

## Open Access Review



# Targeted therapy of multiple myeloma

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## Abstract

Multiple myeloma (MM) is a malignant proliferative disease of monoclonal plasma cells (PCs) and is characterized by uncontrolled proliferation of PCs and excessive production of specific types of immunoglobulins. Since PCs are terminally differentiated B cells, the World Health Organization (WHO) classifies MM as lymphoproliferative B-cell disease. The incidence of MM is 6-7 cases per 100,000 people in the world every year and the second most common cancer in the blood system. Due to the effects of drug resistance and malignant regeneration of MM cells in the microenvironment, all current treatment methods can prolong both overall and symptom-free survival rates of patients with MM but cannot cure MM. Both basic and clinical studies have proven that targeted therapy leads to a clear and significant prolongation of the survival of patients with MM, but when the disease recurs again, resistance to the previous treatment will occur. Therefore, the discovery of new targets and treatment methods plays a vital role in the treatment of MM. This article introduces and summarizes targeted MM therapy, potential new targets, and future precision medicine in MM.

## **Keywords**

Multiple myeloma, targeted therapy, potential targets, immune-based therapies, precision medicine

## Introduction

In terms of disease progression, multiple myeloma (MM) develops from a malignant precursor disease, progresses to monoclonal gammopathy of undetermined significance (MGUS), and then to smoldering MM (SMM) followed by active MM, which is often accompanied by anemia (usually the first symptom), bone pain (the most common symptom), renal insufficiency, infection, bleeding, neurological symptoms, hypercalcemia, amyloidosis, and other symptoms. Due to relapse after treatment, MM will progress to relapsed/refractory MM (RRMM). Most of these cases eventually develop into advanced plasma cell leukemia (PCL). At a molecular level, MM is caused by single or multiple gene mutations, heterotopic, chromosome changes, and other factors and mediated by abnormal chromosome hyper-diploidy,

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immunoglobulin heavy chain shift, cell cycle gene imbalance, nuclear factor-kappaB (NF- $\kappa$ B) pathway changes, and abnormal DNA methylation patterns [1]. In addition, it has been proven that MM cells also rely on the bone marrow (BM) microenvironment to support their growth, survival, and drug resistance. This interaction between cells and the microenvironment promotes the proliferation of tumor cells and weakens the immune surveillance and effector function of MM cells so that tumor cells can grow in an uncontrolled and unrestricted manner. Based on MM pathophysiology and the microenvironment, targeted therapies have been ongoing (Figure 1).



Figure 1. An overview of the different targeted therapies of therapeutic interest in MM. Targeted therapies are divided into 4 sections: 1. Clinically validated targeted therapies already used in treatment of MM including cullin-4 E3 ligase-targeted thalidomide. proteasome inhibitor bortezomib, NF-KB pathway inhibitor lenalidomide and pomalidomide, CD38-targeted daratumumab and isatuximab-irfc, SLAM7-targeted elotuzumab, HDAC-targeted panobinostat exportin-1 inhibitor selinexor; 2. Clinical development of targeted therapies in MM including immune-based therapies, targeting genomic instability, targeting aberrant regulatory network, epigenetic (Epi)-therapeutics, CELMoDs therapeutics, mutation-targeted therapies, targeting the microenvironment, and so on; 3. Other targeted therapies based on preclinical data including targeting CircRNA, NF-KB, negative regulators, and so on; 4. Future personalized treatment strategies in MM including molecular approaches related to diagnosis, risk stratification and precision medicine. SLAM7: signaling lymphocytic activation molecule family member 7; HDAC: histone deacetylase; Tim3: T cell immunoglobulin-3; BiTE: bispecific T-cell engager; BCMA: B-cell maturation antigen; ADC: antibody-drug conjugates; CAR-T: chimeric antigen receptor (CAR) T-cell (CAR-T); ATR: ataxia telangiectasia and Rad3 related; IRF4: interferon regulatory factor 4; ASO: antisense oligonucleotide; c-Myc: cellular myelocytomatosis; BRAF: v-raf murine sarcoma viral oncogene homolog B; MEK: mitogen-activated extracellular signal-regulated kinase; PD-1: programmed cell death-1; PD-L1: programmed death-ligand 1; IL-18: interleukin 18; Circ: Circular; SUVs: standard uptake values; MATV: metabolically active tumour volume; TLG: total lesion glycolysis; RANK: receptor activator of nuclear factor kappa B; RANKL: receptor activator of nuclear faktor kappa B ligand; CXCR4: C-X-C chemokine receptor type 4; CXCL12: C-X-C chemokine ligand 12; TP53: tumor protein p53; RB1: RB transcriptional corepressor 1; MDM2: mouse double minute 2; RUNX: runt-related transcription factor 1; EZH2: enhancer of zeste homolog 2; YY1: Yin Yang 1; CELMoDs: cereblon E3 ligase-modulating drugs

# Clinically validated targeted therapies already used in the treatment of MM

In the past, the Food and Drug Administration (FDA) approved some treatment drugs and modalities for MM (Table 1) [2-13]. In recent years, clinical guidelines have divided the treatment of MM into two categories: (1) those suitable for transplantation and (2) those not suitable for transplantation. For patients who are not suitable for transplantation, cytotoxic drugs are generally selected in combination with chemotherapy. These cytotoxic drugs generally have a slow onset, but relief from symptoms lasts a long time, and the time interval for suppressing recurrence is long, which can prolong the survival period of patients and delay the progression of clinical symptoms. For patients who are suitable for transplantation, targeted therapy is usually used clinically to enable the patient to undergo transplantation as soon as possible after remission. Generally, in patients who need BM transplantation, bortezomib (Velcade), lenalidomide (Revlimid), and dexamethasone (VRD) are used for about 3 to 4 treatment cycles before transplantation is performed. Patients who are not suitable for transplantation can also be treated with the above-mentioned drugs for 8 to 12 cycles, and then lenalidomide is used for maintenance treatment. Even though these treatments have a significant effect on prolonging the survival period of MM [14, 15], due to the heterogeneity of the sensitivity of patients to drugs and the need for long-term medication for patients with MM. Many patients have developed drug resistance to existing drugs, which leads to the recurrence of MM. The survival rate of recurrent MM is extremely low, so the discovery of new drug targets and the research of new therapies are urgently needed with respect to the treatment of MM.

