37.
- 90. Ye C, Brand D, Zheng SG. Targeting IL-2: an unexpected effect in treating immunological diseases. Signal Transduct Target Ther. 2018;3:2.
- 91. Klatzmann D, Abbas AK. The promise of low-dose interleukin-2 therapy for autoimmune and inflammatory diseases. Nat Rev Immunol. 2015;15:283–94.
- 92. Kinne RW, Bräuer R, Stuhlmüller B, Palombo-Kinne E, Burmester GR. Macrophages in rheumatoid arthritis. Arthritis Res. 2000;2:189–202.
- 93. Knab K, Chambers D, Krönke G. Synovial Macrophage and Fibroblast Heterogeneity in Joint Homeostasis and Inflammation. Front Med (Lausanne). 2022;9:862161.
- 94. Tateiwa D, Yoshikawa H, Kaito T. Cartilage and Bone Destruction in Arthritis: Pathogenesis and Treatment Strategy: A Literature Review. Cells. 2019;8:818.
- 95. Kurowska-Stolarska M, Alivernini S. Synovial tissue macrophages in joint homeostasis, rheumatoid arthritis and disease remission. Nat Rev Rheumatol. 2022;18:384–97.
- 96. Misharin AV, Cuda CM, Saber R, Turner JD, Gierut AK, Haines GK 3rd, et al. Nonclassical Ly6C⁻ monocytes drive the development of inflammatory arthritis in mice. Cell Rep. 2014;9:591–604.
- 97. González-Sánchez HM, Baek JH, Weinmann-Menke J, Ajay AK, Charles JF, Noda M, et al. IL-34 and protein-tyrosine phosphatase receptor type-zeta-dependent mechanisms limit arthritis in mice. Lab Invest. 2022;102:846–58.
- 98. Park HE, Oh H, Baek JH. Interleukin-34-regulated T-cell responses in rheumatoid arthritis. Front Med (Lausanne). 2022;9:1078350.
- 99. Baek JH. Interleukin-34: regulator of T lymphocytes in rheumatoid arthritis. Biomed J Sci Tech Res. 2019;20:15062–6.
- 100. Udalova IA, Mantovani A, Feldmann M. Macrophage heterogeneity in the context of rheumatoid arthritis. Nat Rev Rheumatol. 2016;12:472–85.
- 101. Mezghiche I, Yahia-Cherbal H, Rogge L, Bianchi E. Interleukin 23 receptor: Expression and regulation in immune cells. Eur J Immunol. 2024;54:e2250348.
- 102. Gu Q, Yang H, Shi Q. Macrophages and bone inflammation. J Orthop Translat. 2017;10:86–93.
- Schnell A, Littman DR, Kuchroo VK. T_H17 cell heterogeneity and its role in tissue inflammation. Nat Immunol. 2023;24:19–29.
- 104. Hashimoto M, Hirota K, Yoshitomi H, Maeda S, Teradaira S, Akizuki S, et al. Complement drives Th17 cell differentiation and triggers autoimmune arthritis. J Exp Med. 2010;207:1135–43.
- 105. Culshaw S, Millington OR, Brewer JM, McInnes IB. Murine neutrophils present Class II restricted antigen. Immunol Lett. 2008;118:49–54.
- 106. Ottonello L, Cutolo M, Frumento G, Arduino N, Bertolotto M, Mancini M, et al. Synovial fluid from patients with rheumatoid arthritis inhibits neutrophil apoptosis: role of adenosine and proinflammatory cytokines. Rheumatology (Oxford). 2002;41:1249–60.

- 107. Parsonage G, Filer A, Bik M, Hardie D, Lax S, Howlett K, et al. Prolonged, granulocyte-macrophage colony-stimulating factor-dependent, neutrophil survival following rheumatoid synovial fibroblast activation by IL-17 and TNFalpha. Arthritis Res Ther. 2008;10:R47.
- 108. McNamee JP, Bellier PV, Kutzner BC, Wilkins RC. Effect of pro-inflammatory cytokines on spontaneous apoptosis in leukocyte sub-sets within a whole blood culture. Cytokine. 2005;31:161–7.
- 109. Christenson K, Björkman L, Karlsson A, Bylund J. Regulation of neutrophil apoptosis differs after in vivo transmigration to skin chambers and synovial fluid: a role for inflammasome-dependent interleukin-1β release. J Innate Immun. 2013;5:377–88.
- 110. Wright HL, Moots RJ, Edwards SW. The multifactorial role of neutrophils in rheumatoid arthritis. Nat Rev Rheumatol. 2014;10:593–601.
