



Roles of calcium signaling in cancer metastasis to bone

Tianying Xie^{1†}, Sitong Chen^{1†}, Jiang Hao^{2†}, Pengfei Wu³, Xuelian Gu¹, Haifeng Wei^{2*}, Zhenxi Li^{1,2*}, Jianru Xiao^{1,2*}

¹School of Health Science and Engineering, University of Shanghai for Science and Technology, Shanghai 200093, China

²Department of Orthopedic Oncology, Shanghai Changzheng Hospital, Shanghai 200003, China

³Center for Medical Genetics, School of Life Sciences, Central South University, Changsha 410008, Hunan, China

[†]The authors contributed equally to the work.

***Correspondence:** Haifeng Wei, Department of Orthopedic Oncology, Shanghai Changzheng Hospital, Shanghai 200003, China. weihfspine@163.com; Zhenxi Li, School of Health Science and Engineering, University of Shanghai for Science and Technology, Shanghai 200093, China. Department of Orthopedic Oncology, Shanghai Changzheng Hospital, Shanghai 200003, China. zhenxili.ecnu@gmail.com; Jianru Xiao, School of Health Science and Engineering, University of Shanghai for Science and Technology, Shanghai 200093, China. Department of Orthopedic Oncology, Shanghai Changzheng Hospital, Shanghai 200003, China. jianruxiao83@163.com

Academic Editor: Zui Pan, The University of Texas at Arlington, USA

Received: February 7, 2022 **Accepted:** May 16, 2022 **Published:** August 31, 2022

Cite this article: Xie T, Chen S, Hao J, Wu P, Gu X, Wei H, et al. Roles of calcium signaling in cancer metastasis to bone. *Explor Target Antitumor Ther.* 2022;3:445–62. <https://doi.org/10.37349/etat.2022.00094>

Abstract

Bone metastasis is a frequent complication for cancers and an important reason for the mortality in cancer patients. After surviving in bone, cancer cells can cause severe pain, life-threatening hypercalcemia, pathologic fractures, spinal cord compression, and even death. However, the underlying mechanisms of bone metastasis were not clear. The role of calcium (Ca^{2+}) in cancer cell proliferation, migration, and invasion has been well established. Interestingly, emerging evidence indicates that Ca^{2+} signaling played a key role in bone metastasis, for it not only promotes cancer progression but also mediates osteoclasts and osteoblasts differentiation. Therefore, Ca^{2+} signaling has emerged as a novel therapeutical target for cancer bone metastasis treatments. Here, the role of Ca^{2+} channels and Ca^{2+} -binding proteins including calmodulin and Ca^{2+} -sensing receptor in bone metastasis, and the perspective of anti-cancer bone metastasis therapeutics via targeting the Ca^{2+} signaling pathway are summarized.

Keywords

Bone metastasis, calcium, calcium channels, calcium-sensing receptor, calmodulin

Introduction

Bone metastasis is a process in which tumor cells escape from the primary tumor site and colonize the bone microenvironment [1], bringing about a plethora of complications, such as bone pain, pathological fractures, and life-threatening hypercalcemia. It has generally been characterized as osteolytic or osteoblastic (osteosclerotic), leading to bone destruction and new bone formation, respectively [2].

© The Author(s) 2022. This is an Open Access article licensed under a Creative Commons Attribution 4.0 International License (<https://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, sharing, adaptation, distribution and reproduction in any medium or format, for any purpose, even commercially, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made.



There's a good reason for cancer cells' predilection for bone. A seed-and-soil hypothesis was first proposed by Paget [3] in 1889, and shreds of evidence have been found over the years to support this hypothesis. Red marrow areas have a high blood flow, providing a nutritious environment [4]. More importantly, the bone microenvironment is rich in growth factors, including transforming growth factor β (TGF β), insulin-like growth factor I (IGFI) and IGFII, fibroblast growth factors (FGFs), and calcium (Ca²⁺) [5]. These factors are released into the bone microenvironment and/or activated during bone resorption. Many of these growth factors can stimulate the proliferation of cancer cells in bone and induce the production and release of bone-resorbing factors from tumor cells [6]. For example, the secretion of receptor activator of nuclear factor-kappa B ligand (RANKL), located on the plasma membrane of osteoblasts, by activated T cells, binds the receptor activator of nuclear factor-kappa B (RANK) receptor on osteoclast precursors and leads to osteoclast formation [6]. Furthermore, tumor cells also secrete RANKL in a high Ca²⁺ environment and modulate osteoclastic differentiation [7]. The importance of RANKL in bone resorption and Ca²⁺ metabolism has been demonstrated clearly with the use of RANK knockout mice [8]. With findings in osteolytic metastatic lesions suggesting that the remodeling of bone is induced by osteoclasts instead of tumor cells [2], RANK is determined to be essential for osteolytic metastases [9].

It is well established that Ca²⁺ signaling plays a pivotal role in tumor bone metastases with abundant research. Pathway enrichment analysis highlighted that the Ca²⁺ signaling pathway is a potential key regulator for breast cancer bone metastasis [10]. Elevated levels of intracellular Ca²⁺ in prostate cancer cells induce proliferation, angiogenesis, epithelial to mesenchymal transition (EMT), migration, and bone colonization [11]. Ca²⁺ signaling facilitates malignant cells' bone colonization via a variety of mechanisms, interacting with cancer cells, osteoclasts, osteoblasts, and osteogenic niches [12]. However, the mechanism is not well understood. In this review, an insight into the Ca²⁺ signaling in cancer metastasis to bone is provided to our audiences.

Ca²⁺

Ca²⁺, a ubiquitous intracellular messenger, regulates diverse cellular processes, such as gene transcription, apoptosis, autophagy, and cell proliferation. However, cellular Ca²⁺ signaling proteomes, such as Ca²⁺ channels, and Ca²⁺-binding proteins including calmodulin (CaM) and Ca²⁺-sensing receptor (CaSR), are tissue-specific and produce distinct Ca²⁺ signals suitable for tissue physiology [13]. Cytosolic Ca²⁺ signals practically participate in every aspect of cellular life, and rigorous regulation of Ca²⁺ homeostasis is important for preventing dysfunctions that lead to pathological changes [14]. In a pathological environment, remodeling of Ca²⁺ flux contributes to processes important for cancer progressions, such as uncontrolled proliferation, invasiveness of tumor cells, and the development of resistance to cancer therapies [15]. Increases in intracellular Ca²⁺ concentration are involved in cell migration, and impaired Ca²⁺ signaling is important in the metastatic behavior of tumor cells [16]. CaM1–4 remarkably emphasizes the importance of Ca²⁺ signaling by extending Ca²⁺ ions' signals. Ca²⁺/CaM binding activates numerous proteins that contain CaM recruitment sites. Ca²⁺/CaM-dependent protein kinase IIs (CaMKIIs) are autophosphorylated and interphosphorylated after integrating with CaM, leading to prolonged kinase activity [17]. In addition to the elevated cytosolic Ca²⁺ concentration which contributes to major signaling function in most cells, extracellular Ca²⁺ is also an important physiological signal [18]. CaSR, an extracellular Ca²⁺ receptor, couples both various heterotrimeric G-proteins and downstream signaling pathways, mediating pluripotent effects [19].

Ca²⁺ channels

The intricate fluxion of Ca²⁺ ions between extracellular and intracellular stores shapes the movement of Ca²⁺, such as Ca²⁺ release, Ca²⁺ oscillations, and Ca²⁺ spikes, modulating numerous biological functions [20, 21]. It is not surprising that the exchange of Ca²⁺ ions among different components of cells is interconnected and highly coordinated, and uncontrolled remodeling of this well-connected network may lead to cancer cells metastasis to bone.

Extracellular Ca^{2+} concentration is maintained at a high level ($\sim 1\text{--}2$ mmol/L), which is 10–20,000 times that of the cytosolic Ca^{2+} concentration (~ 100 nmol/L). Endoplasmic reticulum (ER) stores intracellular Ca^{2+} ions, with a Ca^{2+} concentration around 100–400 $\mu\text{mol/L}$ [22]. The regulation of this gradient is operated through a variety of mechanisms (Figure 1). Plasma membrane Ca^{2+} ATPases (PMCA) and sarco(endo)plasmic reticular Ca^{2+} ATPases (SERCA) are the main ATP-dependent channels that extrude Ca^{2+} ions from the cytosol to the extracellular space and ER, respectively. Inositol 1,4,5-trisphosphate receptors (IP_3Rs) initiate Ca^{2+} releasing from the ER [23]. After the depletion of the intracellular Ca^{2+} stores, store-operated Ca^{2+} entry (SOCE), a specific Ca^{2+} influx pathway, initiates Ca^{2+} influx through Orai1 Ca^{2+} channels after activation by the ER Ca^{2+} store sensor stromal interaction molecule 1 (STIM1) [24, 25]. Extracellular Ca^{2+} ions enter the cytoplasm through substantial mechanisms and are the primary origin for intracellular Ca^{2+} signaling in cells. Examples include store-operated Ca^{2+} channels (SOCs), the transient receptor potential (TRP) superfamily of ion channels, voltage-gated Ca^{2+} channels (VGCCs) including L-, R-, N-, P/Q-, and T-type channels, and stretch-activated PIEZO channels [23, 25–28].