Years	Main drugs/ treatment method	Composition or function	Application effect
1960	Melphalan Cyclophosphamide corticosteroids	Alkylating agent	
1990	HSCT	Autologous hematopoietic stem-cell transplantation.	
1990	Thalidomide	Immunomodulatory; the first drug to show activity in RRMM; the main function is to suppress angiogenesis.	Toxic and side effects of the dose on peripheral nerves.
2003	Bortezomib	The proteasome inhibitor, which inhibits the NF-κB pathway, is often involved in the combined treatment of NDMM and RRMM and triggers "immunogenic" cell death, which can trigger effective anti-MM immune responses and disease control.	
2005	Lenalidomide	Immunomodulator that inhibits NF-kB pathway, activates caspase and promotes apoptosis [5]. It is the first-line drug for newly discovered MM. It is mainly involved in immune regulation, suppressing angiogenesis and proliferation.	Combined with bortezomib and dexamethasone (VRD), the 4-year overall survival rate was 81%. More than 2% of the patients had grade 3/4 adverse reactions: 63.7% of the patients had blood and lymphatic system diseases, and 13.7% had nervous system diseases [2].
2013	Pomalidomide	Immunomodulator inhibits NF-κB pathway, activates caspase, and promotes apoptosis [5].	Currently, it is approved to be combined with dexamethasone, elozumab, and dalatuzumab [3]. The median of PVD and VD was 11.2 and 7.1 (HR = 0.61, 95% CI: 0.49-0.77; $P < 0.0001$ ). Hematologic toxicity, gastrointestinal side effects, and rash are common.
2015	Daratumumab Elotuzumab	Monoclonal antibodies targeting CD38 and slamf7. Treatment of newly discovered and refractory MM.	D-RVd improved the complete remission rate and the minimal residual disease (MRD) negative rate, and the single drug remission rate was 29.2% [6]. Grade 3/4 neutropenia: d-RVd/RVD 41.4%, 21.6%; lymphopenia: 23.2%, 21.6%; thrombocytopenia: 16.2%, 8.8% [13].

Table 1. The current drugs and treatment methods for MM

Table 1. The current drugs and treatment methods for MM (	(continued)
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Years	Main drugs/ treatment method	Composition or function	Application effect
2015	Panobinostat	Pan-HDAC inhibitor. For RRMM.	In combination with bortezomib and dexamethasone, the PFS was significantly improved, but the side effects were significant [11]. The overall response rate was 34.5%, the median time to response was 1.4 months [9]. Thrombocytopenia 63.6%, fatigue, and diarrhea 20.0%. The incidence of serious adverse reactions was 67.3% [9].
2019	Selinexor	Exportin-1 inhibitor specifically blocks xpo1 protein. For the treatment of RRMM, especially grade 3 refractory myelomas.	It is often combined with dexamethasone. The median duration of PFS was 13.93 months in the SVD group and 9.46 months in the VD group. This means that the median PFS increased by 4.47 months. Nausea, vomiting, loss of appetite. 73% of patients had thrombocytopenia [10].
2020	Isatuximab-irfc	The monoclonal antibody, targeting CD38, is used for RRMM. The immune effect function of MM was restored by blocking immunosuppressive T cells [4].	It was combined with pomalidomide/ dexamethasone (PD). The median PFS of ISA-PD was 11.5 months, the median PFS of PD was 6.5 months, the HR was 0.60, the ORR of ISA-PD was 60.4%, and the ORR of PD was 35.3% [8]. In monotherapy, infusion response was 51%, fatigue 37%, nausea 32%, upper respiratory tract infection 24% and cough 23% [7]. Non-hematological adverse events caused by is a PD: fatigue 64%, ir40%, upper respiratory tract infection, and cough 40% [12].

HSCT: hematopoietic stem cell transplantation; PFS: progression-free survival; ORR: objective response rate; HR: hazard ratio; NDMM: newly diagnosed MM; PVD: pomalidomide + bortezomib (Velcade) + dexamethasone (DEX); VD: bortezomib (Velcade) + dexamethasone (DEX); D-RVd: daratumumab + lenalidomide (Revlimid) + bortezomib (Velcade) + dexamethasone (DEX); SVD: Selinexor + bortezomib (Velcade) + dexamethasone (DEX); ISA-PD: isatuximab with pomalidomide/dexamethasone; xpo1: exportin 1

# **Clinical development of targeted therapies in MM**

## **Immune-based therapies**

Immune-based therapies such as immune checkpoint Tim3 inhibitors, BiTE immunotherapy, ADC immunotherapy, and CAR-T (Table 2) immunotherapy have been becoming therapeutic hot-researches in MM.

	Table 2.	Immune-based	therapies	in	MM
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Therapy	Structure	Trial stage	Dosage and effective rate	Adverse reactions	Approved usage
Tim3 inhibitors such as LY3321367, MG453	The small molecular inhibitor	All of them have completed phase I clinical trials and have shown antitumor activity	Anti Tim3 drug tsr-022, Tim3 blocking antibody LY3321367, MG453		Tim3 antibody has been clearly expressed to improve AML
BiTE® BI 836909 [16-18]	Single-chain variable fragment (scFv) composed of anti-BCMA scFv at the N-terminal and anti-CD3ɛ scFv at the C-terminal, followed by hexahistidine	Animal experiment	0.5, 0.05, 0.005 mg/kg per day for 26 days; median survival was 43.5 days (0.5 and 0.05 mg/kg per day) and 43 days (0.005 mg/ kg per day), whereas the control group was 34 days	Neurotoxicity	In 2014, FDA approved for the treatment of Philadelphia chromosome-negative relapsed/refractory B-cell precursor acute lymphoblastic leukemia [6]

Therapy	Structure	Trial stage	Dosage and effective rate	Adverse reactions	Approved usage
Blenrep	Joint: non cleavable MC; Payload: MMAF	The first stage NCT020 64387, DREAMM-1 [19, 20]	4 mg/kg every 3 weeks ORR 60%; sCR 2 (6%), CR 3 (9%), VGPR 14 (40%); mPFS 12 months; mDOR 14.3 months	Grade 3 and 4 thrombocytopenia (35%), anemia (17%); hemocytopenia (52%); G1, 2 corneal events: blurred vision (52%), dry eyes (37%)	In early August 2020, it was approved in the United States for the treatment of relapsed or refractory MM in adult patients who have received at least four previous therapies, including anti-CD38 monoclonal antibody.
		The second stage, NCT03525678, DREAMM-2 [21]	2.5 or 3.4 mg/kg every 3 weeks; 2.5 mg/kg: ORR 30 (31%); sCR/ CR 3 (3%), VGPR (15%); PD 56 (58%); mPFS 2.9 months; 3.4 mg/kg: ORR 34 (34%); sCR/CR 3 (3%), VGPR 17 (17%); PD 55 (56%); mPFS 4.9 months	Grade 3 and 4 keratopathy (27% in 2.5 mg/kg, 21% in 3.4 mg/kg), thrombocytopenia (20% and 33%) and anemia (20% and 25%)	In latter August 2020, Europe was also on tria
CAR-T: CT053 (NCT03716856, NCT03302403 and NCT03380039) [22]	It includes anti- BCMA human scFv, CD3 ζT cell activation domain, and 4-1BB costimulatory domain	Phase I clinical trial	CT053 was given once after fludarabine/ cyclophosphamide treatment; the total remission rate was 87.5% and the complete remission rate was 70.8%	Hematological toxicity, including a 95.8% decrease in white blood cell count and 66.7% thrombocytopenia. One subject died of BM failure and neutropenia infection. There were 62.5% cytokine release syndrome and 12.5% neurotoxicity	FDA approved two anti-CD19 CAR-T cell products: Yescarta and Kymriah used to treat patients with recurrent and/ or refractory NH [23], Kymriah was initially approved in the USA for relapsed or refractory pediatric or young adult B-cell precursor acute lymphoblastic leukemia in 2017, and subsequently approved for adults with relapsed or refractory diffuse large B-cell lymphoma in early 2018 [24]

