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Does the gut microbiome environment influence response to systemic breast cancer treatment?

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Abstract

The gut microbiome is a novel player in the pathogenesis and treatment of breast cancer. The term "microbiome" is used to describe the diverse community of micro-organisms existing within the gastrointestinal tract. The gut microbiome plays an important role in oestrogen metabolism through its ability to deconjugate oestrogens within the gut resulting in their reabsorption. Therefore, it is not unsurprising that "dysbiosis", the disruption of normal gut microbiota composition, is now thought to play a role in the development of the disease, as women with breast cancer have been shown to have altered gut microbiota and this has been correlated with tumour characteristics. There is emerging evidence to suggest that the gut microbiota may also impact on breast cancer treatment, by mediating both drug efficacy and toxicity. The present review will discuss the influence of the gut microbiota on systemic treatments for breast cancer, including chemotherapy, anti-human epidermal growth factor receptor 2 (HER2) therapy, endocrine therapy and immunotherapy as well as other targeted treatments.

Keywords

Breast cancer, gut microbiome, gut microbiota, dysbiosis, estrobolome

Introduction

Breast cancer remains the most common cancer in women in the UK, accounting for 15% of all new cancer cases, with an annual incidence of 55,200 [1]. It accounts for the 4th most common cause of cancer mortality in the UK, with 11,400 breast cancer deaths every year [1]. However, due to advances in both diagnosis and treatment, mortality rates over the last decade have steadily declined by 21% in the UK [1].

Well established risk factors for breast cancer, such as nulliparity, use of hormonal therapies and positive family history, are only present in around 50% of those diagnosed [2]. However, no specific risk factors for

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the development of breast cancer are identified in the remaining 50% of patients. Yet a novel player in the pathogenesis and treatment of breast cancer is being recognised—the gut microbiome. The "gut microbiota" within the human gastrointestinal (GI) tract is a diverse community of trillions of microorganisms, amounting to over 1,000 different species of viruses, bacteria, fungi, and archaea [3]. The host and microbiota engage in a symbiotic relationship to maintain normobiosis; however, a state of dysbiosis may promote disease pathogenesis [4, 5].

Literature suggests that a state of dysbiosis, that is the disruption of the gut microbiota, may play a role in the pathogenesis of breast cancer. Postmenopausal women with breast cancer have been shown to possess a gut microbiota of altered composition and reduced diversity when compared to healthy controls [6]. Other studies have also implicated differences in microbiota of women with breast cancer compared to controls. Luu et al. [7], for example, showed that intestinal microbiota differs according to tumour characteristics as well as body mass index (BMI). Specifically, patients with histological Grade 3 breast tumours had increased numbers of *Blautia* species within the gut when compared to patients with Grade 1 cancers [7].

When considering factors that influence the gut microbiota, method of delivery at birth, infant feeding experience, genetics, infections, medications, diet and smoking are at the forefront [8]. Indeed, diet, for example, has been shown to rapidly and reproducibly alter the composition of human gut microbiota [9]. The link between diet and obesity has also been demonstrated to be mediated by the gut microbiota, rather than exclusively driven by high calorie diet [8]. This highlights the importance of diet and lifestyle factors in the make-up of the gut microbiota. These can be easily overlooked, but have potential to be used as a treatment target for patients with breast cancer. Furthermore, it is currently unknown whether cancer pathology leads to alteration in the gut microbiota or whether dysbiosis in itself is the carcinogenic factor.

There is emerging evidence to suggest that dysbiosis may not only play a role in the pathogenesis of breast cancer, but may also influence response to systemic treatment of the disease. Microbiota interacts with breast cancer treatments in a plethora of ways, which can influence the degree of efficacy of chemotherapeutic drugs and immune therapies as well as the side effects from such therapies [10]. We present a review of the current evidence for the role of the gut microbiome in influencing systemic breast cancer therapy, specifically with reference to the bacterial component of the microbiota.

Chemotherapy

The gut microbiota is thought to modulate chemotherapeutic agents through a number of key mechanisms, namely translocation, immunomodulation, metabolism, enzymatic degradation and reduced diversity and ecological variation (known as the TIMER framework) [11]. The impact that these processes have on host response to chemotherapeutic drugs are grouped into three categories: facilitation of drug efficacy, compromise of anti-cancer effects and/or mediation of toxicity [11].