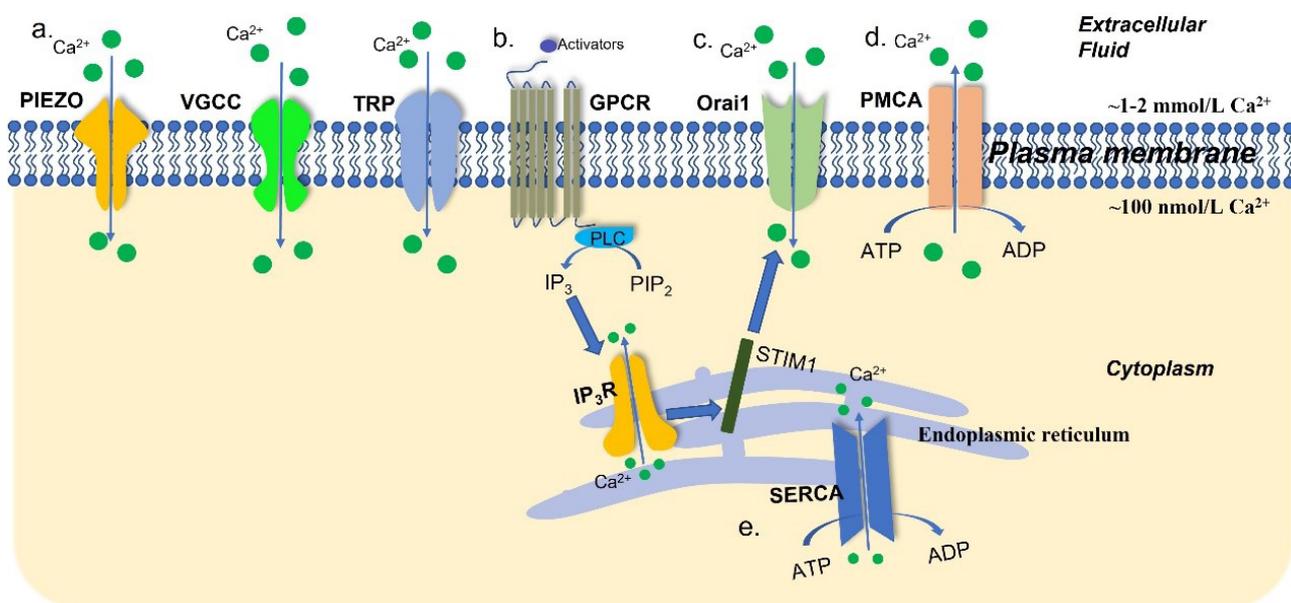


Figure 1. An overview of Ca^{2+} channels, transporters, and pumps in the plasma membrane and ER. Intracellular Ca^{2+} concentration is governed by a tightly mediated mechanism. (a) The TRP channels, VGCCs, and stretch-activated PIEZO channels are the Ca^{2+} channels and transporters in the plasma membrane; (b) after stimulation by activators, G-protein-coupled receptors (GPCRs) facilitate the dephosphorylation of phosphatidylinositol 4,5-bisphosphate (PIP_2) into inositol 1,4,5-trisphosphate (IP_3) by phospholipase C (PLC). In turn, IP_3Rs initiate Ca^{2+} release from the ER; (c) STIM1 senses the depletion of the ER Ca^{2+} stores and activates Ca^{2+} influx via Orai1 Ca^{2+} channels; (d) PMCA extrude Ca^{2+} ions from intracellular space to the extracellular space; (e) SERCA transport Ca^{2+} from the cytoplasm into ER. ADP: adenosine diphosphate

TRP channels related to Ca^{2+}

The mammalian TRP cation channel superfamily has 28 family members [29]. While TRP melastatin 3 α 2 (TRPM3 α 2), TRP vanilloid 5 (TRPV5), and TRPV6 are highly Ca^{2+} -selective, most TRP channels are nonselective [30]. Processes, such as cell apoptosis, proliferation, angiogenesis, invasion, and migration, are under the control of the regulation of the TRP cation channels in intracellular Ca^{2+} concentration (Table 1) [31]. Evidence has shown that TRPV2 mediates the secretion of RANKL via the Ca^{2+} -calcineurin-nuclear factor of activated T cells 3 (NFATc3) signaling pathway in multiple myeloma (MM) cells, and RANKL levels are demonstrated in a Ca^{2+} dose-dependent way (Figure 2a) [7]. Furthermore, NFATc3 was found to bind to the promoter of RANKL and induce RANKL expression at the transcriptional level [7]. The RANKL-induced bone remodeling contributed to the pathogenesis of MM lesions, but it also provided a likely treatment strategy.

Table 1. TRP channels and their functions in different cancers

Family	Members	Cancer type	Effects	References
TRPC	TRPC1	Colorectal cancer (CRC)	Enhanced cell proliferation, migration, invasion, and metastasis and apoptosis resistance	[32, 33]
	TRPC3	Gastric cancer	Tumorigenesis	[34]
		Breast cancer	Enhanced proliferation and apoptosis resistance	[35]
	TRPC5	CRC	Reduction in cancer differentiation	[36]
		Breast cancer	Chemotherapeutic resistance	[37]
	TRPC6	Hepatocellular carcinoma	Enhanced migration and invasion	[38]
	Breast cancer	Proliferation, migration, and invasion	[39]	
	Oesophageal cancer	Essential for G2 phase progression	[40]	
TRPV	TRPV2	Gastric cancer	Gastric cancer	[41]
	TRPV4	Gastric cancer	Enhanced proliferation, migration, and invasion	[42]
	TRPV6	Breast cancer	Tumor metastasis	[43]
TRPM	TRPM3	Clear cell renal cell carcinoma (RCC)	Tumor growth	[44]
	TRPM4	Prostate cancer	Enhanced proliferation	[45]
	TRPM7	Ovarian cancer	EMT and enhanced proliferation	[46, 47]

TRPC: TRP canonical

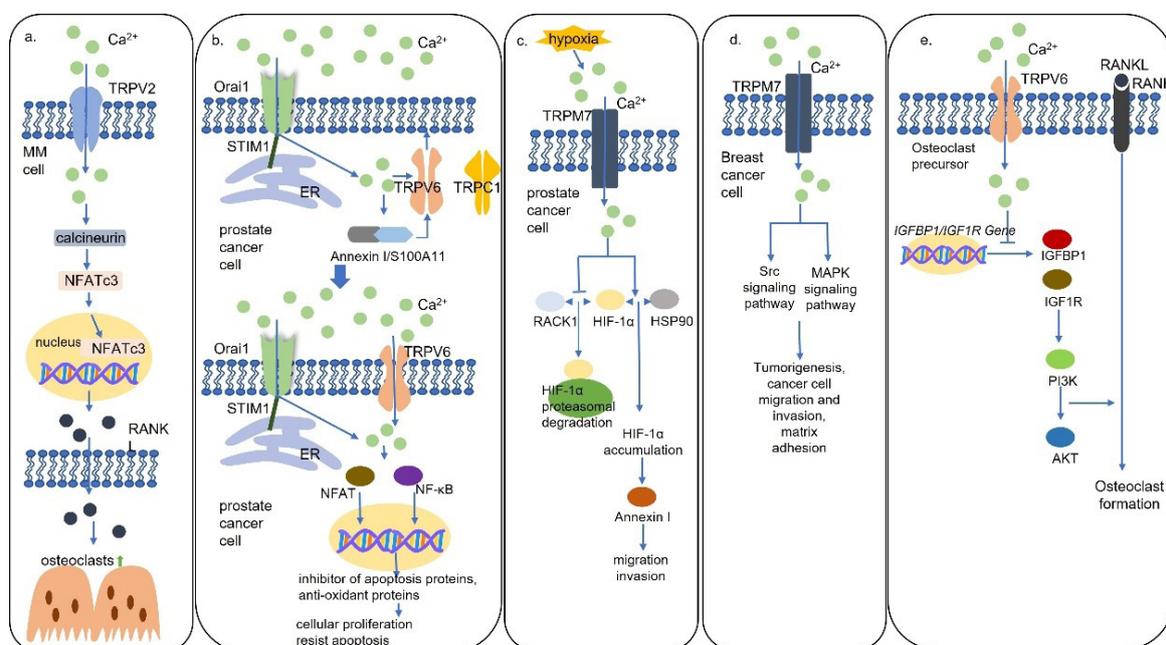


Figure 2. Schematic models for Ca^{2+} channels mediated Ca^{2+} signaling pathways. (a) TRPV2 mediates the secretion of RANKL via the Ca^{2+} -calcineurin-NFATc3 signaling pathway in MM cells to activate osteoclast formation; (b) TRPV6 channels are translocated to the plasma membrane via the STIM1/Orai1 mediated Ca^{2+} /Annexin I/S100A11 pathway. TRPV6 channels activate NFAT and nuclear factor κB (NF- κB) to promote proliferation, apoptosis resistance, and bone metastasis in prostate cancer cells; (c) hypoxia results in TRPM7-dependent hypoxia-inducible factor 1 α (HIF-1 α) accumulation which activates downstream Annexin I to promote cell migration and invasion in prostate cancer cells; (d) TRPM7 channels activate Src and mitogen-activated protein kinase (MAPK) signaling pathways to induce migration and invasion of breast cancer cells; (e) TRPV6 channels inhibit osteoclast formation by inhibiting the IGF/phosphatidylinositol 3-kinase (PI3K)/protein kinase B (AKT) signaling pathway. HSP90: heat shock protein 90; IGFBP1: insulin like growth factor binding protein 1; IGF1R: insulin like growth factor 1 receptor

TRPV6 was found to be present at elevated levels in prostate, breast, thyroid, colon, and ovarian carcinomas [48]. Moreover, TRPV6 messenger RNA (mRNA) expression levels were upregulated with the malignant degree of prostate cancer and the highest levels of TRPV6 were detected in prostate cancer with lymphatic metastases and in recurrent lesions [49]. It was found that the TRPV6 channels are translocated to the plasma membrane involving the STIM1/Orai1/TRPC1-mediated ER Ca^{2+} store depletion via the Ca^{2+} /Annexin I/S100A11 pathway, leading to increased proliferation and apoptosis resistance (Figure 2b) [50]. Furthermore, prostate cancer cells expressing TRPV6 were directly inoculated into the bone marrow cavity of tibias and promoted the generation of osteoblastic lesions suggesting TRPV6 promotes prostate cancer

bone metastasis via numerous osteoblastic lesions [50]. It is well studied that the increased intracellular Ca^{2+} induced by TRPV6 dephosphorylates NFAT to induce cellular proliferation and regulates NF- κ B oscillation to resist apoptosis, but the mechanism of the formation of the osteoblastic lesions is unknown [50].