AML: acute myeloid leukemia; sCR: stringent complete response; CR: complete response; VGPR: very good partial response; mPFS: median PFS; mDOR: median duration of response; MMAF: monomethyl auristatin F; MC: maleimidoca-proyl; DREAMM: the human study with belantamab mafodotin in patients with relapsed/refractory multiple myeloma; NH: non-Hodgkin lymphoma

### Immune checkpoint

Immune checkpoint inhibitors have been launched in clinical practice to treat patients with several types of cancer, including MM [25]. Targeting the cytotoxic T lymphocyte antigen 4 (CTLA4) or PD-1/PD-L1 pathways checkpoint inhibitors have not been effective as a single agent in MM patients. In addition, current combinations of checkpoint inhibitors with immunomodulatory drugs have yielded disappointing results. However, checkpoint inhibitors remain an attractive therapy for MM [26]. T-cell immunoglobulin and Tim3 is an important negative regulator involved in cellular immunity. It is highly expressed in MM cells. In recent years, it has been found that Tim3 regulates the proliferation of MM cells through the NF- $\kappa$ B signaling pathway [phosphatidylinositol 3-kinase (PI3K), protein kinase B (PKB/AKT), mammalian target of rapamycin (mTOR), and NF- $\kappa$ B [27]. Down-regulation of Tim3 expression can inhibit the proliferation of and induce apoptosis in MM cells and has a cumulative inhibitory effect on the NF- $\kappa$ B signaling pathway in conjunction with bortezomib [28]. In other words, if targeting Tim3 is successful in clinical trials, Tim3-targeted therapy can not only be used as a single therapy for MM but also can be combined with other drugs to increase drug efficacy. Therefore, Tim3 may become a new target for the treatment of NDMM/RRMM.

### **BiTE** immunotherapy

BiTE has the specificity of binding double antigens, including an antibody that can bind to the antigen on T cells and an antibody that can bind to the tumor-specific antigen expressed on malignant cells. In this way, cytolytic synapses can be formed between T and malignant cells to enhance the interaction between T and malignant cells [16]. BiTE can activate and cause expansion of T cells by recognizing the CD3 subunit of T cells, produce cytokines, attack target cells, and finally lead to tumor cell lysis [17]. BiTE has the potential to be initiated immediately. BCMA, also known as tumor necrosis factor receptor (TNFR) superfamily 17 (SF17; TNFRSF17) or CD269, is a member of the TNFR superfamily [29]. It is a highly selective expression in normal plasmablasts (PBs) and plasma cells (PCs) but not in other tissues [30]. Of note, almost all MM cell lines express BCMA, and the expression of malignant PCs was higher than that of normal PCs [31]. BI 836909, a novel BCMA-targeted BiTE for the treatment of MM, induces highly specific and efficacious lysis of MM cells *in vitro* and shows anti-tumor activity *in vivo* (Table 2) [6, 18, 21, 32].

### ADC immunotherapy

ADCs are monoclonal antibodies that bind drugs to tumor-associated antigen (TAA). Through the interaction of antigen and antibody, the drug binds to target cells expressing TAA. ADCs enter cells and release effective drug-induced toxicity to induce DNA damage and cell death. Cleavable junctions are treated by enzymes in target cells, while ADCs with non-cleavable junctions need to degrade the antibody in lysosomes to release the effective drug [19]. ADCs have two main advantages: (1) patients do not need to provide any of their own samples and personalized ADCs can be generated during treatment and (2) ADCs are in line with the requirements of modern medical development of precision medicine. Of course, some ADCs are cleaved outside the cells causing the premature release of drugs before entering the target cells, which will lead to the damage of other normal cells or tissues in the body and may also affect the function of other organs. The use of impermeable cell payloads (such as MMAF) or non-cleavable linkers can reduce the premature release outside the cells [17]. At present, blenrep (belantamab mafodotin-blmf), a BCMA-targeted ADC drug, has been approved for the treatment of highly refractory MM (Table 2) [20, 33, 34].

### CAR-T immunotherapy

CAR-T cells are gene-modified T cells that express CAR specific to TAA. After binding, T cells are activated in a human leukocyte antigen (HLA)-independent manner [22, 24, 35, 36]. These CAR constructs are composed of single-chain antibodies (usually mouse or human) targeting TAA, which are connected to the CD3  $\zeta$  intracellular signal and costimulatory domains through extracellular regions and transmembrane domains.

The significant advantage of CAR-T cells is that these cells work for a long time. A single infusion alone may lead to persistent immunity to cancer cells. Compared with other treatments, the reduction in treatment times greatly reduces the damage of drugs or other substances to the body in the process of treatment. The disadvantage is that this type of drug requires a longer production time because it needs to go through the process of collecting white blood cells from patients for *ex vivo* expansion, transfer of autologous T cells, and reinfusion to patients, which is time-consuming. During long-term T-cell production, patients may develop leukopenia or other diseases that are caused by a decline in defense function. Of course, the immune relationship between reduced immune cells and CAR-T cell infusion may also lead to the progression and deterioration of related diseases. Another potential drawback is the use of pretreated lymphadenectomy. The decrease of endogenous lymphocyte level and the expansion of CAR-T cells may affect the rescue therapy after the failure of CAR-T cell therapy. CT053 is a BCMA-targeted CAR-T cell therapy and has become a breakthrough treatment in MM (Table 2) [23, 37, 38].

### Targeting genomic instability

There are currently several small molecule inhibitors targeting chromosomal instability such as poly(ADP-ribose) polymerase (PARP), Akt, Aurora kinase, and spindle kinase inhibitors which have been tested in mouse models and in early phase I/II trials [39]. Thyroid hormone receptor-interacting protein (Trip) 13 has been identified as a chromosomal instability gene and is associated with drug

resistance, disease recurrence, and adverse prognoses in MM patients [40]. In animal experiments, a small molecule inhibitor (DCZ0415) of Trip13 can inhibit MM cell proliferation. It acts as a toxic agent of the spindle, which causes spindle multipole, destroys the repair of non-homologous end joining (NHEJ), inhibits the synthesis of DNA, and inhibits NF- $\kappa$ B activity in MM cells. More importantly, DCZ0415 works in conjunction with the melphalan and HDAC inhibitor panobinostat to suppress MM cell proliferation [41]. DCZ0415 also inhibits the protective effect of the BM microenvironment on myeloma cells [42]. Therefore, the discovery of more and new targets and new methods by targeting chromosomal instability such as Trip3 for the treatment of MM is important.