Translocation, the process through which bacteria pass across the gut barrier into the systemic circulation, allows species of the gut microbiota to possess the potential to contribute to chemotherapy's morbidity and also improve its' efficacy [12]. Literature in this field relates to the study of haematological malignancy rather than malignant disease of the breast. However, in mice models, both doxorubicin and cyclophosphamide have been shown to induce the translocation of selected Gram-positive bacteria into secondary lymphoid organs [13]. Once translocation has occurred, a specific subset of T helper 17 cells and memory T helper 1 cells are generated, which contribute to the aforementioned chemotherapy agents' efficacy [13]. Conversely, evidence exists to support sterile mice or mice which have been treated with antibiotics to reduce the diversity of the microbiota had tumours which were resistant to the effects of chemotherapy [13].

With regard to immunomodulation, Iida et al. [14] showed that commensal bacteria also impact response to chemotherapy by modulating the microenvironment of subcutaneous tumours. In sterile mice or those treated with antibiotics, tumour-infiltrating myeloid-derived cells had a poor response to platinum chemotherapy resulting from poor cytokine production [14]. However, those mice treated with CpG-oligonucleotides (CpG-ODN) displayed tumour necrosis with reduced production of reactive oxygen species and less cytotoxicity [14].

The gut microbiota not only impacts the therapeutic efficacy of chemotherapy, but also is thought to have an impact on its toxic effects. Mucositis, often manifesting itself as diarrhoea, is a common adverse effect of chemotherapy, which causes morbidity and mortality [15]. As a result of systemic chemotherapy, the gut microbiota demonstrates reduced diversity and ecological variation patterns [11]. Namely a reduction in gut villous length and a reduction in diversity of the microbiota have been shown [16]. Patients receiving chemotherapy for haematological malignancy is thought to have reduced levels of *Bifidobacterium, Clostridium* cluster XIV5, *Faecalibacterium prausnitzii* within the GI tract, and increased levels of *Enterobacteriaceae* and *Bacteroides* [16]. The increased presence of these species may contribute to the development of mucositis, weight loss and bacteraemia [16], through metabolism and enzymatic degradation of the chemotherapeutic agents is yet to be examined. Randomised controlled trials have been performed to investigate the efficacy of probiotics in preventing cancer therapy-induced mucositis with 3 out of 6 trials showing a significant reduction in the incidence of diarrhea [16].

In terms of emergent interventions currently available, most are aimed at reducing toxicity from chemotherapy rather than improving efficacy. A study in mice demonstrated that a diet high in protein, *L*-leucine, fish oil and specific oligosaccharides can reduce the incidence and severity of *Pseudomonas* translocation from cyclophosphamide-induced immunosuppression, for example [17]. 5-fluorouracil (5-FU) has been shown to have an enhanced effect in the presence of ginseng in colorectal cancer treatment; however, it is not known whether the same can be said for breast cancer [18]. Polysaccharides from the ink of squid have been shown to enrich the levels of bifidobacteria and reduce levels of *Bacteroidetes*, again being a potential treatment option to reduce adverse effects of chemotherapy [19].

Probiotics, prebiotics and synbiotics are another treatment strategy to reduce the risk of mucositis during chemotherapy [20]. Meanwhile, prebiotics, namely oligofructose and inulin, have been shown to both inhibit malignant tumour growth and potentiate the effects of six different chemotherapeutic agents in animal models [21].

Anti-human epidermal growth factors receptor 2 therapy

There is strong evidence that the composition of the gut microbiota can also potentiate the effects or reduce the efficacy of anti-human epidermal growth factors receptor 2 (HER2) therapies in breast cancer. Di Modica et al. [22] studied the relationship between commensal bacteria composition and clinical efficacy of trastuzumab in HER2 positive patients, finding that administration of antibiotics impaired the efficacy of anti-HER2 therapy. The effect was achieved through reducing the CD4⁺ T cells' and natural killer (NK) cells' recruitment due to reduced bacterial biodiversity in the gut; mice transplanted with faeces of antibiotic-treated mice also did not benefit from anti-HER2 treatment, thus supporting the initial findings [22]. Mice who achieved a complete pathological tumour response to anti-HER2 therapy had a significantly higher microbial diversity in the gut and non-responders had reduced Clostridiales and increased *Bacteroidales* species [22]. These represent potential treatment strategies to enhance the therapeutic efficacy of trastuzumab (through manipulation of intestinal flora).

Another study in patients found two microbiota clusters, namely Clostridiales and *Bacteroides*, which discriminated patients according to whether they had a complete response to anti-HER2 therapy [23]. Responders were found to have a higher abundance of Clostridiales and lower abundance of *Bacteroides*, supporting the evidence provided by the study discussed above [22, 23]. Furthermore, in the same study, intestinal flora-depleted mice were transplanted with faecal material from responders and non-responders patients. The mice's response to trastuzumab was identical to that of patients from which stool was obtained [23]. This suggests a causal relationship between gut microbiota and response to anti-HER2 therapies, independent of tumour molecular characteristics. Manipulating the gut microbiota could thus increase the efficacy of trastuzumab, perhaps through transplantation of faecal material as done in the study discussed above. The gut microbiota could also be used as a marker of treatment response, through measurement of the different species of bacteria within it.