The TRPM7 channels, which play a pivotal role in cell motility, are non-selective channels permeable predominantly to Mg^{2+} and Ca^{2+} [51]. In prostate cancer, an increase in serum $\text{Ca}^{2+}/\text{Mg}^{2+}$ ratio, which is regulated by TRPM7 and facilitates Ca^{2+} entry, leads to an increase in cell proliferation [52]. In a hypoxic environment, the increased TRPM7 results in HIF-1 α accumulation. TRPM7-HIF-1 α signaling activates downstream Annexin I protein expression mediating EMT, cell migration, and invasion (Figure 2c) [53]. In addition, it is also found that TRPM7 modulates the migration and invasion of breast cancer cells through the Src-MAPK signaling pathway (Figure 2d) [54]. Notably, TRPM7 overexpression promotes neuroblastoma cells to spread to the liver and bone marrow, but the mechanism is unknown [55].

It is well established that many members of the TRP cation channel superfamily play important roles in the mediation of tumor progression. The TRP cation channels also make a contribution to osteoclast differentiation, for example, TRPC1, TRPV4, and TRPV5 are essential for the regulation of osteoclastogenesis [56–58]. In a recent study, osteoporosis and enhanced bone absorption were found in TRPV6 knockout mice [59]. TRPV6 channels decreased the ratios of phosphorylation in the PI3K-AKT pathway which mediates the regulation of osteoclast formation and bone resorption (Figure 2e) [59]. The mechanism of the negative regulation of osteoclast differentiation and fusion and bone absorption by TRPV6 was revealed on a molecular level, and TRPV6 was confirmed to play an important role in bone metabolism [59]. Taken together with the formation of osteoblastic lesions induced by TRPV6, the TRPV6 channels play essential roles in the modulation of tumor progression and osteoclast activation. It makes one wonder, is there a connection between the TRPV6 generated osteoblastic lesions in prostate cancer and the TRPV6 negative regulated osteoclast differentiation? Further studies are required to elucidate the function of the TRP channels on cancer cell bone metastases and the possible link between metastases and the TRP regulation of osteoclast and osteoblast.

SOCs

SOCE follows the depletion of ER Ca^{2+} storage. Both STIM1 and STIM2 are important for the maintenance of intracellular Ca^{2+} concentration [60]. After reduction of ER intraluminal Ca^{2+} , STIM1 is activated and translocated to ER-plasma membrane junctions, where STIM proteins leash and gate Orai1 Ca^{2+} entry channels. STIM2 is more sensitive to changes in ER Ca^{2+} than STIM1, but it is a significantly weaker activator of Orai channels than STIM1.

In all, Orai1, -2, and -3 have been identified as plasma membrane Ca^{2+} channels. Although all three proteins are highly homologous to each other, they display notable differences in their features. Orai1 is the most potent to induce Ca^{2+} influx among its homologs, and its depletion significantly inhibits SOCE [61]. SOCE serves a wide set of signaling functions by elevating the cytosolic Ca^{2+} concentration. SOCE has potential roles in cellular proliferation and is inactivated during the division phase (M-phase) of the cell cycle. During the M-phase, STIM1 clustering is inhibited and Orai1 is internalized, thus uncoupling Ca^{2+} store depletion from Orai1 gating [62].

STIM1 and Orai1 are new targets for cancer treatment. Before STIM1's role in Ca^{2+} influx was suspected, it was implicated that STIM1 could be a tumor suppressor [63]. The role of STIM and Orai in cancer is better studied particularly in the case of breast cancer. Breast cancer cell lines are not homogenous regarding STIM/Orai expression. Orai1 and STIM1 are predominant in the estrogen receptor-negative breast cancer cell lines, but Orai3 and STIM1/2 are the main SOC in estrogen receptor-positive breast cancer cells [64]. The Orai3-induced Ca^{2+} influx contributed to breast cancer proliferation and survival but not in normal cells, consistent with the down-regulation of Orai3 arresting cell cycle progression and inducing apoptosis in breast cancer cells [65].

Focal adhesions, which are mediated by the interaction of integrin with the extracellular matrix, are relatively stable structures and tend to inhibit cell migration [66, 67]. Cell migration requires a dynamic state of focal adhesion [67]. STIM1 and Orai1 were shown to regulate tumor cell migration partially involving the

mediation of the focal adhesion [68]. Increased Ca^{2+} influx might induce tumor cell migration depending on the activation of the focal adhesion kinase (FAK), the Ca^{2+} -dependent protease calpain, and other Ca^{2+} -sensitive proteins in focal adhesion turnover. SOCE inhibitor SKF96365 inhibited breast cancer cells' metastasis in mouse models, providing a strong argument that SOCE is vital for breast tumor cell migration and metastasis.

Small conductance Ca^{2+} -activated potassium channel protein 3 (SK3), a potassium channel, is a member of the small conductance Ca^{2+} -activated potassium channel family [69]. An SK3-Orai1 complex, localized within lipid rafts, was found to be critical for the control of cancer cell migration and osteolytic bone metastases [70]. The SK3-Orai1 complex controls constitutive Ca^{2+} entry and tumor cell migration through store-independent Ca^{2+} signaling. Knocking down of the SK3 channels resulted in a lower metastatic score in breast cancer. Moreover, bone metastases achieved this lower metastatic score, but this reduction was not seen in lung metastases. The formation of the osteolytic lesions increased external Ca^{2+} concentration which amplified Ca^{2+} entry, establishing a vicious circle. Furthermore, the increased intracellular Ca^{2+} upregulated the activity of the Ca^{2+} -sensitive protease calpain which could be attributed to bone metastases. Ohmlin, a lipid inhibitor of SK3 channels [71], moved the SK3-Orai1 complex outside of lipid rafts and impaired the subsequent SK3-dependent Ca^{2+} entry, tumor cell migration, and bone metastases [70]. Therefore, ohmlin could be a promising therapeutic application in preventing and treating breast cancer bone metastases. However, the role of calpain in breast cancer bone metastases needs further study.

Serum- and glucocorticoid-inducible kinase 1 (SGK1) mediates osteoclast differentiation, bone resorption, and bone metastasis via the Orai1 [72]. The expression levels of SGK1 are essentially upregulated during RANKL-induced osteoclastogenesis [72]. It was found that treatment with GSK650394, an SGK1 inhibitor, down-regulated Orai1 levels during osteoclastogenesis and overexpressed Orai1 markedly alleviated the inhibitory effects of GSK650394 on osteoclast differentiation [72]. In addition, SGK1 is functionally relevant for cell migration, which is critically dependent on SOCE [73], and treatment with GSK650394 significantly decreased breast cancer bone metastases in mouse models [72]. These findings present a new perspective on RANKL-induced osteoclastogenesis and breast tumor bone metastases through SGK1-mediated Orai1 overexpression. However, reintroduction of Orai1 did not fully rescue the GSK650394 abolished Ca^{2+} influx [72]. There are possibly other SGK1-mediated Ca^{2+} channels that can be regarded as future therapeutic targets.

VGCCs

The VGCCs transport intracellular Ca^{2+} cations into intracellular Ca^{2+} transients initiating numerous physiological activities [26]. VGCCs are composed of three different subfamilies, the CaV1 (L-type) Ca^{2+} channel family, the CaV2 Ca^{2+} channel family, and the CaV3 (T-type) Ca^{2+} channel family, and are specified to ten members, CaV1.1, CaV1.2, CaV1.3, CaV1.4, CaV2.1, CaV2.2, CaV2.3, CaV3.1, CaV3.2, and CaV3.3 [74]. The CaV1 subfamily and the CaV2 subfamily are primarily responsible for the initiation of contraction, secretion, regulation of gene expression, integration of synaptic input in neurons, and synaptic transmission at ribbon synapses in specialized sensory cells, and initiation of synaptic transmission at fast synapses [26]. However, pieces of evidence have suggested that CaV3 mediates cellular processes including tumorigenesis and cancer progression by regulating intracellular Ca^{2+} levels [75].

T-type VGCCs expression levels are upregulated in many cancers and, thus, CaV3 channels are regarded as promising therapeutic targets. CaV3.1 isoform is a tumor-suppressor candidate and is reported to promote apoptosis and prevent tumor proliferation in breast cancer cells [76]. The CaV3.2 channels were not involved in the proliferation of MCF-7 breast cancer cells [76]. However, CaV3.1 was aberrantly upregulated and indicated a positive role in the regulation of proliferation in prostate cancer [77]. CaV3.1 together with CaV3.2 isoforms were found to increase gradually from normal skin to common nevi, dysplastic nevi, and melanoma samples with differences in distribution. Notably, metastatic melanoma showed the highest CaV3.2 expression levels which significantly differed from all other groups [78]. These results suggest that CaV3.1 and CaV3.2 channels may contribute to tumorigenesis and metastases. Therefore, further studies are required to identify the role of T-type VGCCs in cancers and bone metastasis, which is significant for the regulation of Ca^{2+} homeostasis.