ATR inhibition, combined with piperlongumine, triggers synergistic MM cell cytotoxicity by enhancing oxidative stress while concomitantly blocking replicative stress response [43]. This novel combination targeted therapy, that is the synthetic lethal approach, provides an unmet medical need for a poor-prognosis subset of MM patients with extensive chromosomal instability and replicative stress [43].

## Targeting aberrant regulatory network

IRF4 can promote proliferation and survival of both MM stem and tumor cells, and the high level of IRF4 is directly related to the decline of overall survival rate in patients with MM [44]. The ASO of IRF4 inhibits the proliferation and survival of MM cells by reducing the expression of IRF4. IRF4 ASO monotherapy can prevent the formation of a xenograft tumor model and MM cell proliferation and lead to improvement in animal survival rates. In addition, IRF4 ASO presented more obvious advantages for MM treatment than drugs targeting other pathogenic molecules in MM in that it can eliminate the progenitor cells and malignant PCs of MM and retain normal hematopoietic stem cell development [45]. That is, if IRF4 ASO is successful, it may target disease cells more accurately and not cause much damage to other cells. Thus, side effects in MM patients may be greatly reduced. A study has shown that lenalidomide also causes down-regulation of IRF4 expression [46]. However, the use of selective IRF4 ASO may be superior to lenalidomide. ASO can be combined with MM drugs for treatment and has synergistic effects. Therefore, it is possible to study IRF4 ASO since IRF4 represents a potential therapeutic target of MM, especially with respect to the survival of patients who are difficult to treat with a standard of care drugs. Myelocytomatosis oncogene (MYC) is a direct target of IRF4 in activated B cells and myeloma. Critically, IRF4 was itself a direct target of MYC transactivation, generating an autoregulatory circuit in myeloma cells [47]. A novel c-Myc inhibitor, compound 7594-0037, can be used as a novel c-Myc inhibitor and is a potential candidate therapeutic drug for MM [48].

### Epi-therapeutics: a novel treatment modality in MM

Epigenetic aberrations are the pathogenic events that play important roles in leading to MM. HDACs influence the activity of several transcription factors disrupted in MM through histone modifications [49, 50]. Clinical trials have failed to show a clinical benefit with HDAC is as single-agent therapy. ACY-1215, a selective HDAC6 inhibitor, is a promising epigenetic targeted therapy for myeloma that has shown *in vitro* and *in vivo* effectiveness [51] and is now tested in two ongoing phases 1-2 clinical trials (ClinicalTrials.gov. NCT01323751 and NCT01583283) [52].

### **CELMoDs**

CELMoDs are larger molecules and work by quickly degrading a specific set of proteins (called Ikaros and Aiolos) [53-55]. They are potent modulators of the cereblon E3 ligase complex. They also stimulate the immune system and kill myeloma cells directly, even for myeloma that has become resistant to certain therapies. A new type of myeloma treatment called CC-92480 (a CELMoD) showed benefit in phase I international clinical trials [53-55]. It is synergistic with other therapies and has "excellent tumor penetration". The other CELMoD currently in development is iberdomide [53-55].

### **Mutation-targeted therapies**

The mutational spectrum of MM with only a limited number of common recurrent mutations is now observed [56, 57]. Mutations in Kirsten rat sarcoma 2 viral oncogene homolog (KRAS), neuroblastoma RAS viral oncogene homolog (NRAS), and BRAF account for 50% of all cases at presentation and these numbers

increase at relapse MM [58]. Inhibition of MEK, located downstream of the cellular homolog of viral raf gene (RAF) in the mitogen-activated protein kinases (MAPK) pathway, has emerged as a potential strategy to treat these patients [58]. Trametinib shows promise as a myeloma therapeutics based on responses seen in a heavily pretreated population [59]. BRAF-targeting vemurafenib has also been shown to result in responses. Targeting of the rat sarcoma (RAS)/MAPK pathway clearly could work in MM. However, it is compromised by intra-clonal and spatiotemporal heterogeneity. This will require changes to the way response is assessed, with markers directed to the subclone being treated becoming important [58].

## Targeting the microenvironment

BM microenvironment, composed of stromal cells, osteoblasts, osteoclasts, endothelial cells, immune cells, and a non-cellular compartment, provides the supportive role of malignant MM cell differentiation, migration, proliferation, survival, and drug resistance. Therefore, it is very interesting to target the BM microenvironment. Targeting specific niches within BM will have the potential for the treatment in MM and may facilitate earlier intervention to disrupt an environment that is permissive for myeloma progression [60]. PD-1 plays a critical role in the suppressive microenvironment of MM. Immunotherapies using anti-PD-1/PD-L1 strategies are promising treatment options for patients with MM [61]. The MM-niche-derived IL-18 drove the generation of myeloid-derived suppressor cells (MDSCs), leading to accelerated disease progression [62]. A preclinical study suggested that IL-18 could be a potential therapeutic target in MM [62].

# Other targeted therapies based on preclinical data

## CircRNA

Circ\_0007841 has been shown to be up-regulated in BM biopsy cells from MM patients [63]. Circ\_0007841 targets miR-129-5p and inhibit the effect of miR-129-5p, while miR-129-5p can inhibit the expression of jagged canonical Notch ligand 1 (JAG1) mRNA. Thus, circ\_0007841 leads to MM cell proliferation, transformation, and drug resistance by promoting JAG1 overexpression. On the other hand, circ\_0007841 gene knockout can inhibit the proliferation, metastasis, and drug resistance of MM cells by regulating the miR-129-5p/JAG1 axis. Another study found that circ\_0007841 could also directly target miR-338-3p and cause acceleration of the progression of MM [64]. Circ\_0007841 can up-regulate the expression of bromodomain-containing protein 4 (BRD4) by binding with miR-338-3p to promote the activation of PI3K/Akt signaling, thus promoting the proliferation and migration of MM cells. Therefore, circ\_0007841 MM may become an important target for MM treatment in the future.

### NF-ĸB

NF- $\kappa$ B was previously one of the important targets for the treatment of MM. Five members in the NF- $\kappa$ B family have been found: (1) NF- $\kappa$ B1 (p50 and its precursor p105), (2) NF- $\kappa$ B2 (p52 and its precursor p100), (3) RelA (p65), (4) RelB and (5) c-Rel [65].

