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B-cell lymphoma 2 family members and sarcomas: a promising target in a heterogeneous disease

Rui Caetano Oliveira^{1,2,3*}[®], João Gama⁴[®], José Casanova^{3,5,6}[®]

¹Centro de Anatomia Patológica Germano de Sousa, 3000 Coimbra, Portugal ²Coimbra Institute for Clinical and Biomedical Research (iCBR), 3000 Coimbra, Portugal

³Centre of Investigation on Genetics and Oncobiology (CIMAGO), 3000 Coimbra, Portugal

⁴Pathology Department, Centro Hospitalar e Universitário de Coimbra, 3000 Coimbra, Portugal

⁵Orthopedic Oncology Department, Centro Hospitalar e Universitário de Coimbra, 3000 Coimbra, Portugal

⁶Faculdade de Medicina da Universidade de Coimbra, 3000 Coimbra, Portugal

*Correspondence: Rui Caetano Oliveira, Centro de Anatomia Patológica Germano de Sousa, R. São Teotónio lote 5 nº21, 3000 Coimbra, Portugal. ruipedrocoliveira@hotmail.com

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Abstract

Targeting the B-cell lymphoma 2 (Bcl-2) family proteins has been the backbone for hematological malignancies with overall survival improvements. The Bcl-2 family is a major player in apoptosis regulation and, has captured the researcher's interest in the treatment of solid tumors. Sarcomas are a heterogeneous group of diseases, comprising several entities, with high morbidity and mortality and with few specific therapies available. The treatment for sarcomas is based on platinum regimens, with variable results and poor outcomes, especially in advanced lesions. The high number of different sarcoma entities makes treatment standardization as well as the performance of clinical trials difficult. The use of Bcl-2 family members modifiers has revealed promising results in *in vitro* and *in vivo* models and may be a valid option, especially when used in combination with chemotherapy. In this article, a revision of these results and possibilities for the use of Bcl-2 family members inhibitors in sarcomas was performed.

Keywords

B-cell lymphoma 2, sarcomas, therapy

Introduction

Cancer is a heterogeneous group of diseases characterized by high proliferation, microenvironment modulation, and breakthrough of physical barriers, and evasion of programmed cell death—apoptosis. Apoptosis is essentially a subtype of programmed cell death, fundamental in several physiological and pathological events, eliminating damaged cells [1].

There are two pathways for apoptosis: an extrinsic pathway, mediated by ligands and cell surface death receptors, and an intrinsic pathway, that results from mitochondrial processes [2].

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One of the main regulators of the intrinsic pathway is the B-cell lymphoma 2 (Bcl-2) family of proteins. Recent pharmaceutical developments have explored this pathway as a treatment option, with considerable success [3].

Sarcomas are rare tumors, usually detected in bone and soft tissues, but which have high morbidity and mortality. They comprise more than 50 histological subtypes, prompting additional difficulty for treatment standardization and homogenous therapy [4]. This heterogeneity of sarcomas makes the performance of clinical trials and quality of life evaluation even more difficult [5, 6].

Sarcomas have origin in several genetic alterations, such as fusion proteins, tumor suppressor and cell cycle regulatory gene mutations, and DNA copy-number variations. These changes can be grouped in sarcomas with simple karyotypes and specific genetic alterations [such as EWS RNA binding protein 1 (*EWSR1*) fusions in Ewing sarcoma, paired box 3 (*PAX3*) fusions in alveolar rhabdomyosarcomas, among others] and in sarcomas with complex genetic modifications [7].

Due to this genetic heterogeneity, especially in the setting of advanced disease, therapies are limited, usually based on chemotherapy, with a low overall survival [8]. Due to resistance to monotherapies, a combined treatment with doxorubicin and ifosfamide (core treatment in osteosarcoma [9] and in some histological subtypes of soft tissue sarcoma such as leiomyosarcoma [10]) is usually employed to overcome resistance, but with only a mild improvement in overall survival and with high toxicity reports [11].

In this review, there is a focus on the Bcl-2 functions in cancer, with an emphasis on sarcomas, and their role in precision therapy, mainly of the Bcl-2 homology 3 (BH3)-mimetics/Bcl-2 inhibitors.