Connexin 43

Connexin 43 [Cx43, encoded by gap junction protein alpha 1 (*GJA1*)] belongs to the connexin family which is the major constituent of gap junctions, widely connects osteocytes and osteoblasts in bone, and directs Ca^{2+} flow [79, 80]. Prostate cancer and breast cancer bone metastasis showed the highest levels of Cx43 expression among all sites of metastases, suggesting that bone colonization requires Ca^{2+} flows from osteoblasts to cancer cells via the Cx43-based gap junctions [12]. Importantly, arsenic trioxide (As_2O_3) can inhibit Ca^{2+} signaling through downregulation of Cx43 and affecting Ca^{2+} influx, making it a promising therapeutic agent for clinical practice (Figure 3) [12].

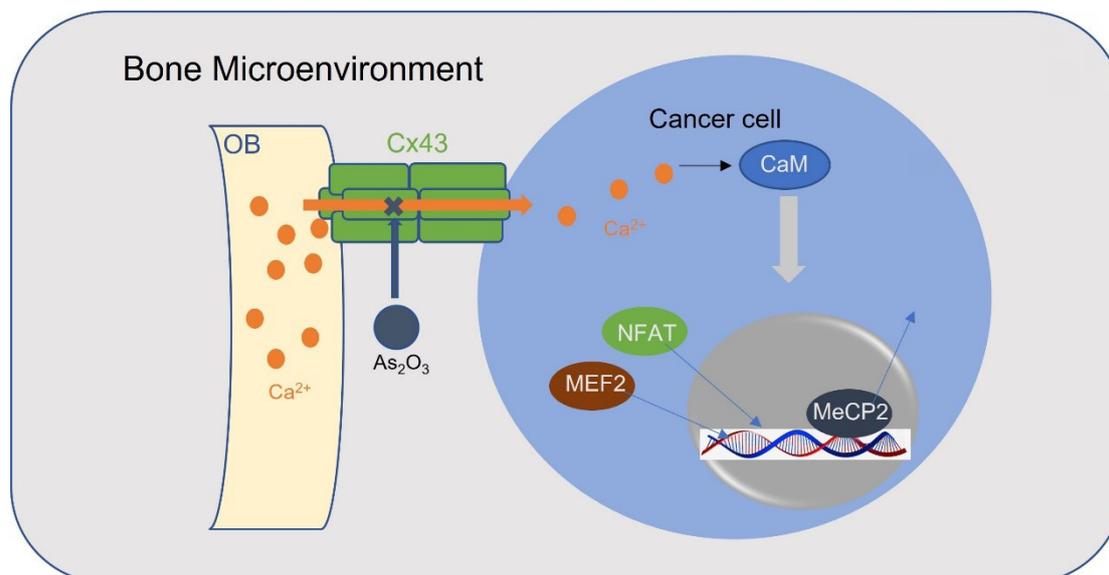


Figure 3. A schematic model for osteoblasts transports Ca^{2+} ions into cancer cells through Cx43 gap junctions. Ca^{2+} activates myocyte enhancer factor 2 (MEF2) and NFAT and releases methyl-CpG-binding protein 2 (MeCP2) from silenced promoters in a CaM-dependent manner. OB: osteoblast

CaMs

Ca^{2+} transducer CaMs are prominent Ca^{2+} sensors [81]. Ca^{2+} /CaM complexes bind to several classes of proteins and enzymes, including the CaM-dependent phosphatase calcineurin, myosin light-chain kinase, and Ca^{2+} /CaMK family, as well as many other enzymes, channels, transport systems, and transcription factors (TFs) [82]. CaM-dependent proteins have been known in tumor progression, including cell migration, tumor cell invasiveness, and metastasis, and they are emerging as potential anti-cancer therapeutic targets. High expression levels of CaM were discovered in neuroblastoma tumor cells, and, especially, an abundance of CaM was seen in bone marrow metastases [83]. Treatment with tamoxifen, an anti-CaM drug, was effective against neuroblastoma with bone marrow metastasis in a dose-dependent manner, but not with liver metastasis.

Ca^{2+} /CaMKs

Ca^{2+} /CaM-dependent protein kinase kinase α (CaMKK α) and β are the upstream kinases in the CaMK signaling cascade [84]. CaMKK α and β are activated through Ca^{2+} /CaM binding and intramolecular phosphorylation. Activated CaMKKs phosphorylate and activate CaMKI and CaMKIV, adenosine monophosphate (AMP)-activated protein kinase (AMPK), or AKT (PKB). These kinases then phosphorylate downstream proteins, such as cyclic AMP (cAMP) response element-binding protein (CREB), activating transcription factor-1 (ATF-1), CAAT-enhancer-binding protein (C/EBP), and serum response factor (SRF) [85]. Notably, different from CaMKK α , which is solely dependent on Ca^{2+} /CaM for activity, CaMKK β also can be activated in the absence of Ca^{2+} /CaM. Glycogen synthase kinase 3 β (GSK3 β) and cyclin-dependent kinase 5 (CDK5) regulate CaMKK β activities through phosphorylation [86]. Unlike CaMKI and CaMKIV, CaMKII requires Ca^{2+} /CaM complexes for activation, independent of CaMKKs [87].

CaMKK β was identified as a downstream target protein of androgen receptor that has been known as prostate cancer bone metastasis enhancer (Table 2). Androgen-dependent regulation of CaMKK β allowed tumor cells to migrate toward a more nutrient-rich environment, such as bone marrow, by activating and phosphorylating AMPK [88, 89]. Furthermore, CaMKK β is a critical regulator of bone remodeling and macrophage function, creating a favorable microenvironment for colonizing and tumor growth of prostate cancer cells [90]. CaMKK β stimulates osteoclast differentiation via CaMKK β -CaMKIV-phosphorylated cAMP response element binding (pCREB) signaling cascade its downstream target, the NFATc1, the primary mediator during osteoclastogenesis [91]. However, in osteoblast, the CaMKK β -CaMKIV pathway suppresses type I adenylate cyclase-cAMP regulated activities of protein kinase A (PKA), resulting in inhibited osteoblast differentiation [91]. Among immune cells, CaMKK β was found to be restrictedly expressed in cells of the monocytic/macrophage lineage [92]. CaMKK β ablation impaired macrophages' ability such as cytokine secretion, and morphological changes, and CaMKK β knockdown mice showed resistance to irritants that lead to systemic inflammation. Above all, dysregulation of CaMKK β remodels bone into a favorable environment for tumor cells. Knockdown of CaMKK β inhibits tumor growth, resists macrophage-induced inflammation, and improves the bone microenvironment. Further studies are still needed to investigate the molecular mechanisms of how CaMKK β mediates prostate cancer cells' metastatic abilities.

Table 2. Ca²⁺/CaMKs and their functions in different cancers

Components	Cancer type	Effects	References
CaMKK	Lung cancer	Tumor metastasis	[93]
	Prostate cancer	Tumor growth and castration resistance	[94]
	Glioma	Migration, invasion, and proliferation	[95]
CaMKI	Breast cancer	Control of cell cycle progression	[96]
CaMKII	Glioma	Migration and invasion	[97]
	Melanoma and hepatoma	Reprogramming of macrophages	[98]
	Prostate cancer	Inhibition of cancer growth and invasion, and induction of apoptosis	[99]
CaMKIV	Hepatic cancer	Cancer cell growth	[100]

Furthermore, increased cytolitic Ca²⁺ levels induced by Cx43 activate CaMKII mediating tumor cells' bone colonization [12]. Nuclear Ca²⁺ signaling induces the CaMKII-dependent MeCP2 phosphorylation on serine 421 of MeCP2 and releases MeCP2 from silenced promoters in many cellular contexts [101, 102]. Decreased levels of MeCP2 enriched TFs, NFAT, and MEF2 which are associated with the promotion of EMT, migration, angiogenesis, and invasion [103, 104], in bone metastases. Moreover, evidence has suggested that CaMKII may be co-regulatory with the Notch signaling pathway which plays a critical role in the development of osteometric properties by prostate cancer bone metastatic cells [105].

It has been established that CaMKII is involved in the differentiation of both osteoblasts and osteoclasts. A collagen-binding motif derived from osteopontin induces an influx of extracellular Ca²⁺ via Ca²⁺ channels and promotes osteoblastic differentiation via Ca²⁺/CaMKII/extracellular signal-regulated kinase (ERK)/activating protein-1 (AP-1) signaling pathway [106]. More importantly, increased osteoclastic resorption and subsequent bone loss are common features of bone metastases. Once osteoclasts are stimulated, activated CaM complexes combine with CaMKII to regulate the expression of NFATc1 and tartrate-resistant acid phosphatase (TRAP/ACP5), an osteoclast marker, leading to macrophage differentiation into osteoclasts [107]. Zoledronic acid, a bisphosphonate, significantly decreases the Ca²⁺ levels, inhibits the expression of CaM and CaMKII, and prevents osteoclasts differentiation, providing effective therapy for patients with skeletal involvement from advanced cancers [107]. Furthermore, CaMKII induces *c-fos* gene expression and subsequent AP-1 activation, which can, in turn, drive NFAT2 expression and is involved in osteoclast differentiation and bone remodeling [108]. CaMKII also mediates leukemia inhibitory factor (LIF)-induced phosphorylation of serine-782 in the glycoprotein 130 (gp130) tail, which leads to internalization and downregulation of the gp130 receptor on the cell surface, suggesting that CaMKII may promote osteoclastogenesis by inhibiting the gp130 receptor signaling cascade [108]. Zoledronic acid has proven to be efficient to treat bone metastases

targeting osteoclastogenesis, but zoledronic acid has nonnegligible side effects and limited application [107]. The molecular mechanisms of Ca^{2+} signaling leading to osteoclastogenesis may provide more specific targets to the treatment regimen for bone metastases, and needs further studies.