Two activation pathways of NF-κB have been demonstrated: 1) classical activation pathway, which is the main activation pathway of some receptors and cytokines. The extracellular signal factor binds to the receptor on the cell membrane to activate IkappaB (IκB) kinase (IKK). IKK phosphorylates the serine at the regulatory site of IκB subunit of NF-κB-IκB complex, which facilitates IκB subunit ubiquitination, which is then degraded by a protease and releases an NF-κB dimer. Free NF-κB will enter the nucleus and bind to the genes with NF-κB binding sites to initiate the transcription process and 2) non-classical activation pathway, mainly consisting of the p52/RelB dimer. The non-classical NF-κB kinase of the IKK complex selectively phosphorylates the p52 precursor, p100. Phosphorylation of p100 results in the removal of the C-terminal part of the protein (also known as IκBδ) and its degradation in the proteasome. This process produces a functional p52 protein, which can then dimerize with RelB and accumulate in the nucleus in which regulates its target gene. At least 17% of primary MM cells and 42% of MM cell lines were found to have mutations in several major components of the NF-κB pathway. Although the NF-κB activation pathway is well known and has led to the production of corresponding drugs, the change of new genes and the activation of new sub-pathways in this pathway will become a new target for MM treatment.

### Negative regulators or tumor-suppressors in PBs and PCs

MM characterized by cancerous proliferation of PBs and PCs remains incurable in many patients. Differentially-expressed molecules in MM PCs versus healthy PCs have been explored in order to identify novel targets for treating MM. Some molecules such as BCMA, are highly expressed in cancerous PBs and PCs and have been used as the key targets in the treatment of MM. Apart from positive regulators, negative regulators or tumor-suppressors have also been implicated to be potential biomarkers and therapeutic targets in some cancers [66, 67]. Some tumor suppressors such as down-regulation of candidate 2 (CECR2), contribute to tumor growth in laryngeal squamous cell carcinoma [68]. We searched for novel MM therapeutic targets by comparing mRNA expression patterns between the *Mus* musculus (mouse) myeloma PB-like myeloma SP 2/0 cell line and lipopolysaccharide (LPS)-induced PB/PC. We found that some molecules, such as Gm6377 and BC094916, were decreased in both normal PCs/PBs and SP 2/0 cells [69, 70]. Specifically, some molecules, such as Loc108167440 and Gm40600, were down-regulated in SP 2/0 cells but not in normal PCs and PBs [71, 72]. Of note, the overexpression of reduced forms of molecules, such as Gm6377, BC094916, Loc108167440, and Gm40600 suppressed SP 2/0 cell proliferation and suppressed tumor progression in the SP 2/0 xenograft mouse model [69-72]. These results suggest that modulation of negative regulators or tumor-suppressors may provide a potential therapeutic method for treating MM.

## Future personalized treatment strategies in MM

## Molecular approaches related to diagnosis of MM

Recent advances in the treatment of MM have pointed to accurate diagnosis of MM. It is possible that a better understanding of the genetic and epigenetic interactions in MM may reveal a new understanding of MM pathogenesis, new disease biomarkers, and hopefully the development of novel and a more effective diagnosis method. Present studies are evaluating a combined use of "imaging markers", such as SUVs, MATV, TLG, ADC from positron emission tomography (PET), and magnetic resonance imaging (MRI) techniques respectively, and several "biomarkers" spanning from chemokine immune-modulators, such as PD-1, RANK/RANKL, CXCR4/CXCL12 to transcription factors, such as TP53, RB1, MDM2, runt-related transcription factor (RUNX) family, EZH2, YY1, mitotic arrest deficient 2 (MAD2) [73]. Imaging markers are used to characterize both tumor heterogeneity and metabolic data, whereas biomarkers may improve diagnosis and prognosis leading to precision medicine [73]. Although imaging and molecular integration could allow both early diagnosis and stratification of cancer prognosis, large-scale clinical trials will be necessary to translate pilot studies in the current clinical setting [73].

### **Risk stratification in MM**

The presence of del(17p), t(4;14), t(14;16), t(14;20), gain 1q, or p53 mutation is considered high-risk in MM [74]. The presence of any two high-risk factors is considered double-hit myeloma; three or more high-risk factors are triple-hit myeloma [74]. Standard risk patients can opt for delayed autologous stem cell transplantation (ASCT) at first relapse. Patients not candidates for transplant are treated with lenalidomide (Revlimid) plus dexamethasone (DEX, RD) until progression, or alternatively, a triplet regimen such as VRD for approximately 12-18 months. After ASCT, lenalidomide maintenance is considered for standard-risk patients especially in those who are not in VGPR or better, while maintenance with a bortezomib-based regimen is needed for patients with the intermediate or high-risk disease [75].

### **Precision medicine in MM**

A better understanding of diagnosis and risk stratification in MM will further inspire the development of precision therapy for MM. To date, the choice of therapy for an individual MM patient has been based on clinical factors such as age and comorbidities [76]. The widespread evolution, validation, and clinical utilization of molecular technologies, such as fluorescence *in situ* hybridization and next-generation sequencing has enabled the identification of a number of prognostic and predictive biomarkers for PFS, overall survival, and treatment response [76]. This will improve myeloma patient outcomes incorporating

such biomarkers into the routine diagnostic workup of patients will allow for the use of personalized, biologically-based treatments [76].

# Conclusions

Due to the heterogeneity and resistance of MM patients to a variety of drugs, only a few drugs or limited targeted treatment can produce remission for patients with MM. In addition, the limitation of drug types makes it impossible to effectively treat patients with MM-associated drug resistance. Therefore, the discovery of new targets is undoubtedly the best hope for patients with MM, especially for patients with RRMM. Because any single pathway may have different side effects and the regulation of a single molecule is prone to drug resistance, many single molecule-driven drugs need to be combined with other drugs in order to effectively conduct symptomatic treatment. However, due to cumulative toxic and side effects of multi-drug combination, it is still difficult to achieve precise treatment and targeted therapy. Among these newly discovered targets, BCMA is now the most popular because it targets the PCs of MM and reverses the damaged hematopoietic microenvironment. Moreover, BCMA does not induce drug resistance easily because it plays an important role in the survival of normal long-lived PCs. Importantly, BCMA-based therapy has received good clinical feedback, so it is the most promising new target for the effective treatment of MM. In the near future, we hope that more targets similar to BCMA will become new therapies for the treatment of MM. In the present, incorporating the molecular approaches related to diagnosis and risk stratification into the routine diagnostic workup of patients has been allowing for the use of personalized, biologically-based treatments in MM.