The Bcl-2 family

This family is essentially defined by its founder member, the *BCL-2* gene located at chromosome 18, which was first identified by chromosomal analysis of follicular lymphoma [12]. In follicular lymphoma, the translocation t(12;18) leads to a constitutive *BCL-2* expression, which promotes oncogenesis cell death resistance [13]. After this discovery, more than 15 proteins have been added to this family, each with one or more BH domains [3].

The family of Bcl-2 proteins controls the cell death mechanism by regulating the mitochondrial outer membrane permeability, which induces the irreversible release of intermembrane space proteins with caspase activation and apoptosis [14].

Further studies allowed the subclassification into anti-apoptotic [BCL-2, Bcl-extra large (BCL-xL), Bcl-2-like protein 2 (BCL-W), induced myeloid leukemia cell differentiation protein (MCL-1), Bcl-2-related protein A1 (BFL-1/A1)] and pro-apoptotic members. The pro-apoptotic Bcl-2 family members can be further subdivided into multi-BH domains proteins, which contain BH1, BH2 and BH3 domains [Bcl-2-associated X protein (BAX), Bcl-2 homologous antagonist/killer (BAK), Bcl-2 related ovarian killer (BOK)], and BH3-only members [BCL2 associated agonist of cell death (BAD), BH3 interacting domain death agonist (BID), Bcl-2-interacting killer (BIK), Phorbol-12-myristate-13-acetate-induced protein 1 (NOXA), p53 upregulated modulator of apoptosis (PUMA)] [14]. This affects the molecular structure, with the BH3-only members exhibiting a less structured conformation when in solution (except for BID) and the remaining members having a highly conserved tertiary structure [15].

This is associated with distinct functions, and currently, there are two (non-exclusive) models: a direct model where the BH3-only proteins can directly activate Bax and Bak, which are pro-apoptotic, and can be hampered by anti-apoptotic proteins [16]; and an indirect model, where BH3-only proteins challenge the inactive dimerization between anti and pro-apoptotic proteins [17]. The indirect model is very interesting because when Bax and Bak are released from their interaction with anti-apoptotic proteins, they are already in an active state and do not require further activation by BH3-only proteins. This mechanism is the main reason why BH3-mimetics are more efficient in inducing cell death in cells that have a high expression of antiapoptotic proteins, such as cancer cells [18].

Both mechanisms can coexist, but regardless of the mechanism, the result is the oligomerization and conformation changes in Bax and Bak, with the formation of pores in the mitochondrial outer membrane, cytochrome c release, caspase activation and apoptosis induction [19–22].

In physiological conditions, there is an equilibrium between the pro-survival and pro-apoptotic molecules. In cancer, this balance is disrupted, with a shift towards a pro-survival state. The pro-survival proteins (BCL-2 family proteins such as Bcl-2 and Mcl-1) sequester the BH3-only members, preventing them from activating the effectors Bax, Bok and Bak and, therefore, neutralizing the formation of pores in the mitochondria membrane and consequent initiation of apoptosis [23, 24]. As explained before, the use of BH3 mimetic drugs may target Bcl-2. Bcl-xL and Mcl-1, freeing the BH3-only proteins to initiate apoptosis, would be of high therapeutic value.

The first selective Bcl-2 inhibitor was navitoclax (targets Bcl-2, Bcl-xL, and Bcl-W), followed by Obatoclax (targets Bcl-2, Bcl-xL, Mcl-1, and Bcl-W) and venetoclax (selectively targets Bcl-2), with the latter being (probably) the most well-known, due to its success, and favorable safety profile, in the treatment of chronic lymphocytic leukemia, acute myeloid leukemia, some subtypes of lymphoma and multiple myeloma [25]. A simple representation of the main proteins of the Bcl-2 family can be seen in Figure 1.