Ca²⁺/CaM-dependent phosphatase

Calcineurin is a conserved Ca^{2+} -CaM-dependent serine-threonine phosphatase that controls signaling pathways relevant to the migration, invasiveness, and metastatic potency of cancer cells. Increased cytosolic Ca^{2+} levels activate calcineurin mediating tumor cell bone colonization [12]. Calcineurin showed a similar effect as CaMKII to increase NFAT and MEF2 expression levels and inhibition of calcineurin also impedes bone colonization [12]. Calcineurin dephosphorylates resident NFAT proteins in the cytoplasm and triggers NFAT nuclear accumulation and activation [109]. RANK activation evokes Ca^{2+} oscillation by Ca^{2+} released from the ER and SOCE promotes CRC bone metastases through the calcineurin/NFATc1/ACP5 axis [110]. In addition, calcineurin/NFATc1 signaling promotes breast cancer metastasis to bone and brain and upregulates IGFI [111]. Regulator of calcineurin 1 isoform 4 (RCAN1.4) was found to reduce calcineurin activity and block nuclear translocation of NFATc1 [112]. Hence, RCAN1.4 is competent to reduce proliferation, migration, and metastases [112]. Moreover, RCAN1.4 was identified as a super suppressor of breast cancer and a potential therapeutic target for late-stage breast cancer patients with bone and brain lesions by ablation of calcineurin/NFATc1 signaling [111].

Furthermore, calcineurin A α (CnA α), an isoform of calcineurin, is significantly overexpressed in small cell lung cancer (SCLC) tissues with bone metastasis in contrast to tumor cells where bone metastasis was absent [113]. CnA α is located in nuclear SCLC cells with bone metastases, but in non-metastatic tumors, CnA α is mainly located in the cytosol [113]. Downregulation of CnA α by lentiviral vector-mediated RNA interference (RNAi) reduced cell migration and invasion, and inhibited adhesion to the bone matrix, hampering metastasis development of SCLC with no change in the apoptosis rate of tumor cells [114].

CaSR

CaSR is a GPCR that activates biased signaling in response to ligand stimulation [115]. With distinct ligand stimulation, CaSR preferentially activates relevant G proteins, including $G_{q/11}$, $G_{i/o}$, $G_{12/13}$, and Gs, facilitating selective regulation of the wide array of cellular effects [115]. CaSR senses fluctuations in extracellular Ca^{2+} and regulates intracellular and extracellular Ca^{2+} concentrations [19].

It has been known that CaSR controls Ca^{2+} homeostasis through its modulation of the parathyroid glands and kidneys, therefore contributing to chondrocytes, osteoblasts, and osteoclasts differentiation, leading to skeletal development and bone turnover [116]. Moreover, the role of extracellular Ca^{2+} and CaSR in cancers has been identified, promoting tumor cell proliferation, migration, and bone metastasis [117]. Another study showed a high CaSR expression in RCC, and high extracellular Ca^{2+} levels enhanced migratory potential and proliferation of bone metastasizing primary RCC cells [118].

Parathyroid hormone-related protein (PTHrP) is important for the induction of osteoclasts maturation and differentiation. Unlike suppressed PTHrP secretion by elevated Ca^{2+} in normal tissue, high Ca^{2+} concentrations stimulate CaSR to secrete PTHrP in prostate cancer, breast cancer, and lung cancer cells [119–121]. These cancers are referred to as humoral hypercalcemia of malignancies (HHMs), because of their systemic secretion of PTHrP which induce the secretion of RANKL in osteoblast, which in turn promotes osteoclast formation [122]. This Ca^{2+} -CaSR-PTHrP axis stimulates the differentiation of osteoclast precursors into mature osteoclast, therefore promoting bone resorption and Ca^{2+} release [122], initiating a vicious cycle, which contributes to the increased levels of Ca^{2+} and bone destruction.

In bone metastatic prostate cells, Ca^{2+} /CaSR upregulates the expression of cyclin D1, a key component of the cell cycle, to support cancer cell growth, but this upregulation is absent in the nonskeletal metastases [123]. Furthermore, activation of CaSR triggered prostate cancer cells' attachment, but the mechanism remains unknown [123]. In lung adenocarcinoma, CaSR was overexpressed in patients with bone metastasis, and overexpression of CaSR increased NF- κ B protein levels and subsequent matrix metalloproteinases 2 and

9 to enhance tumor cell invasion [120]. These results suggested that CaSR facilitates the development of bone metastasis.

Conclusions

The process and mechanism of bone metastasis are so complicated that there is no clear therapeutic target. The roles of Ca²⁺ signaling in tumor cells' metastasis to the bone have been well established. As a ubiquitous second message, Ca²⁺ interacts with cancer cells to promote proliferation, migration, and invasion. Moreover, bone has the biggest Ca²⁺ storage in the human body and Ca²⁺ signaling mediates osteoclasts and osteoblasts differentiation which can facilitate bone metastasis. Thus, Ca²⁺ ions' role in bone metastases is beyond tumor cells alone. It tells a better story along with osteoclasts, osteoblasts, and immune cells. Cancer cells' colonization in the bone environment depends on the destroyed bone structures and systemic inflammation induced by immune cells. Increased concentrations of intracellular Ca²⁺ have been proven to contribute to the progress of bone metastasis. However, future scholars should also investigate whether Ca²⁺ acts as a negative regulator of bone metastases. Numerous Ca²⁺ channels and Ca²⁺ signaling pathways have provided us with a plethora of potential therapeutical targets for cancer treatment. However, many Ca²⁺-associated channels, proteins, and kinases have not been investigated, and for most of the signaling pathways that have been studied, the specific mechanisms in migration, invasion, and metastasis of different types of cancers are only just beginning to be understood. Medications targeting the Ca²⁺ signaling toolkit are limited. Therefore, a better understanding of the exact molecular functions and mechanisms of Ca²⁺ signaling in bone metastases is needed and further efforts can focus on the Ca²⁺ channels, Ca²⁺-related signaling cascades, and their effects on bone metastases.

Abbreviations

AKT: protein kinase B

Ca²⁺: calcium

CaM: calmodulin

CaMKIIs: calcium/calmodulin-dependent protein kinase IIs

CaMKK α : calcium/calmodulin-dependent protein kinase kinase α

CaSR: calcium-sensing receptor

CnA α : calcineurin A α

CRC: colorectal cancer

Cx43: connexin 43

EMT: epithelial to mesenchymal transition

ER: endoplasmic reticulum

gp130: glycoprotein 130

GPCR: G-protein-coupled receptor

HIF-1 α : hypoxia-inducible factor 1 α

IGFI: insulin-like growth factor I

IP₃Rs: inositol 1,4,5-trisphosphate receptors

MAPK: mitogen-activated protein kinase

MeCP2: methyl-CpG-binding protein 2

MEF2: myocyte enhancer factor 2

MM: multiple myeloma

NFATc3: nuclear factor of activated T cells 3

NF- κ B: nuclear factor κ B

PI3K: phosphatidylinositol 3-kinase
PMCA: plasma membrane calcium ATPases
PTHrP: parathyroid hormone-related protein
RANK: receptor activator of nuclear factor-kappa B
RANKL: receptor activator of nuclear factor-kappa B ligand
RCAN1.4: regulator of calcineurin 1 isoform 4
RCC: renal cell carcinoma
SCLC: small cell lung cancer
SERCA: sarco(endo)plasmic reticular calcium ATPases
SGK1: serum- and glucocorticoid-inducible kinase 1
SK3: small conductance calcium-activated potassium channel protein 3
SOCE: store-operated calcium entry
SOCs: store-operated calcium channels
STIM1: stromal interaction molecule 1
TRP: transient receptor potential
TRPC: transient receptor potential canonical
TRPM3 α 2: transient receptor potential melastatin 3 α 2
TRPV5: transient receptor potential vanilloid 5
VGCCs: voltage-gated calcium channels

Declarations

Author contributions

TX, SC, JH, and ZL conceptualized the manuscript; TX, SC, and JH wrote this review; PW, HW, and ZL revised the manuscript. All authors have read and approved the submitted version of the manuscript.

Conflicts of interest

The authors declare that there are no conflicts of interest.

Ethical approval

Not applicable.

Consent to participate

Not applicable.

Consent to publication

Not applicable.

Availability of data and materials

Not applicable.

Funding

Not applicable.

Copyright

© The Author(s) 2022.