# **Abbreviations**

ADC: antibody-drug conjugates ASO: antisense oligonucleotide BCMA: B-cell maturation antigen BiTE: bispecific T-cell engager BM: bone marrow BRAF: v-raf murine sarcoma viral oncogene homolog B CAR: chimeric antigen receptor CAR-T: chimeric antigen receptor T-cell CELMoDs: cereblon E3 ligase-modulating drugs Circ: circular c-Myc: cellular myelocytomatosis CR: complete response DREAMM: the human study with belantamab mafodotin in patients with relapsed/refractory multiple myeloma D-RVd: daratumumab + lenalidomide + bortezomib + dexamethasone FDA: Food and Drug Administration HDAC: histone deacetylase HR: hazard ratio IKK: IkappaB kinase IL-18: interleukin 18 IRF4: interferon regulatory factor 4 ISA-PD: isatuximab with pomalidomide/dexamethasone IкB: IkappaB MDM2: mouse double minute 2 MM: multiple myeloma

MMAF: monomethyl auristatin F mPFS: median progression-free survival NDMM: newly diagnosed multiple myeloma NF-κB: nuclear factor-kappaB ORR: objective response rate **PBs:** plasmablasts PCs: plasma cells PD: pomalidomide/dexamethasone PD-1: programmed cell death-1 PD-L1: programmed death-ligand 1 PFS: progression-free survival RRMM: relapsed/refractory multiple myeloma RUNX: runt-related transcription factor 1 scFv: single-chain variable fragment sCR: stringent complete response TAA: tumor-associated antigen Tim3: T cell immunoglobulin-3 Trip: thyroid hormone receptor-interacting protein VD: bortezomib + dexamethasone VGPR: very good partial response VRD: bortezomib, lenalidomide, and dexamethasone

# **Declarations**

## **Author contributions**

SZ contributed conception, design of the study, and wrote the first draft of the manuscript. SZ and RW contributed to manuscript revision, read and approved the submitted version.

### **Conflicts of interest**

The authors declare that they have no conflicts of interest.

**Ethical approval** Not applicable.

**Consent to participate** Not applicable.

Consent to publication

Not applicable.

Availability of data and materials

Not applicable.

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# References

- 1. Manier S, Salem KZ, Park J, Landau DA, Getz G, Ghobrial IM. Genomic complexity of multiple myeloma and its clinical implications. Nat Rev Clin Oncol. 2017;14:100-13.
- 2. Attal M, Lauwers-Cances V, Hulin C, Leleu X, Caillot D, Escoffre M, et al; IFM 2009 Study. Lenalidomide, bortezomib, and dexamethasone with transplantation for myeloma. N Engl J Med. 2017;376:1311-20.
- 3. Dimopoulos MA, Dytfeld D, Grosicki S, Moreau P, Takezako N, Hori M, et al. Elotuzumab plus pomalidomide and dexamethasone for multiple myeloma. N Engl J Med. 2018;379:1811-22.
- 4. Feng X, Zhang L, Acharya C, An G, Wen K, Qiu L, et al. Targeting CD38 suppresses induction and function of T regulatory cells to mitigate immunosuppression in multiple myeloma. Clin Cancer Res. 2017;23:4290-300.
- 5. Holstein SA, McCarthy PL. Immunomodulatory drugs in multiple myeloma: mechanisms of action and clinical experience. Drugs. 2017;77:505-20.
- 6. Lonial S, Weiss BM, Usmani SZ, Singhal S, Chari A, Bahlis NJ, et al. Daratumumab monotherapy in patients with treatment-refractory multiple myeloma (SIRIUS): an open-label, randomised, phase 2 trial. Lancet. 2016;387:1551-60.
- 7. Martin T, Strickland S, Glenn M, Charpentier E, Guillemin H, Hsu K, et al. Phase I trial of isatuximab monotherapy in the treatment of refractory multiple myeloma. Blood Cancer J. 2019;9:41.
- 8. Richardson PG, Beksaç M, Špička I, Mikhael J. Isatuximab for the treatment of relapsed/refractory multiple myeloma. Expert Opin Biol Ther. 2020;20:1395-404.
- 9. Richardson PG, Schlossman RL, Alsina M, Weber DM, Coutre SE, Gasparetto C, et al. Panorama 2: panobinostat in combination with bortezomib and dexamethasone in patients with relapsed and bortezomib-refractory myeloma. Blood. 2013;122:2331-7.
- 10. Richter J, Madduri D, Richard S, Chari A. Selinexor in relapsed/refractory multiple myeloma. Ther Adv Hematol. 2020;11:2040620720930629.
- 11. Tzogani K, van Hennik P, Walsh I, De Graeff P, Folin A, Sjöberg J, et al. EMA review of panobinostat (Farydak) for the treatment of adult patients with relapsed and/or refractory multiple myeloma. Oncologist. 2018;23:631-6.
- 12. Usmani SZ, Karanes C, Bensinger WI, D'Souza A, Raje N, Tuchman SA, et al. Final results of a phase 1b study of isatuximab short-duration fixed-volume infusion combination therapy for relapsed/refractory multiple myeloma. Leukemia. 2021;[Epub ahead of print].
- 13. Voorhees PM, Kaufman JL, Laubach J, Sborov DW, Reeves B, Rodriguez C, et al. Daratumumab, lenalidomide, bortezomib, and dexamethasone for transplant-eligible newly diagnosed multiple myeloma: the GRIFFIN trial. Blood. 2020;136:936-45.
- 14. Liu J, Liu W, Mi L, Zeng X, Cai C, Ma J, et al; Union for China Lymphoma Investigators of the Chinese Society of Clinical Oncology; Union for China Leukemia Investigators of the Chinese Society of Clinical Oncology. Incidence and mortality of multiple myeloma in China, 2006-2016: an analysis of the global burden of disease study 2016. J Hematol Oncol. 2019;12:136.
- 15. Liu W, Liu J, Song Y, Wang X, Zhou M, Wang L, et al; Union for China Leukemia Investigators of the Chinese Society of Clinical Oncology, Union for China Lymphoma Investigators of the Chinese Society of Clinical Oncology. Mortality of lymphoma and myeloma in China, 2004-2017: an observational study. J Hematol Oncol. 2019;12:22.
- 16. Huehls AM, Coupet TA, Sentman CL. Bispecific T-cell engagers for cancer immunotherapy. Immunol Cell Biol. 2015;93:290-6.