Endocrine therapy

It is well known that the gut microbiota plays a role in oestrogen metabolism [24]. The collective numbers of bacterial genes in the GI tract that are capable of metabolizing oestrogens have been named the "Estrobolome" [25]. This "aggregate of enteric bacterial genes" has been considered as a potential factor which can modulate risk of oestrogen receptor positive breast cancer.

Endogenous oestrogen is found in a number of forms: oestradiol (the predominant form in premenopausal women); oestrone (predominant in post-menopausal women) an oestriol (predominant in gravid women). Collectively, these oestrogen hormones are produced predominantly within the ovaries, adrenals, adipose tissue and during their circulation they are subject to conjugation by first-pass hepatic metabolism. The conjugated oestrogens are then excreted renally or via bile into faeces. Conjugated oestrogens which are excreted into bile are subjected to the "estrobolome". Unconjugated oestrogens within the GI tract can be reabsorbed into the circulation, and it is known that bacterial species can deconjugate oestrogens previously conjugated by first pass metabolism, leading to their reabsorption (Figure 1).



Endogenous Oestrogen Production

Figure 1. Enterohepatic circulation of oestrogens and effect of the "Estrobolome"

Particular bacteria have been identified to have the ability to deconjugate oestrogens via hydrolytic enzymes beta-glucuronidases and beta-glucosidases [26]. In deconjugating the oestrogens, their reuptake is promoted and therefore it is speculated that the composition of the gut microbiota can influence the risk of breast cancer (Figure 1). Women with lower plasma oestrogen concentrations have been found to have reduced faecal glucoronidase activity [27]. In the mouse model, several bacterial taxa with beta-glucoronidase activity have been shown to change upon administration of conjugated oestrogens and bazedoxifene, a common treatment for peri-menopausal symptoms [28]. In this study, Chen et al. [28] identified that faecal glucoronidase activity decreased after treatment with conjugated oestrogens and bazedoxifene. This is the first published evidence to support the ability of oestrogen therapy to modulate the gut microbiota. In relation to the treatment of breast cancer, the authors suggest that probiotic supplementation to modulate gut microbial activity may present an opportunity to alter the half-life or serum concentration

of tamoxifen (TAM), a selective oestrogen receptor modulator, in order to optimise its therapeutic effect. Indeed, lower serum concentrations of the active TAM metabolite have been associated with poorer outcome in the disease [29]. However, any specific positive or negative influence of the gut microbiota on the hormonal treatment of breast cancer remains unknown.

Immunotherapy

There is increasing evidence to suggest that the degree of clinical response to immune checkpoint inhibitors is shaped by the characteristics of the gut microbiota. The two most widely recognised immune checkpoint inhibitors are cytotoxic T lymphocyte-associated antigen 4 (CTLA-4) and programmed death-ligand 1/ programmed cell death protein-1 (PD-L1/PD-1). These are emerging therapies in the treatment of breast cancer, however, evidence for the impact of the gut microbiota on these therapies exists largely in the context of metastatic melanoma [30]. Ipilimumab is a fully human monoclonal antibody directed against CTLA-4, a major negative regulator of T cell activation which improved overall survival of patients with metastatic melanoma. Anti-tumour effects of CTLA-4 blockade have been found to depend on distinct *Bacteriodes* species both in mice and human models, particularly B. *thetaiotaomicron* and B. *fragilis* being associated with efficacious CTLA-4 blockade [31].

A number of studies into anti-PD-1 and the relationship with malignant melanoma have found that treatment responders harbour more diverse gut microbiota and increased anti-tumour T cell responses compared to non-responders [32-34]. It is thought that the microbiota of immunotherapy responders may upregulate the immune response through enhancing antigen presentation, or increasing T cell recruitment in the local tumour environment. *Bifidobacterium* and *Akkermansia muciniphila* have been some of a number of specific bacterial species correlated with positive response to anti-PD-L1 immunotherapy [35].

Other targeted therapies

Study into the impact of the gut microbiota on targeted therapies for breast cancer is in its infancy, and the gut microbiota itself has been identified as a therapeutic target. Commensal dysbiosis, the disruption of the normal gut microbiota, has been identified as a potential biomarker for high-risk breast cancer or as a potential therapeutic target in reducing tumour-promoting inflammation and subsequent metastatic dissemination [36].