References

1. Coleman RE, Croucher PI, Padhani AR, Clézardin P, Chow E, Fallon M, et al. Bone metastases. *Nat Rev Dis Primers*. 2020;6:83.
2. Roodman GD. Mechanisms of bone metastasis. *N Engl J Med*. 2004;350:1655–64.
3. Paget S. The distribution of secondary growths in cancer of the breast. 1889. *Cancer Metastasis Rev*. 1989;8:98–101.
4. Kahn D, Weiner GJ, Ben-Haim S, Ponto LL, Madsen MT, Bushnell DL, et al. Positron emission tomographic measurement of bone marrow blood flow to the pelvis and lumbar vertebrae in young normal adults. *Blood*. 1994;83:958–63. Erratum in: *Blood*. 1994;84:3602.
5. Hauschka PV, Mavrikos AE, Iafrati MD, Doleman SE, Klagsbrun M. Growth factors in bone matrix. Isolation of multiple types by affinity chromatography on heparin-Sepharose. *J Biol Chem*. 1986;261:12665–74.
6. Yin JJ, Pollock CB, Kelly K. Mechanisms of cancer metastasis to the bone. *Cell Res*. 2005;15:57–62.
7. Bai H, Zhu H, Yan Q, Shen X, Lu X, Wang J, et al. TRPV2-induced Ca^{2+} -calcineurin-NFAT signaling regulates differentiation of osteoclast in multiple myeloma. *Cell Commun Signal*. 2018;16:68.
8. Li J, Sarosi I, Yan XQ, Morony S, Capparelli C, Tan HL, et al. RANK is the intrinsic hematopoietic cell surface receptor that controls osteoclastogenesis and regulation of bone mass and calcium metabolism. *Proc Natl Acad Sci U S A*. 2000;97:1566–71.
9. Chu GC, Chung LW. RANK-mediated signaling network and cancer metastasis. *Cancer Metastasis Rev*. 2014;33:497–509.
10. Chen X, Pei Z, Peng H, Zheng Z. Exploring the molecular mechanism associated with breast cancer bone metastasis using bioinformatic analysis and microarray genetic interaction network. *Medicine (Baltimore)*. 2018;97:e12032.
11. Ardura JA, Álvarez-Carrión L, Gutiérrez-Rojas I, Alonso V. Role of calcium signaling in prostate cancer progression: effects on cancer hallmarks and bone metastatic mechanisms. *Cancers (Basel)*. 2020;12:1071.
12. Wang H, Tian L, Liu J, Goldstein A, Bado I, Zhang W, et al. The osteogenic niche is a calcium reservoir of bone micrometastases and confers unexpected therapeutic vulnerability. *Cancer Cell*. 2018;34:823–39.e7.
13. Bootman MD. Calcium signaling. *Cold Spring Harb Perspect Biol*. 2012;4:a011171.
14. Wu L, Lian W, Zhao L. Calcium signaling in cancer progression and therapy. *FEBS J*. 2021;288:6187–205.
15. Patergnani S, Danese A, Bouhamida E, Aguiari G, Previati M, Pinton P, et al. Various aspects of calcium signaling in the regulation of apoptosis, autophagy, cell proliferation, and cancer. *Int J Mol Sci*. 2020;21:8323.
16. Prevarskaya N, Skryma R, Shuba Y. Calcium in tumour metastasis: new roles for known actors. *Nat Rev Cancer*. 2011;11:609–18.
17. Clapham DE. Calcium signaling. *Cell*. 2007;131:1047–58.
18. Bootman MD, Collins TJ, Peppiatt CM, Prothero LS, MacKenzie L, De Smet P, et al. Calcium signalling—an overview. *Semin Cell Dev Biol*. 2001;12:3–10.
19. Conigrave AD, Ward DT. Calcium-sensing receptor (CaSR): pharmacological properties and signaling pathways. *Best Pract Res Clin Endocrinol Metab*. 2013;27:315–31.
20. Berridge MJ, Bootman MD, Roderick HL. Calcium signalling: dynamics, homeostasis and remodelling. *Nat Rev Mol Cell Biol*. 2003;4:517–29.
21. Okada H, Okabe K, Tanaka S. Finely-tuned calcium oscillations in osteoclast differentiation and bone resorption. *Int J Mol Sci*. 2020;22:180.
22. Yang Z, Yue Z, Ma X, Xu Z. Calcium homeostasis: a potential vicious cycle of bone metastasis in breast cancers. *Front Oncol*. 2020;10:293.

23. Marchi S, Giorgi C, Galluzzi L, Pinton P. Ca²⁺ fluxes and cancer. *Mol Cell*. 2020;78:1055–69.
24. Mikoshiba K, Furuichi T, Miyawaki A. Structure and function of IP₃ receptors. *Semin Cell Biol*. 1994;5:273–81.
25. Roberts-Thomson SJ, Chalmers SB, Monteith GR. The calcium-signaling toolkit in cancer: remodeling and targeting. *Cold Spring Harb Perspect Biol*. 2019;11:a035204.
26. Catterall WA. Voltage-gated calcium channels. *Cold Spring Harb Perspect Biol*. 2011;3:a003947.
27. He L, Si G, Huang J, Samuel ADT, Perrimon N. Mechanical regulation of stem-cell differentiation by the stretch-activated Piezo channel. *Nature*. 2018;555:103–6.
28. Moran MM, McAlexander MA, Bíró T, Szallasi A. Transient receptor potential channels as therapeutic targets. *Nat Rev Drug Discov*. 2011;10:601–20.
29. Koivisto AP, Belvisi MG, Gaudet R, Szallasi A. Advances in TRP channel drug discovery: from target validation to clinical studies. *Nat Rev Drug Discov*. 2022;21:41–59.
30. Wu LJ, Sweet TB, Clapham DE. International Union of Basic and Clinical Pharmacology. LXXVI. Current progress in the mammalian TRP ion channel family. *Pharmacol Rev*. 2010;62:381–404.
31. Gautier M, Dhennin-Duthille I, Ay AS, Rybarczyk P, Korichneva I, Ouadid-Ahidouch H. New insights into pharmacological tools to TR(i)P cancer up. *Br J Pharmacol*. 2014;171:2582–92.
32. Sun Y, Ye C, Tian W, Ye W, Gao YY, Feng YD, et al. TRPC1 promotes the genesis and progression of colorectal cancer via activating CaM-mediated PI3K/AKT signaling axis. *Oncogenesis*. 2021;10:67.
33. Villalobos C, Hernández-Morales M, Gutiérrez LG, Núñez L. TRPC1 and ORAI1 channels in colon cancer. *Cell Calcium*. 2019;81:59–66.
34. Lin DC, Zheng SY, Zhang ZG, Luo JH, Zhu ZL, Li L, et al. TRPC3 promotes tumorigenesis of gastric cancer via the CNB2/GSK3β/NFATc2 signaling pathway. *Cancer Lett*. 2021;519:211–25.
35. Wang Y, Qi YX, Qi Z, Tsang SY. TRPC3 regulates the proliferation and apoptosis resistance of triple negative breast cancer cells through the TRPC3/RASA4/MAPK pathway. *Cancers (Basel)*. 2019;11:558.
36. Chen Z, Tang C, Zhu Y, Xie M, He D, Pan Q, et al. TrpC5 regulates differentiation through the Ca²⁺/Wnt5a signalling pathway in colorectal cancer. *Clin Sci (Lond)*. 2017;131:227–37.
37. Ma X, Chen Z, Hua D, He D, Wang L, Zhang P, et al. Essential role for TrpC5-containing extracellular vesicles in breast cancer with chemotherapeutic resistance. *Proc Natl Acad Sci U S A*. 2014;111:6389–94.
38. Xu J, Yang Y, Xie R, Liu J, Nie X, An J, et al. The NCX1/TRPC6 complex mediates TGFβ-driven migration and invasion of human hepatocellular carcinoma cells. *Cancer Res*. 2018;78:2564–76.
39. Jardin I, Diez-Bello R, Lopez JJ, Redondo PC, Salido GM, Smani T, et al. TRPC6 channels are required for proliferation, migration and invasion of breast cancer cell lines by modulation of Orai1 and Orai3 surface exposure. *Cancers (Basel)*. 2018;10:331.
40. Shi Y, Ding X, He ZH, Zhou KC, Wang Q, Wang YZ. Critical role of TRPC6 channels in G2 phase transition and the development of human oesophageal cancer. *Gut*. 2009;58:1443–50.
41. Kato S, Shiozaki A, Kudou M, Shimizu H, Kosuga T, Ohashi T, et al. TRPV2 promotes cell migration and invasion in gastric cancer via the transforming growth factor-β signaling pathway. *Ann Surg Oncol*. 2022;29:2944–56.
42. Xie R, Xu J, Xiao Y, Wu J, Wan H, Tang B, et al. Calcium promotes human gastric cancer via a novel coupling of calcium-sensing receptor and TRPV4 channel. *Cancer Res*. 2017;77:6499–512.
43. Xu X, Li N, Wang Y, Yu J, Mi J. Calcium channel TRPV6 promotes breast cancer metastasis by NFATC2IP. *Cancer Lett*. 2021;519:150–60.
44. Hall DP, Cost NG, Hegde S, Kellner E, Mikhaylova O, Stratton Y, et al. TRPM3 and miR-204 establish a regulatory circuit that controls oncogenic autophagy in clear cell renal cell carcinoma. *Cancer Cell*. 2014;26:738–53.