- 17. Ross SL, Sherman M, McElroy PL, Lofgren JA, Moody G, Baeuerle PA, et al. Bispecific T cell engager (BiTE®) antibody constructs can mediate bystander tumor cell killing. PLoS One. 2017;12:e0183390.
- 18. Hipp S, Deegen P, Wahl J, Blanset D, Thomas O, Rattel B, et al. BI 836909, a novel bispecific T cell engager for the treatment of multiple myeloma induces highly specific and efficacious lysis of multiple myeloma cells *in vitro* and shows anti-tumor activity *in vivo*. Blood. 2015;126:2999.
- 19. Kinneer K, Meekin J, Tiberghien AC, Tai YT, Phipps S, Kiefer CM, et al. SLC46A3 as a potential predictive biomarker for antibody-drug conjugates bearing noncleavable linked maytansinoid and pyrrolobenzodiazepine warheads. Clin Cancer Res. 2018;24:6570-82.
- 20. Lonial S, Lee HC, Badros A, Trudel S, Nooka AK, Chari A, et al. Belantamab mafodotin for relapsed or refractory multiple myeloma (DREAMM-2): a two-arm, randomised, open-label, phase 2 study. Lancet Oncol. 2020;21:207-21.
- 21. Topp MS, Attal M, Langer C, Moreau P, Facon T, Düll J, et al. Phase 1 dose-escalation study of BI 836909, an anti-BCMA bi-specific 1-cell engager, in relapsed andfor refractory multiple myeloma (RRMM). J Clin Oncol. 2016;34:TPS8067.
- 22. Danhof S, Hudecek M, Smith EL. CARs and other T cell therapies for MM: the clinical experience. Best Pract Res Clin Haematol. 2018;31:147-57.
- 23. Hao S, Jin J, Yu K, Li Z, Zhang W, Yang M, et al. CT053, anti-BCMA CAR T-cell therapy for relapsed/ refractory multiple myeloma: proof of concept results from a phase I study. Clin Lymphoma Myeloma Leuk. 2019;19:E54-5.
- 24. Ghosh A, Mailankody S, Giralt SA, Landgren CO, Smith EL, Brentjens RJ. CAR T cell therapy for multiple myeloma: where are we now and where are we headed? Leuk Lymphoma. 2018;59:2056-67.
- 25. Caserta S, Innao V, Musolino C, Allegra A. Immune checkpoint inhibitors in multiple myeloma: a review of the literature. Pathol Res Pract. 2020;216:153114.
- 26. Paul B, Kang S, Zheng Z, Kang Y. The challenges of checkpoint inhibition in the treatment of multiple myeloma. Cell Immunol. 2018;334:87-98.
- 27. Vrábel D, Pour L, Ševčíková S. The impact of NF-κB signaling on pathogenesis and current treatment strategies in multiple myeloma. Blood Rev. 2019;34:56-66.
- 28. Liu Z, Xiang C, Han M, Meng N, Luo J, Fu R. Study on Tim3 regulation of multiple myeloma cell proliferation via NF-κB signal pathways. Front Oncol. 2020;10:584530.
- 29. Sanchez E, Li M, Kitto A, Li J, Wang CS, Kirk DT, et al. Serum B-cell maturation antigen is elevated in multiple myeloma and correlates with disease status and survival. Br J Haematol. 2012;158:727-38.
- 30. Carpenter RO, Evbuomwan MO, Pittaluga S, Rose JJ, Raffeld M, Yang S, et al. B-cell maturation antigen is a promising target for adoptive T-cell therapy of multiple myeloma. Clin Cancer Res. 2013;19:2048-60.
- 31. Eckhert E, Hewitt R, Liedtke M. B-cell maturation antigen directed monoclonal antibody therapies for multiple myeloma. Immunotherapy. 2019;11:801-11.
- 32. Hipp S, Tai YT, Blanset D, Deegen P, Wahl J, Thomas O, et al. A novel BCMA/CD3 bispecific T-cell engager for the treatment of multiple myeloma induces selective lysis *in vitro* and *in vivo*. Leukemia. 2017;31:1743-51.
- 33. Trudel S, Lendvai N, Popat R, Voorhees PM, Reeves B, Libby EN, et al. Targeting B-cell maturation antigen with GSK2857916 antibody-drug conjugate in relapsed or refractory multiple myeloma (BMA117159): a dose escalation and expansion phase 1 trial. Lancet Oncol. 2018;19:1641-53.
- 34. Trudel S, Lendvai N, Popat R, Voorhees PM, Reeves B, Libby EN, et al. Antibody-drug conjugate, GSK2857916, in relapsed/refractory multiple myeloma: an update on safety and efficacy from dose expansion phase I study. Blood Cancer J. 2019;9:37.
- 35. Kriegsmann K, Kriegsmann M, Cremer M, Schmitt M, Dreger P, Goldschmidt H, et al. Cell-based immunotherapy approaches for multiple myeloma. Br J Cancer. 2019;120:38-44.

- 36. Mikkilineni L, Kochenderfer JN. Chimeric antigen receptor T-cell therapies for multiple myeloma. Blood. 2017;130:2594-602.
- 37. Maude SL, Frey N, Shaw PA, Aplenc R, Barrett DM, Bunin NJ, et al. Chimeric antigen receptor T cells for sustained remissions in leukemia. N Engl J Med. 2014;371:1507-17.
- 38. Schuster SJ. CD19-directed CAR T cells gain traction. Lancet Oncol. 2019;20:2-3.
- 39. Beksac M, Balli S, Akcora Yildiz D. Drug targeting of genomic instability in multiple myeloma. Front Genet. 2020;11:228.
- 40. Tao Y, Yang G, Yang H, Song D, Hu L, Xie B, et al. TRIP13 impairs mitotic checkpoint surveillance and is associated with poor prognosis in multiple myeloma. Oncotarget. 2017;8:26718-31.
- 41. Wang Y, Huang J, Li B, Xue H, Tricot G, Hu L, et al. A small-molecule inhibitor targeting TRIP13 suppresses multiple myeloma progression. Cancer Res. 2020;80:536-48.
- 42. Chauhan D, Uchiyama H, Akbarali Y, Urashima M, Yamamoto K, Libermann TA, et al. Multiple myeloma cell adhesion-induced interleukin-6 expression in bone marrow stromal cells involves activation of NF-kappa B. Blood. 1996;87:1104-12.
- 43. Cottini F, Hideshima T, Suzuki R, Tai YT, Bianchini G, Richardson PG, et al. Synthetic lethal approaches exploiting DNA damage in aggressive myeloma. Cancer Discov. 2015;5:972-87.
- 44. Wang L, Yao ZQ, Moorman JP, Xu Y, Ning S. Gene expression profiling identifies IRF4-associated molecular signatures in hematological malignancies. PLoS One. 2014;9:e106788.
- 45. Mondala PK, Vora AA, Zhou T, Lazzari E, Ladel L, Luo X, et al. Selective antisense oligonucleotide inhibition of human IRF4 prevents malignant myeloma regeneration via cell cycle disruption. Cell Stem Cell. 2021;28:623-36.e9.
- 46. Lopez-Girona A, Heintel D, Zhang LH, Mendy D, Gaidarova S, Brady H, et al. Lenalidomide downregulates the cell survival factor, interferon regulatory factor-4, providing a potential mechanistic link for predicting response. Br J Haematol. 2011;154:325-36.
- 47. Shaffer AL, Emre NC, Lamy L, Ngo VN, Wright G, Xiao W, et al. IRF4 addiction in multiple myeloma. Nature. 2008;454:226-31.
- 48. Yao R, Xie Y, Sun X, Zhang M, Zhou J, Liu L, et al. Identification of a novel c-Myc inhibitor 7594-0037 by structure-based virtual screening and investigation of its anti-cancer effect on multiple myeloma. Drug Des Devel Ther. 2020;14:3983-93.
- 49. Todoerti K, Barbui V, Pedrini O, Lionetti M, Fossati G, Mascagni P, et al. Pleiotropic anti-myeloma activity of ITF2357: inhibition of interleukin-6 receptor signaling and repression of miR-19a and miR-19b. Haematologica. 2010;95:260-9.
- 50. Lamottke B, Kaiser M, Mieth M, Heider U, Gao Z, Nikolova Z, et al. The novel, orally bioavailable HSP90 inhibitor NVP-HSP990 induces cell cycle arrest and apoptosis in multiple myeloma cells and acts synergistically with melphalan by increased cleavage of caspases. Eur J Haematol. 2012;88:406-15.
- 51. Santo L, Hideshima T, Kung AL, Tseng JC, Tamang D, Yang M, et al. Preclinical activity, pharmacodynamic, and pharmacokinetic properties of a selective HDAC6 inhibitor, ACY-1215, in combination with bortezomib in multiple myeloma. Blood. 2012;119:2579-89.
- 52. Dimopoulos K, Gimsing P, Grønbæk K. The role of epigenetics in the biology of multiple myeloma. Blood Cancer J. 2014;4:e207.
- 53. Thakurta A, Pierceall WE, Amatangelo MD, Flynt E, Agarwal A. Developing next generation immunomodulatory drugs and their combinations in multiple myeloma. Oncotarget. 2021;12:1555-63.
- 54. You W, Pang J. Pharmacokinetics, bioavailability and metabolism of CC-92480 in rat by liquid chromatography combined with electrospray ionization tandem mass spectrometry. Biomed Chromatogr. 2021;35:e5139.