Using a mouse model of hormone receptor positive mammary cancer, Buchta Rosean et al. [36] identified commensal dysbiosis as a host-intrinsic regulator of tissue inflammation, fibrosis and tumour dissemination which all are known contributors to poor survival outcomes in hormone receptor positive breast cancer. In this study, two mammary cancer cell lines were injected orthotopically into the abdominal mammary fat pad. The first used was BRPKp110 [37], a cell line for poorly metastatic hormone receptor positive mammary cancer. The second was a highly metastatic cell line, polyomavirus middle T (PyMT) [38]. Prior to this, gut dysbiosis was induced within the mouse models using an antibiotic cocktail which resulted in a significant reduction in community richness within the gut.

After tumour cell implantation, dysbiotic mice had a significant increase in total accumulated myeloid cells (including macrophages) within the normal mammary tissue adjacent to tumour. Flow cytometry was also used to evaluate for tumour cell dissemination in the poorly metastatic mouse model identifying significantly more disseminated tumour cells within axillary lymph node and lung tissue. Although it remains unknown whether host-intrinsic differences in immune function promote early dissemination of tumour cells, lymph node involvement is well known to be one of the most reliable predictors of recurrent disease and hence is often used as a marker to necessitate an upstaging of treatment [39]. Hence, future study in this field is likely to translate this work into the clinical setting with the view to using gut dysbiosis as a biomarker or therapeutic target.

Current trials/trial evidence

A number of active clinical trials and observational studies are currently ongoing to investigate the impact of the gut microbiota on breast cancer treatments. One study, for example, is exploring the ability of gut microbiota composition to predict progression-free survival and whether there is any correlation with response to treatment [40]. Similarly, another study is investigating whether the gut microbial composition can influence immune response to the tumour, resulting in individual differences in response to anti-cancer therapies [41]. Multiple other studies are also looking at the gut microbiota composition in relation to the treatment of breast cancer. These studies are summarized in Table 1.

Table 1. Summary of ongoing clinical trials and observational studies in breast cancer treatment in relation	to gut microbiota
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Study	Aims	Study type	Sample size
"Engineering gut microbiome to target breast cancer" [42]	To determine whether probiotics will help the body's immune system react to breast cancer	Clinical trial	7
"Gut microbiome and GI toxicities as determinants of response to neoadjuvant chemo for advanced breast cancer" [41]	To determine whether gut microbial composition can influence immune response to the tumour, resulting in inter-individual differences in the response to anti-cancer therapies	Clinical trial	40
"Intestinal microbiota of breast cancer patients undergoing chemotherapy" [43]	To study specific types (or groups) of bacteria that promote the occurrence, development, and mechanism of breast cancer	Observational	80
"Breast cancer and its relationship with the microbiota" [44]	To examine associated between composition and functionality of the mammary/gut microbiota with breast cancer risk, and to determine if exposure to environmental contaminants [endocrine disruptors, endocrine disrupting chemicals (EDCs)] can alter the microbiota?	Observational	200
"Determinants of acquired endocrine resistance in metastatic breast cancer: a pilot study" [45]	To identify markers of endocrine resistance in ctDNA and the gut microbiome in patients with estrogen (Oestrogen) receptor positive (ER ⁺) HER2 ⁻ metastatic breast cancer	Observational	20
"Study to investigate efficacy of a novel probiotic on the bacteriome and mycobiome of breast cancer" [46]	To examine the effect of probiotics on the breast tumour microbiome and gut microbiome in breast cancer	Clinical trial	100
"Effects of chemotherapy on intestinal bacteria in patients with newly diagnosed breast cancer" [47]	To study effects of chemotherapy on intestinal bacteria/ organisms in patients newly diagnosed with breast cancer	Observational	36
"ARGONAUT: stool and blood sample bank for cancer patients" [40]	To determine whether the microbiota composition can predict progression-free survival.	Observational	4,000
	Whole genome sequencing and metabolomics will be used to characterize the patient's microbiome, and whether there is any correlation with response to treatment		
"Gut and intratumoural microbiome effect on the neoadjuvant chemotherapy-induced immunosurveillance in triple negative breast cancer" [48]	To determine if the probability of pathologic complete response in triple negative breast cancer patients treated with standard of care neoadjuvant chemotherapy is correlated with variability in the composition of intestinal and intra-tumoral microbiota and subsequent short-term alterations in composition	Observational	49
"The clinical study of modern therapies on flora in body fluids and blood of malignant tumour patients" [49]	To study what modern therapies lead to the influence of microecological environment including diversity and abundance of bacteria in patients who received malignant tumours	Observational	500

Future perspective

Genetic profiling and gene-environment interactions are potential future players which can be used to target treatment in breast cancer. An interaction between CASP8-rs1045485 and alcohol consumption has been replicated in the pathogenesis of breast cancer, however, several others have not, including LSP1-rs3817198 and parity and 1p11.2-rs11249433 and ever being parous [50]. Additionally, it is not known what effect these interactions have on the gut microbiota itself.