- 111. Trzybulska D, Olewicz-Gawlik A, Graniczna K, Kisiel K, Moskal M, Cieślak D, et al. Quantitative analysis of elastase and cathepsin G mRNA levels in peripheral blood CD14⁺ cells from patients with rheumatoid arthritis. Cell Immunol. 2014;292:40–4. Erratum in: Cell Immunol. 2015;296:155.
- 112. Velvart M, Fehr K. Degradation in vivo of articular cartilage in rheumatoid arthritis and juvenile chronic arthritis by cathepsin G and elastase from polymorphonuclear leukocytes. Rheumatol Int. 1987;7:195–202.
- Jones JE, Causey CP, Knuckley B, Slack-Noyes JL, Thompson PR. Protein arginine deiminase 4 (PAD4): Current understanding and future therapeutic potential. Curr Opin Drug Discov Devel. 2009;12: 616–27.
- 114. Metzler KD, Goosmann C, Lubojemska A, Zychlinsky A, Papayannopoulos V. A myeloperoxidasecontaining complex regulates neutrophil elastase release and actin dynamics during NETosis. Cell Rep. 2014;8:883–96.
- 115. Romero V, Fert-Bober J, Nigrovic PA, Darrah E, Haque UJ, Lee DM, et al. Immune-mediated poreforming pathways induce cellular hypercitrullination and generate citrullinated autoantigens in rheumatoid arthritis. Sci Transl Med. 2013;5:209ra150.
- 116. Wu S, Peng W, Liang X, Wang W. Anti-citrullinated protein antibodies are associated with neutrophil extracellular trap formation in rheumatoid arthritis. J Clin Lab Anal. 2021;35:e23662.
- 117. Khandpur R, Carmona-Rivera C, Vivekanandan-Giri A, Gizinski A, Yalavarthi S, Knight JS, et al. NETs are a source of citrullinated autoantigens and stimulate inflammatory responses in rheumatoid arthritis. Sci Transl Med. 2013;5:178ra40.
- 118. Kristyanto H, Blomberg NJ, Slot LM, van der Voort EIH, Kerkman PF, Bakker A, et al. Persistently activated, proliferative memory autoreactive B cells promote inflammation in rheumatoid arthritis. Sci Transl Med. 2020;12:eaaz5327.
- 119. Carmona-Rivera C, Carlucci PM, Moore E, Lingampalli N, Uchtenhagen H, James E, et al. Synovial fibroblast-neutrophil interactions promote pathogenic adaptive immunity in rheumatoid arthritis. Sci Immunol. 2017;2:eaag3358. Erratum in: Sci Immunol. 2020;5.
- 120. Nakazawa D, Shida H, Kusunoki Y, Miyoshi A, Nishio S, Tomaru U, et al. The responses of macrophages in interaction with neutrophils that undergo NETosis. J Autoimmun. 2016;67:19–28.
- 121. Cross A, Bucknall RC, Cassatella MA, Edwards SW, Moots RJ. Synovial fluid neutrophils transcribe and express class II major histocompatibility complex molecules in rheumatoid arthritis. Arthritis Rheum. 2003;48:2796–806.
- 122. Taylor PC, Peters AM, Paleolog E, Chapman PT, Elliott MJ, McCloskey R, et al. Reduction of chemokine levels and leukocyte traffic to joints by tumor necrosis factor α blockade in patients with rheumatoid arthritis. Arthritis Rheum. 2000;43:38–47.
- 123. Talbot J, Bianchini FJ, Nascimento DC, Oliveira RD, Souto FO, Pinto LG, et al. CCR2 Expression in Neutrophils Plays a Critical Role in Their Migration Into the Joints in Rheumatoid Arthritis. Arthritis Rheumatol. 2015;67:1751–9.

- 124. Grespan R, Fukada SY, Lemos HP, Vieira SM, Napimoga MH, Teixeira MM, et al. CXCR2-specific chemokines mediate leukotriene B4-dependent recruitment of neutrophils to inflamed joints in mice with antigen-induced arthritis. Arthritis Rheum. 2008;58:2030–40.
- 125. Min SH, Wang Y, Gonsiorek W, Anilkumar G, Kozlowski J, Lundell D, et al. Pharmacological targeting reveals distinct roles for CXCR2/CXCR1 and CCR2 in a mouse model of arthritis. Biochem Biophys Res Commun. 2010;391:1080–6.
- 126. Willis VC, Banda NK, Cordova KN, Chandra PE, Robinson WH, Cooper DC, et al. Protein arginine deiminase 4 inhibition is sufficient for the amelioration of collagen-induced arthritis. Clin Exp Immunol. 2017;188:263–74.