- 55. Hansen JD, Correa M, Nagy MA, Alexander M, Plantevin V, Grant V, et al. Discovery of CRBN E3 ligase modulator CC-92480 for the treatment of relapsed and refractory multiple myeloma. J Med Chem. 2020;63:6648-76.
- 56. Walker BA, Boyle EM, Wardell CP, Murison A, Begum DB, Dahir NM, et al. Mutational spectrum, copy number changes, and outcome: results of a sequencing study of patients with newly diagnosed myeloma. J Clin Oncol. 2015;33:3911-20.
- 57. Walker BA, Wardell CP, Murison A, Boyle EM, Begum DB, Dahir NM, et al. APOBEC family mutational signatures are associated with poor prognosis translocations in multiple myeloma. Nat Commun. 2015;6:6997.
- 58. Morgan GJ, Jones JR. Integration of genomics into treatment: are we there yet? Am Soc Clin Oncol Educ Book. 2017;37:569-74.
- 59. Heuck CJ, Jethava Y, Khan R, van Rhee F, Zangari M, Chavan S, et al. Inhibiting MEK in MAPK pathway-activated myeloma. Leukemia. 2016;30:976-80.
- 60. Lomas OC, Tahri S, Ghobrial IM. The microenvironment in myeloma. Curr Opin Oncol. 2020;32:170-5.
- 61. Atanackovic D, Luetkens T, Radhakrishnan S, Kroger N. Coinhibitory molecule PD-1 as a therapeutic target in the microenvironment of multiple myeloma. Curr Cancer Drug Targets. 2017;17:839-45.
- 62. Nakamura K, Kassem S, Cleynen A, Chrétien ML, Guillerey C, Putz EM, et al. Dysregulated IL-18 is a key driver of immunosuppression and a possible therapeutic target in the multiple myeloma microenvironment. Cancer Cell. 2018;33:634-48.e5.
- 63. Wang Y, Lin Q, Song C, Ma R, Li X. Depletion of circ\_0007841 inhibits multiple myeloma development and BTZ resistance via miR-129-5p/JAG1 axis. Cell Cycle. 2020;19:3289-302.
- 64. Wang Y, Lin Q, Song C, Ma R, Li X. Circ\_0007841 promotes the progression of multiple myeloma through targeting miR-338-3p/BRD4 signaling cascade. Cancer Cell Int. 2020;20:383.
- 65. Oeckinghaus A, Ghosh S. The NF-kappaB family of transcription factors and its regulation. Cold Spring Harb Perspect Biol. 2009;1:a000034.
- 66. Kapora E, Feng S, Liu W, Sakhautdinova I, Gao B, Tan W. MicroRNA-505-5p functions as a tumor suppressor by targeting cyclin-dependent kinase 5 in cervical cancer. Biosci Rep. 2019;39:BSR20191221.
- 67. Kumar D, Patel SA, Hassan MK, Mohapatra N, Pattanaik N, Dixit M. Reduced IQGAP2 expression promotes EMT and inhibits apoptosis by modulating the MEK-ERK and p38 signaling in breast cancer irrespective of ER status. Cell Death Dis. 2021;12:389.
- 68. Wang X, Xu C, Wang S, Huang W, Liu Y, Zhang X, et al. A novel tumor suppressor CECR2 down regulation links glutamine metabolism contributes tumor growth in laryngeal squamous cell carcinoma. Clin Transl Oncol. 2021;23:1942-54.
- 69. Zhai B, Hou C, Xu R, Fang Y, Ma N, Xing C, et al. Gm6377 suppressed SP 2/0 xenograft tumor by down-regulating Myc transcription. Clin Transl Oncol. 2020;22:1463-71.
- 70. Xu R, Fang Y, Hou C, Zhai B, Jiang Z, Ma N, et al. BC094916 suppressed SP 2/0 xenograft tumor by down-regulating Creb1 and Bcl2 transcription. Cancer Cell Int. 2018;18:138.
- 71. Zhai B, Hou C, Xu R, Fang Y, Xiao H, Chen G, et al. Loc108167440 suppressed myeloma cell growth by P53-mediated apoptosis. Leuk Lymphoma. 2019;60:2541-8.
- 72. Fang Y, Xu R, Zhai B, Hou C, Ma N, Wang L, et al. Gm40600 suppressed SP 2/0 isograft tumor by reducing Blimp1 and Xbp1 proteins. BMC Cancer. 2019;19:700.
- 73. Schiano C, Soricelli A, De Nigris F, Napoli C. New challenges in integrated diagnosis by imaging and osteo-immunology in bone lesions. Expert Rev Clin Immunol. 2019;15:289-301.
- 74. Rajkumar SV. Multiple myeloma: 2020 update on diagnosis, risk-stratification and management. Am J Hematol. 2020;95:548-67.

- 75. Rajkumar SV. Multiple myeloma: 2016 update on diagnosis, risk-stratification, and management. Am J Hematol. 2016;91:719-34.
- 76. Pawlyn C, Davies FE. Toward personalized treatment in multiple myeloma based on molecular characteristics. Blood. 2019;133:660-75.