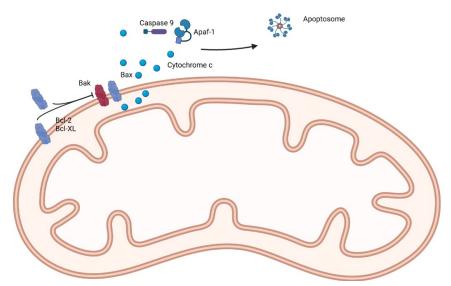


Figure 1. A simple representation of the main proteins of the Bcl-2 family. Apaf-1: apoptotic protease activating factor 1. Created in BioRender™ (https://www.biorender.com/)

Cancers with frequent Bcl-2 alterations

Considering the functions of the Bcl-2 family of proteins is not surprising that they have increased expression in cancer. Evasion of apoptosis is a fundamental process in oncogenesis [26]. This upregulation is found because of several mechanisms, with chromosomal translocation, gene amplification, and increased gene expression/translation as the most common ones.

Bcl-2 translocation is frequently found in follicular lymphoma, as previously mentioned, but is also expressed in other hematologic malignancies such as lymphoma of mucosa-associated lymphoid tissue [27], diffuse large Bcl [28] and acute myeloid leukemia [29]. The Bcl-2 translocation results in overexpression of the Bcl-2 protein, and therefore in the inhibition of the apoptosis [30]. In these hematological malignancies, the Bcl-2 family alterations have been pointed out as new therapeutic possibilities, with the use of selective Bcl-2 inhibitors such as venetoclax, among others [25]. However, the use of these drugs in hematology is not the focus of this review.

Following the studies in hematologic malignancies, many studies have found amplifications of Bcl-2 family genes, such as *MCL1* and *BCL2L1* (responsible for encoding Bcl-xL) in solid tumors, such as lung, breast, uterine and bladder cancer, which was confirmed by The Cancer Genome Atlas data [3]. These

findings have paved the way for clinical trials in lung cancers with navitoclax, a novel Bcl-2 family inhibitor, with interesting results [31].

Studies with colorectal cell lines have found the anti-tumor effect of Bcl-xL inhibitor [32], the same happening in studies with lung cancer cell lines [33]. In a recent study in breast cancer, there was a role of Bcl-xL in cell migration and mitochondrial metabolism, favouring the onset and dissemination of metastases, prompting for modulation of the VDAC1/Bcl-xL interaction as a promising target for anti-tumor therapy in the context of metastatic breast cancer [34]. Bcl-xL inhibitors (A-1155463 and A-1331852, for example) have emerged as a very interesting option, since they appear to have the potential to enhance the efficacy of docetaxel in solid tumors and avoid the exacerbation of neutropenia observed with navitoclax (because inhibits both Bcl-xL and Bcl-2) [35]. Bcl-xL inhibitors are known for their platelet toxicity, but this can be minimized by the adoption of lead-in dosing schemes or by the preselection of thrombocytopenia low-risk patients [36]. Besides the reduction of dose when using a Bcl-xL inhibitor, a key study that investigated the neutropenia induced by navitoclax, showed that this was mainly driven by the Bcl-2 inhibition, and not by the Bcl-xL one [35, 37].

A very intersecting study in 2020 by Scherr et al. [38] found by RNA expression, in more than 1,500 colorectal cancers, that Bcl-xL was the only overactivated antiapoptotic Bcl-2 protein. Using colorectal cancer cells, they were able to demonstrate that the use of WEHI-539 (a Bcl-xL inhibitor) was able to induce apoptosis, and after Bcl-xL knockdown, there was an increase in the response to irinotecan and 5-fluorouracil (5-FU). Still, in the same study, the authors translated those findings to animal models, in which they found that the addition of the Bcl-xL inhibitor to 5-FU caused the strongest anti-tumoral effect, proving support for association therapies. Several studies with a focus in tumor cells state that a Bcl-xL inhibitor as a single agent is not adequate to promote apoptosis [39]. There are reports in cancer cells and animal models, that the use of BTSA1.2 (an orally bioavailable Bax activator) in combination with Bcl-xL has a synergist anti-tumoral effect while being safe for healthy tissues [39, 40].