45. Sagredo AI, Sagredo EA, Cappelli C, Báez P, Andaur RE, Blanco C, et al. TRPM4 regulates AKT/GSK3- β activity and enhances β -catenin signaling and cell proliferation in prostate cancer cells. *Mol Oncol*. 2018;12:151–65.
46. Liu L, Wu N, Wang Y, Zhang X, Xia B, Tang J, et al. TRPM7 promotes the epithelial-mesenchymal transition in ovarian cancer through the calcium-related PI3K/AKT oncogenic signaling. *J Exp Clin Cancer Res*. 2019;38:106.
47. Chen Y, Liu L, Xia L, Wu N, Wang Y, Li H, et al. TRPM7 silencing modulates glucose metabolic reprogramming to inhibit the growth of ovarian cancer by enhancing AMPK activation to promote HIF-1 α degradation. *J Exp Clin Cancer Res*. 2022;41:44.
48. Zhuang L, Peng JB, Tou L, Takanaga H, Adam RM, Hediger MA, et al. Calcium-selective ion channel, CaT1, is apically localized in gastrointestinal tract epithelia and is aberrantly expressed in human malignancies. *Lab Invest*. 2002;82:1755–64.
49. Peng JB, Brown EM, Hediger MA. Epithelial Ca²⁺ entry channels: transcellular Ca²⁺ transport and beyond. *J Physiol*. 2003;551:729–40.
50. Raphaël M, Lehen'kyi V, Vandenberghe M, Beck B, Khalimonchuk S, Vanden Abeele F, et al. TRPV6 calcium channel translocates to the plasma membrane via Orai1-mediated mechanism and controls cancer cell survival. *Proc Natl Acad Sci U S A*. 2014;111:E3870–9.
51. Jimenez I, Prado Y, Marchant F, Otero C, Eltit F, Cabello-Verrugio C, et al. TRPM channels in human diseases. *Cells*. 2020;9:2604.
52. Sun Y, Selvaraj S, Varma A, Derry S, Sahmoun AE, Singh BB. Increase in serum Ca²⁺/Mg²⁺ ratio promotes proliferation of prostate cancer cells by activating TRPM7 channels. *J Biol Chem*. 2013;288:255–63.
53. Yang F, Cai J, Zhan H, Situ J, Li W, Mao Y, et al. Suppression of TRPM7 inhibited hypoxia-induced migration and invasion of androgen-independent prostate cancer cells by enhancing RACK1-mediated degradation of HIF-1 α . *Oxid Med Cell Longev*. 2020;2020:6724810.
54. Meng X, Cai C, Wu J, Cai S, Ye C, Chen H, et al. TRPM7 mediates breast cancer cell migration and invasion through the MAPK pathway. *Cancer Lett*. 2013;333:96–102.
55. Middelbeek J, Visser D, Henneman L, Kamermans A, Kuipers AJ, Hoogerbrugge PM, et al. TRPM7 maintains progenitor-like features of neuroblastoma cells: implications for metastasis formation. *Oncotarget*. 2015;6:8760–76.
56. Ong EC, Nesin V, Long CL, Bai CX, Guz JL, Ivanov IP, et al. A TRPC1 protein-dependent pathway regulates osteoclast formation and function. *J Biol Chem*. 2013;288:22219–32.
57. Masuyama R, Vriens J, Voets T, Karashima Y, Owsianik G, Vennekens R, et al. TRPV4-mediated calcium influx regulates terminal differentiation of osteoclasts. *Cell Metab*. 2008;8:257–65.
58. van der Eerden BC, Hoenderop JG, de Vries TJ, Schoenmaker T, Buurman CJ, Uitterlinden AG, et al. The epithelial Ca²⁺ channel TRPV5 is essential for proper osteoclastic bone resorption. *Proc Natl Acad Sci U S A*. 2005;102:17507–12.
59. Ma J, Zhu L, Zhou Z, Song T, Yang L, Yan X, et al. The calcium channel TRPV6 is a novel regulator of RANKL-induced osteoclastic differentiation and bone absorption activity through the IGF-PI3K-AKT pathway. *Cell Prolif*. 2021;54:e12955.
60. Soboloff J, Rothberg BS, Madesh M, Gill DL. STIM proteins: dynamic calcium signal transducers. *Nat Rev Mol Cell Biol*. 2012;13:549–65.
61. Várnai P, Hunyady L, Balla T. STIM and Orai: the long-awaited constituents of store-operated calcium entry. *Trends Pharmacol Sci*. 2009;30:118–28.
62. Courjaret R, Machaca K. STIM and Orai in cellular proliferation and division. *Front Biosci (Elite Ed)*. 2012;4:331–41.
63. Sabbioni S, Barbanti-Brodano G, Croce CM, Negrini M. *GOK*: a gene at 11p15 involved in rhabdomyosarcoma and rhabdoid tumor development. *Cancer Res*. 1997;57:4493–7.

64. Motiani RK, Abdullaev IF, Trebak M. A novel native store-operated calcium channel encoded by Orai3: selective requirement of Orai3 *versus* Orai1 in estrogen receptor-positive *versus* estrogen receptor-negative breast cancer cells. *J Biol Chem*. 2010;285:19173–83.
65. Faouzi M, Hague F, Potier M, Ahidouch A, Sevestre H, Ouadid-Ahidouch H. Down-regulation of Orai3 arrests cell-cycle progression and induces apoptosis in breast cancer cells but not in normal breast epithelial cells. *J Cell Physiol*. 2011;226:542–51.
66. Guo W, Giancotti FG. Integrin signalling during tumour progression. *Nat Rev Mol Cell Biol*. 2004;5:816–26.
67. Webb DJ, Parsons JT, Horwitz AF. Adhesion assembly, disassembly and turnover in migrating cells - over and over and over again. *Nat Cell Biol*. 2002;4:E97–100.
68. Yang S, Zhang JJ, Huang XY. Orai1 and STIM1 are critical for breast tumor cell migration and metastasis. *Cancer Cell*. 2009;15:124–34.
69. Köhler M, Hirschberg B, Bond CT, Kinzie JM, Marrion NV, Maylie J, et al. Small-conductance, calcium-activated potassium channels from mammalian brain. *Science*. 1996;273:1709–14.
70. Chantôme A, Potier-Cartereau M, Clarysse L, Fromont G, Marionneau-Lambot S, Guéguinou M, et al. Pivotal role of the lipid Raft SK3-Orai1 complex in human cancer cell migration and bone metastases. *Cancer Res*. 2013;73:4852–61.
71. Girault A, Haelters JP, Potier-Cartereau M, Chantome A, Pinault M, Marionneau-Lambot S, et al. New alkyl-lipid blockers of SK3 channels reduce cancer cell migration and occurrence of metastasis. *Curr Cancer Drug Targets*. 2011;11:1111–25.
72. Zhang Z, Xu Q, Song C, Mi B, Zhang H, Kang H, et al. Serum- and glucocorticoid-inducible kinase 1 is essential for osteoclastogenesis and promotes breast cancer bone metastasis. *Mol Cancer Ther*. 2020;19:650–60.
73. Eylonstein A, Gehring EM, Heise N, Shumilina E, Schmidt S, Sztejn K, et al. Stimulation of Ca²⁺-channel Orai1/STIM1 by serum- and glucocorticoid-inducible kinase 1 (SGK1). *FASEB J*. 2011;25:2012–21.
74. Zamponi GW, Striessnig J, Koschak A, Dolphin AC. The physiology, pathology, and pharmacology of voltage-gated calcium channels and their future therapeutic potential. *Pharmacol Rev*. 2015;67:821–70.
75. Bhargava A, Saha S. T-type voltage gated calcium channels: a target in breast cancer? *Breast Cancer Res Treat*. 2019;173:11–21.
76. Ohkubo T, Yamazaki J. T-type voltage-activated calcium channel Cav3.1, but not Cav3.2, is involved in the inhibition of proliferation and apoptosis in MCF-7 human breast cancer cells. *Int J Oncol*. 2012;41:267–75.
77. Hu S, Li L, Huang W, Liu J, Lan G, Yu S, et al. CAV3.1 knockdown suppresses cell proliferation, migration and invasion of prostate cancer cells by inhibiting AKT. *Cancer Manag Res*. 2018;10:4603–14.
78. Maiques O, Macià A, Moreno S, Barceló C, Santacana M, Veà A, et al. Immunohistochemical analysis of T-type calcium channels in acquired melanocytic naevi and melanoma. *Br J Dermatol*. 2017;176:1247–58.
79. Plotkin LI, Bellido T. Beyond gap junctions: Connexin43 and bone cell signaling. *Bone*. 2013;52:157–66.
80. Osswald M, Jung E, Sahm F, Solecki G, Venkataramani V, Blaes J, et al. Brain tumour cells interconnect to a functional and resistant network. *Nature*. 2015;528:93–8.
81. Chin D, Means AR. Calmodulin: a prototypical calcium sensor. *Trends Cell Biol*. 2000;10:322–8.
82. O'Day DH, Taylor RJ, Myre MA. Calmodulin and calmodulin binding proteins in *Dictyostelium*: a primer. *Int J Mol Sci*. 2020;21:1210.
83. Iwakawa M, Ando K, Ohkawa H, Koike S, Chen YJ. A murine model for bone marrow metastasis established by an i.v. injection of C-1300 neuroblastoma in A/J mice. *Clin Exp Metastasis*. 1994;12:231–7.
84. Brzozowski JS, Skelding KA. The multi-functional calcium/calmodulin stimulated protein kinase (CaMK) family: emerging targets for anti-cancer therapeutic intervention. *Pharmaceuticals (Basel)*. 2019;12:8.