The gut microbiota is influenced by host genetics, with the *Christensenellaceae* family of bacteria identified as the most heritable taxon [51]. This family has been shown to co-occur with other heritable bacteria and archaea. Additionally, *Christensenellaceae* and its co-inhabitants are known to be enriched in subjects with low BMI and when transplanted to germfree mice they demonstrate reduced weight gain [51]. These findings indicate a direct relationship between genetics and gut microbiota, which can potentially be targeted for breast cancer treatment.

One study has identified germline genetic polymorphisms across 24 different cancer types, which are associated with variable tissue gene expression [52]. Expression of inducible T-cell co-stimulator ligand (*ICOSLG*) and endoplasmic reticulum aminopeptidase 2 (*ERAP2*) (major regulators of immunity) were shown to be under strong genetic control and germline variants associated with a strong immune response were also defined [52]. This demonstrates that germline genetics can potentially be used to tailor immunotherapy in breast cancer patients based on the germline phenotype.

Genetic and environmental factors no doubt play a major role in the make-up of the immune system and the gut microbiota, however, specific gene-environment interactions in relation of these to breast cancer are not yet known.

As previously discussed, nutrition, lifestyle, diet, microbiome, and environment are some of the exogenous factors that influence the genome and metabolome of neoplastic and immune cells. The field of molecular pathological epidemiology (MPE) is a promising direction in investigating the role of exogenous factors in relation to the treatment of breast cancer and can provide insights into environment-tumor-host interactions. This is done by integrating pathological principles into data science fields, including bioinformatics, biostatistics, and epidemiology [53]. Novel players which aim to transform the field of MPE include artificial intelligence, digital pathology, systems biology and *in vivo* pathology techniques as we continue to advance the field of precision medicine [53]. This transdisciplinary field can contribute to improved understanding of the interactive role of immune cells, tumor cells, microbiome, and environment.

This integrative science approach can be used to investigate a variety of neoplastic and non-neoplastic diseases, and it can enhance causal inference and technological advances continuously enable more complex analyses [54]. The drawbacks include sample size limitations, requirement for interdisciplinary experts, standardized guidelines and education programs and need for rigorous validation of findings [54].

Conclusions

A vast number of potential connections exist between the gut microbiota and malignant breast disease. However, investigation into the specific interactions is in its infancy and there is still much to understand about the effect of the gut microbiota on systemic breast cancer treatments. Our literature review highlights the role of the microbiota in influencing response to chemotherapeutic drugs and their side effects. The gut microbiota has the capacity to metabolize systemic treatments in cancer and modulate the immune system in its response to treatment. Future work will expand the evidence base with specific regard to breast cancer, and further explore the potential of gut dysbiosis to act as a biomarker and therapeutic target itself.

Abbreviations

CTLA-4: cytotoxic T lymphocyte-associated antigen 4 GI: gastrointestinal HER2: human epidermal growth factor receptor 2

Declarations

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Author contributions

EB, SM, GU and BE contributed to conception and design of the review. EB and SM wrote the first draft of the manuscript. All authors contributed to manuscript revision, read, and approved the submitted version. First authorship is shared between SM and EB as both authors equally contributed and worked on the manuscript.

Conflicts of interest

The authors declare that they have no conflicts of interest.