85. Hook SS, Means AR. Ca²⁺/CaM-dependent kinases: from activation to function. *Annu Rev Pharmacol Toxicol.* 2001;41:471–505.
86. Green MF, Scott JW, Steel R, Oakhill JS, Kemp BE, Means AR. Ca²⁺/calmodulin-dependent protein kinase kinase β is regulated by multisite phosphorylation. *J Biol Chem.* 2011;286:28066–79.
87. Shifman JM, Choi MH, Mihalas S, Mayo SL, Kennedy MB. Ca²⁺/calmodulin-dependent protein kinase II (CaMKII) is activated by calmodulin with two bound calciums. *Proc Natl Acad Sci U S A.* 2006;103:13968–73.
88. Frigo DE, Howe MK, Wittmann BM, Brunner AM, Cushman I, Wang Q, et al. CaM kinase kinase β -mediated activation of the growth regulatory kinase AMPK is required for androgen-dependent migration of prostate cancer cells. *Cancer Res.* 2011;71:528–37.
89. Shima T, Mizokami A, Miyagi T, Kawai K, Izumi K, Kumaki M, et al. Down-regulation of calcium/calmodulin-dependent protein kinase kinase 2 by androgen deprivation induces castration-resistant prostate cancer. *Prostate.* 2012;72:1789–801.
90. Dadwal UC, Chang ES, Sankar U. Androgen receptor-CaMKK2 axis in prostate cancer and bone microenvironment. *Front Endocrinol (Lausanne).* 2018;9:335.
91. Cary RL, Waddell S, Racioppi L, Long F, Novack DV, Voor MJ, et al. Inhibition of Ca²⁺/calmodulin-dependent protein kinase kinase 2 stimulates osteoblast formation and inhibits osteoclast differentiation. *J Bone Miner Res.* 2013;28:1599–610.
92. Racioppi L, Noeldner PK, Lin F, Arvai S, Means AR. Calcium/calmodulin-dependent protein kinase kinase 2 regulates macrophage-mediated inflammatory responses. *J Biol Chem.* 2012;287:11579–91.
93. Jin L, Chun J, Pan C, Kumar A, Zhang G, Ha Y, et al. The PLAG1-GDH1 axis promotes anoikis resistance and tumor metastasis through CamKK2-AMPK signaling in LKB1-deficient lung cancer. *Mol Cell.* 2018;69:87–99.e7.
94. Wang N, Yao M, Xu J, Quan Y, Zhang K, Yang R, et al. Autocrine activation of CHRM3 promotes prostate cancer growth and castration resistance via CaM/CaMKK-mediated phosphorylation of AKT. *Clin Cancer Res.* 2015;21:4676–85.
95. Liu DM, Wang HJ, Han B, Meng XQ, Chen MH, Yang DB, et al. CAMKK2, regulated by promoter methylation, is a prognostic marker in diffuse gliomas. *CNS Neurosci Ther.* 2016;22:518–24.
96. Rodriguez-Mora OG, LaHair MM, McCubrey JA, Franklin RA. Calcium/calmodulin-dependent kinase I and calcium/calmodulin-dependent kinase kinase participate in the control of cell cycle progression in MCF-7 human breast cancer cells. *Cancer Res.* 2005;65:5408–16.
97. Yu-Ju Wu C, Chen CH, Lin CY, Feng LY, Lin YC, Wei KC, et al. CCL5 of glioma-associated microglia/macrophages regulates glioma migration and invasion via calcium-dependent matrix metalloproteinase 2. *Neuro Oncol.* 2020;22:253–66.
98. Dai X, Meng J, Deng S, Zhang L, Wan C, Lu L, et al. Targeting CAMKII to reprogram tumor-associated macrophages and inhibit tumor cells for cancer immunotherapy with an injectable hybrid peptide hydrogel. *Theranostics.* 2020;10:3049–63.
99. Tan H, Zhang G, Yang X, Jing T, Shen D, Wang X. Peimine inhibits the growth and motility of prostate cancer cells and induces apoptosis by disruption of intracellular calcium homeostasis through Ca²⁺/CaMKII/JNK pathway. *J Cell Biochem.* 2020;121:81–92.
100. Lin F, Marcelo KL, Rajapakshe K, Coarfa C, Dean A, Wilganowski N, et al. The camKK2/camKIV relay is an essential regulator of hepatic cancer. *Hepatology.* 2015;62:505–20.
101. Buchthal B, Lau D, Weiss U, Weislogel JM, Bading H. Nuclear calcium signaling controls methyl-CpG-binding protein 2 (MeCP2) phosphorylation on serine 421 following synaptic activity. *J Biol Chem.* 2012;287:30967–74.
102. Li H, Zhong X, Chau KF, Santistevan NJ, Guo W, Kong G, et al. Cell cycle-linked MeCP2 phosphorylation modulates adult neurogenesis involving the Notch signalling pathway. *Nat Commun.* 2014;5:5601.

103. Mancini M, Toker A. NFAT proteins: emerging roles in cancer progression. *Nat Rev Cancer*. 2009;9:810–20.
104. Di Giorgio E, Hancock WW, Brancolini C. MEF2 and the tumorigenic process, hic sunt leones. *Biochim Biophys Acta Rev Cancer*. 2018;1870:261–73.
105. Mamaeva OA, Kim J, Feng G, McDonald JM. Calcium/calmodulin-dependent kinase II regulates notch-1 signaling in prostate cancer cells. *J Cell Biochem*. 2009;106:25–32.
106. Shin MK, Kim MK, Bae YS, Jo I, Lee SJ, Chung CP, et al. A novel collagen-binding peptide promotes osteogenic differentiation via Ca²⁺/calmodulin-dependent protein kinase II/ERK/AP-1 signaling pathway in human bone marrow-derived mesenchymal stem cells. *Cell Signal*. 2008;20:613–24.
107. Wang L, Fang D, Xu J, Luo R. Various pathways of zoledronic acid against osteoclasts and bone cancer metastasis: a brief review. *BMC Cancer*. 2020;20:1059.
108. Seales EC, Micoli KJ, McDonald JM. Calmodulin is a critical regulator of osteoclastic differentiation, function, and survival. *J Cell Biochem*. 2006;97:45–55.
109. Hogan PG, Chen L, Nardone J, Rao A. Transcriptional regulation by calcium, calcineurin, and NFAT. *Genes Dev*. 2003;17:2205–32.
110. Liang Q, Wang Y, Lu Y, Zhu Q, Xie W, Tang N, et al. RANK promotes colorectal cancer migration and invasion by activating the Ca²⁺-calcineurin/NFATc1-ACP5 axis. *Cell Death Dis*. 2021;12:336.
111. Deng R, Huang JH, Wang Y, Zhou LH, Wang ZF, Hu BX, et al. Disruption of super-enhancer-driven tumor suppressor gene RCAN1.4 expression promotes the malignancy of breast carcinoma. *Mol Cancer*. 2020;19:122.
112. Jin H, Wang C, Jin G, Ruan H, Gu D, Wei L, et al. Regulator of calcineurin 1 gene isoform 4, down-regulated in hepatocellular carcinoma, prevents proliferation, migration, and invasive activity of cancer cells and metastasis of orthotopic tumors by inhibiting nuclear translocation of NFAT1. *Gastroenterology*. 2017;153:799–811.e33.
113. Liu Y, Zhang Y, Min J, Liu LL, Ma NQ, Feng YM, et al. Calcineurin promotes proliferation, migration, and invasion of small cell lung cancer. *Tumour Biol*. 2010;31:199–207.
114. Ma NQ, Liu LL, Min J, Wang JW, Jiang WF, Liu Y, et al. The effect of down regulation of calcineurin A α by lentiviral vector-mediated RNAi on the biological behavior of small-cell lung cancer and its bone metastasis. *Clin Exp Metastasis*. 2011;28:765–78.
115. Leach K, Hannan FM, Josephs TM, Keller AN, Møller TC, Ward DT, et al. International Union of Basic and Clinical Pharmacology. CVIII. Calcium-sensing receptor nomenclature, pharmacology, and function. *Pharmacol Rev*. 2020;72:558–604.
116. Goltzman D, Hendy GN. The calcium-sensing receptor in bone—mechanistic and therapeutic insights. *Nat Rev Endocrinol*. 2015;11:298–307.
117. Tuffour A, Kosiba AA, Zhang Y, Peprah FA, Gu J, Shi H. Role of the calcium-sensing receptor (CaSR) in cancer metastasis to bone: identifying a potential therapeutic target. *Biochim Biophys Acta Rev Cancer*. 2021;1875:188528.
118. Joeckel E, Haber T, Prawitt D, Junker K, Hampel C, Thüroff JW, et al. High calcium concentration in bones promotes bone metastasis in renal cell carcinomas expressing calcium-sensing receptor. *Mol Cancer*. 2014;13:42.
119. Sanders JL, Chattopadhyay N, Kifor O, Yamaguchi T, Brown EM. Ca²⁺-sensing receptor expression and PTHrP secretion in PC-3 human prostate cancer cells. *Am J Physiol Endocrinol Metab*. 2001;281:E1267–74.
120. Liu L, Fan Y, Chen Z, Zhang Y, Yu J. CaSR induces osteoclast differentiation and promotes bone metastasis in lung adenocarcinoma. *Front Oncol*. 2020;10:305.
121. Das S, Clézardin P, Kamel S, Brazier M, Mentaverri R. The CaSR in pathogenesis of breast cancer: a new target for early stage bone metastases. *Front Oncol*. 2020;10:69.

122. Zagzag J, Hu MI, Fisher SB, Perrier ND. Hypercalcemia and cancer: differential diagnosis and treatment. *CA Cancer J Clin.* 2018;68:377–86.
123. Liao J, Schneider A, Datta NS, McCauley LK. Extracellular calcium as a candidate mediator of prostate cancer skeletal metastasis. *Cancer Res.* 2006;66:9065–73.