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References

- 1. Cancerresearchuk.org [Internet]. London: Cancer Research UK; c2020 [cited 2020 Mar]. Available from: https://www.cancerresearchuk.org/health-professional/cancer-statistics/statistics-by-cancer-type/ breast-cancer
- 2. Lacey JV Jr, Kreimer AR, Buys SS, Marcus PM, Chang SC, Leitzmann MF, et al. Breast cancer epidemiology according to recognized breast cancer risk factors in the Prostate, Lung, Colorectal and Ovarian (PLCO) Cancer Screening Trial Cohort. BMC Cancer. 2009;9:84.
- 3. Selber-Hnatiw S, Rukundo B, Ahmadi M, Akoubi H, Al-Bizri H, Aliu AF, et al. Human gut microbiota: toward an ecology of disease. Front Microbiol. 2017;8:1265.
- 4. Fernández MF, Reina-Pérez I, Astorga JM, Rodríguez-Carrillo A, Plaza-Díaz J, Fontana L. Breast cancer and its relationship with the microbiota. Int J Environ Res Public Health. 2018;15:1747.
- 5. Schwabe RF, Jobin C. The microbiome and cancer. Nat Rev Cancer. 2013;13:800-12.
- 6. Goedert JJ, Jones G, Hua X, Xu X, Yu G, Flores R, et al. Investigation of the association between the fecal microbiota and breast cancer in postmenopausal women: a population-based case-control pilot study. J Natl Cancer Inst. 2015;107:djv147.
- 7. Luu TH, Michel C, Bard JM, Dravet F, Nazih H, Bobin-Dubigeon C. Intestinal proportion of *Blautia* sp. is associated with clinical stage and histoprognostic grade in patients with early-stage breast cancer. Nutr Cancer. 2017;69:267-75.
- 8. Wen L, Duffy A. Factors influencing the gut microbiota, inflammation, and type 2 diabetes. J Nutr. 2017;147:1468S-75S.
- 9. David LA, Maurice CF, Carmody RN, Gootenberg DB, Button JE, Wolfe BE, et al. Diet rapidly and reproducibly alters the human gut microbiome. Nature. 2013;505:559-63.
- 10. Saad, R, Rizkallah MR, Aziz RK. Gut pharmacomicrobiomics: the tip of an iceberg of complex interactions between drugs and gut-associated microbes. Gut Pathog. 2012;4:16.
- 11. Alexander JL, Wilson ID, Teare J, Marchesi JR, Nicholson JK, Kinross JM. Gut microbiota modulation of chemotherapy efficacy and toxicity. Nat Rev Gastroenterol Hepatol. 2017;14:356-65.
- 12. Samet A, Sledzińska A, Krawczyk B, Hellmann A, Nowicki A, Kur J, et al. Leukemia and risk of recurrent *Escherichia coli* bacteremia: genotyping implicates *E. coli* translocation from the colon to the bloodstream. Eur J Clin Microbiol Infect Dis. 2013;32:1393-400.

- 13. Viaud S, Saccheri F, Mignot G, Yamazaki T, Daillère R, Hannani D, et al. The intestinal microbiota modulates the anticancer immune effects of cyclophosphamide. Science. 2013;342:971-6.
- 14. Iida N, Dzutsev A, Stewart CA, Smith L, Bouladoux N, Weingarten RA, et al. Commensal bacteria control cancer response to therapy by modulating the tumor microenvironment. Science. 2013;342:967-70.
- 15. Paci A, VealG, Bardin C, LevêqueD, Widmer N, Beijnen J, et al. Review of therapeutic drug monitoring of anticancer drugs part 1--cytotoxics. Eur J Cancer. 2014;50:2010-9.
- 16. Montassier E, Batard E, Massart S, Gastinne T, Carton T, Caillon J, et al. 16S rRNA gene pyrosequencing reveals shift in patient faecal microbiota during high-dose chemotherapy as conditioning regimen for bone marrow transplantation. Microb Ecol. 2014;67:690-9.
- 17. Faber J, van Limpt K, Diane Kegler, Luiking Y, Garssen J, van Helvoort A, et al. Bacterial translocation is reduced by a specific nutritional combination in mice with chemotherapy-induced neutropenia. J Nutr. 2011;141:1292-8.
- 18. Li XL, Wang CZ, Mehendale SR, Sun S, Wang Q, Yuan CS, et al. Panaxadiol, a purified ginseng component, enhances the anti-cancer effects of 5-fluorouracil in human colorectal cancer cells. Cancer Chemother Pharmacol. 2009;64:1097-104.
- 19. Tang Q, Zuo T, Lu S, Wu J, Wang J, Zheng R, et al. Dietary squid ink polysaccharides ameliorated the intestinal microflora dysfunction in mice undergoing chemotherapy. Food Funct. 2014;5:2529-35.
- 20. Redman MG, Ward EJ, Phillips RS. The efficacy and safety of probiotics in people with cancer: a systematic review. Ann Oncol. 2014;25:1919-29.
- 21. Taper HS, Roberfroid MB. Nontoxic potentiation of cancer chemotherapy by dietary oligofructose or inulin. Nutr Cancer. 2000;38:1-5.
- 22. Di Modica M, Regondi V, Gargari G, Bonizzi A, Arioli S, Belmonte B, et al. Abstract 4959: the gut microbiota contributes to the effectiveness of HER2-targeted therapy. Immunology. 2019;79:4959.
- 23. Tagliabue E, Modica M, Gargari G, Regondi V, Bonizzi A, Arioli S, et al. Abstract P4-10-32: commensal gut microbiota influences efficacy of trastuzumab in patients with HER2-positive breast carcinoma. Cancer Res. 2020;80:4-10-32.
- 24. Parida S, Sharma D. The microbiome-estrogen connection and breast cancer risk. Cells. 2019;8:1642.
- 25. Plottel CS, Blaser MJ. Microbiome and malignancy. Cell Host Microbe. 2011;10:324-35.
- 26. Kwa M, Plottel CS, Blaser MJ, Adams S. The intestinal microbiome and estrogen receptor-positive female breast cancer. J Natl Cancer Inst. 2016;108:djw029.
- 27. Goldin BR, Adlercreutz H, Gorbach SL, Warram JH, Dwyer JT, Swenson L, et al. Estrogen excretion patterns and plasma levels in vegetarian and omnivorous women. N Engl J Med. 1982;307:1542-7.
- 28. Chen KLA, Liu X, Zhao YC, Hieronymi K, Rossi G, Auvil LS, et al. Long-term administration of conjugated estrogen and bazedoxifene decreased murine fecal β-glucuronidase activity without impacting overall microbiome community. Sci Rep. 2018;8:8166.
- 29. Helland T, Henne N, Bifulco E, Naume B, Borgen E, Kristensen VN, et al. Serum concentrations of active tamoxifen metabolites predict long-term survival in adjuvantly treated breast cancer patients. Breast Cancer Res. 2017;19:125.
- 30. Voutsadakis IA. Immune blockade inhibition in breast cancer. Anticancer Res. 2016;36:5607-22.
- 31. Vétizou M, Pitt JM, Daillère R, Lepage P, Waldschmitt N, Flament F, et al. Anticancer immunotherapy by CTLA-4 blockade relies on the gut microbiota. Science. 2015;350:1079-84.
- 32. Roy S, Trinchieri G. Microbiota: a key orchestrator of cancer therapy. Nat Rev Cancer. 2017;17:271-85.
- 33. Routy B, Le Chatelier E, Derosa L, Duong CPM, Alou MT, Daillère R, et al. Gut microbiome influences efficacy of PD-1-based immunotherapy against epithelial tumors. Science. 2018;359:91-7.
- 34. Matson V, Fessler J, Bao R, Chongsuwat T, Zha Y, Alegre ML, et al. The commensal microbiome is associated with anti-PD-1 efficacy in metastatic melanoma patients. Science. 2018;359:104-8.

- 35. Sivan A, Corrales L, Hubert N, Williams JB, Aquino-Michaels K, Earley ZM, et al. Commensal *Bifidobacterium* promotes antitumor immunity and facilitates anti-PD-L1 efficacy. Science. 2015;350:1084-9.
- 36. Buchta Rosean C, Bostic RR, Ferey JCM, Feng TY, Azar FN, Tung KS, et al. Preexisting commensal dysbiosis is a host-intrinsic regulator of tissue inflammation and tumor cell dissemination in hormone receptor-positive breast cancer. Cancer Res. 2019;79:3662-75.
- 37. Allegrezza MJ, Rutkowski MR, Stephen TL, Svoronos N, Perales-Puchalt A, Nguyen JM, et al. Trametinib drives T-cell-dependent control of KRAS-mutated tumors by inhibiting pathological myelopoiesis. Cancer Res. 2016;76:6253-65.
- 38. Bos PD, Plitas G, Rudra D, Lee SY, Rudensky AY. Transient regulatory T cell ablation deters oncogenedriven breast cancer and enhances radiotherapy. J Exp Med. 2013;210:2435-66.
- 39. Yamashita H, Ogiya A, Shien T, Horimoto Y, Masuda N, Inao T, et al. Clinicopathological factors predicting early and late distant recurrence in estrogen receptor-positive, HER2-negative breast cancer. Breast Cancer. 2016;23:830-43.
- 40. Persephone Biosciences. ARGONAUT: Stool and blood sample bank for cancer patients [Internet]. ClinicalTrials.gov: National Library of Medicine (US); c2020 [updated 2021 Feb 23; cited 2021 Apr 9]. Available from: https://www.clinicaltrials.gov/ct2/show/ NCT04638751?term=gut+microbiome&cond=Breast+Cancer&draw=3&rank=13
- University of Arkansas. Gut microbiome & gastrointestinal toxicities as determinants of response to neoadjuvant chemo for advanced breast cancer [Internet]. ClinicalTrials.gov: National Library of Medicine (US); c2016 [updated 2021 Jun 15; cited 2020 Apr 9]. Available from: https://www.clinicaltrials.gov/ ct2/show/NCT02696759?term=gut+microbiome&cond=Breast+Cancer&draw=4&rank=3
- 42. Mayo Clinic. Engineering gut microbiome to target breast cancer [Internet]. ClinicalTrials.gov: National Library of Medicine (US); c2017 [updated 2021 May 21; cited 2021 Apr 9]. Available from: https://www.clinicaltrials.gov/ct2/show/NCT03358511?term=gut+microbiome&cond=Breast+Cancer&draw=2
- 43. First Affiliated Hospital of Harbin Medical University. Intestinal microbiota of breast cancer patients undergoing chemotherapy [Internet]. ClinicalTrials.gov: National Library of Medicine (US); c2019 [updated 2019 Oct 25; cited 2021 Apr 9]. Available from: https://www.clinicaltrials.gov/ct2/show/ NCT04138979?term=gut+microbiome&cond=Breast+Cancer&draw=2&rank=4
- 44. Universidad de Granada. Breast cancer and its relationship with the microbiota [Internet]. ClinicalTrials.gov: National Library of Medicine (US); c2019 [updated 2021 May 13; cited 2021 Apr 9]. Available from: https://www.clinicaltrials.gov/ct2/show/NCT03885648?term=gut+microbiome&cond=Breast+Cancer&draw=2&rank=5
- 45. Sunnybrook Health Sciences Centre. Determinants of acquired endocrine resistance in metastatic breast cancer: a pilot study [Internet]. ClinicalTrials.gov: National Library of Medicine (US); c2020 [updated 2021 Oct 19; cited 2021 Apr 9]. Available from: https://www.clinicaltrials.gov/ct2/show/ NCT04579484?term=gut+microbiome&cond=Breast+Cancer&draw=2&rank=6
- 46. Case Comprehensive Cancer Center. Study to investigate efficacy of a novel probiotic on the bacteriome and mycobiome of breast cancer [Internet]. ClinicalTrials.gov: National Library of Medicine (US); c2020 [updated 2021 May 7; cited 2021 Apr 9]. Available from: https://www.clinicaltrials.gov/ct2/show/ NCT04362826?term=gut+microbiome&cond=Breast+Cancer&draw=2&rank=9
- 47. University of Southern California. Effects of chemotherapy on intestinal bacteria in patients with newly diagnosed breast cancer [Internet]. ClinicalTrials.gov: National Library of Medicine (US); c2015 [updated 2019 Jun 18; cited 2021 Apr 9]. Available from: https://www.clinicaltrials.gov/ct2/show/ NCT02370277?term=gut+microbiome&cond=Breast+Cancer&draw=2&rank=10
- Health. Gut 48. Hackensack Meridian and intratumoral microbiome effect on the neoadjuvant chemotherapy-induced immunosurveillance in triple negative breast cancer [Internet]. ClinicalTrials.gov: National Library of Medicine (US); c2018 [updated

2021 Jul 2; cited 2021 Apr 9]. Available from: https://www.clinicaltrials.gov/ct2/show/ NCT03586297?term=gut+microbiome&cond=Breast+Cancer&draw=3&rank=16

- 49. The First Affiliated Hospital of Dalian Medical University. The clinical study of modern therapies on flora in body fluids and blood of malignant tumor patients [Internet]. ClinicalTrials.gov: National Library of Medicine (US); c2018 [updated 2019 Dec 18; cited 2021 Apr 9]. Available from: https://www.clinicaltrials. gov/ct2/show/NCT04202848?term=gut+microbiome&cond=Breast+Cancer&draw=4&rank=27
- 50. Fletcher O, Dudbridge F. Candidate gene-environment interactions in breast cancer. BMC Med. 2014;12:195.
- 51. Goodrich JK, Waters JL, Poole AC, Sutter JL, Koren O, Blekhman R, et al. Human genetics shape the gut microbiome. Cell. 2014;159:789-99.
- 52. Lim YW, Chen-Harris H, Mayba O, Lianoglou S, Wuster A, Bhangale T, et al. Germline genetic polymorphisms influence tumor gene expression and immune cell infiltration. Proc Natl Acad Sci USA. 2018;115:E11701-10.
- 53. Ogino S, Nowak JA, Hamada T, Milner DA Jr, Nishihara R. Insights into pathogenic interactions among environment, host, and tumor at the crossroads of molecular pathology and epidemiology. Annu Rev Pathol. 2019;14:83-103.
- 54. Ogino S, Nishihara R, Vander Weele TJ, Wang M, Nishi A, Lochhead P, et al. Review article: the role of molecular pathological epidemiology in the study of neoplastic and non-neoplastic diseases in the era of precision medicine. Epidemiology. 2016;27:602-11.