This mechanism in which solid tumors rely on Bcl-xL for survival has led to several studies focusing on selective Bcl-xL inhibitors [41]. We must also integrate the "non-apoptotic" role of Bcl-xL, namely its mechanism of action for tumor survival, non-related with BH3 and Bcl-2 directly, which should be referred to. Bcl-2 family proteins have a role in mitochondrial structure and functions. Bcl-xL is associated with mitochondrial fission and synaptic activity [42], and it was associated with a different apoptotic function: instead of the death of the entire cell, only a small appendage of a neuron would die, in an event so-called normal synaptic plasticity [41]. What's more curious in these interactions is that on a mouse model with a mutated Bcl-xL, unable to be cleaved, the cells were more resistant to an ischemic event, which is the opposite of its expected function in killing tumor cells [43]. Bcl-xL is also implicated in the calcium homeostasis, more specifically with an increase of calcium leak in the endoplasmic reticulum by direct binding to the inositol 1,4,5-trisphosphate (IP₃) receptors, inducing a low basal calcium concentration in the reticulum, and thus a reduction in stress-mediated calcium release [44, 45].

Therefore, the non-canonical functions of this family of proteins have also to be considered in order to fully understand their biological properties and therapeutic potential, making this a very intriguing puzzle.

Sarcomas with regular Bcl-2 expression

There are sarcomas that are usually associated with Bcl-2 expression, namely on immunohistochemistry. This section will describe them and the possibilities for Bcl-2 targeted therapy.

Synovial sarcoma

Synovial sarcoma (SS) is a sarcoma most often diagnosed in teenagers and young adults. It is highly aggressive, and the standard treatment is chemotherapy with variable results. Nowadays, the diagnosis is based on the finding of the *SS18-SSX* gene fusion, but in the past, a high expression of Bcl-2 by immunohistochemistry was a core element [7]. Since the gene fusion is not a viable target, the modulation

of Bcl-2 expression via inhibitor-based therapies can be a valuable option. However, despite the expression of Bcl-2 in SS, the targeting of Bcl-2 has not been successful in preclinical trials with venetoclax [46].

A recent study has confirmed the high levels of Bcl-2 in SS [47] but found very low levels of *NOXA*, the endogenous inhibitor of MCL-1. Due to that low level of *NOXA*, *MCL-1* was not regulated and, expressed its anti-apoptotic function, resulting in high levels of Bcl-2.

Using cell lines and patient-derived xenografts, the authors demonstrated that low levels of *NOXA* were associated with resistance to venetoclax, similar to what happens in hematologic cancer, inducing an MCL-1 mediated resistance [48]. Using vector methodology, they established a SS cell line with high *NOXA* expression and assessed the sensitivity of the tumor cells to venetoclax-mediated cell death, since *NOXA* specifically binds with MCL-1 and inactivates it [14]. The study demonstrated a synergistic effect of the combined inhibition of Bcl-2 and MCL-1. The inhibition of MCL-1 was able to increase the sensitivity of tumor cells to venetoclax (*in vitro*) and induced tumor regression *in vivo* with the same agent. The mechanism is that inhibition of MCL-1 allows liberation of pro-apoptotic factors, and consequently the induction of apoptosis [47].

Interestingly, one tumor cell line, defined as atypical, without *SS18-SSX* fusion and Bcl-2 expression was resistant to the combination therapy. These findings have a high potential for clinical translation and are in line with other studies that state that the Bcl-2 protein level is not enough for assessing the sensitivity of cancer cells to venetoclax and, MCL-1 and Bcl-xL proteins have a pivotal role in antagonizing the activity of venetoclax [49, 50]. Other resistance mechanisms are described in the literature, such as specific mutations in Bcl-2 (G101V and D103Y) and high resistance to venetoclax [51, 52] but also mutations in BAX (G179E), which cause a diminution in its binding capacity to the mitochondrial membrane, due to translocation to the mitochondria, and therefore preventing BAX-mediated apoptosis [53]. The G179E BAX mutation is also characterized by inducing cross-resistance (at least partial) to other anti-tumoral drugs [53].

Mutation in BAX (G179E) is defined by an inactive form, so in order to target Bax, this has to be activated [54]. There are several drugs available that activate Bax indirectly, but direct Bax activation can provide a specific anti-tumoral approach [54]. However, Bax is rather special, since it has unique and pivotal sites which are not shared by the other members of the Bcl-2 family, prompting mechanisms for a targeted approach with a desired outcome and less adverse effects [55]. Bax participates in both the intrinsic and extrinsic pathways of apoptosis, which does not happen with BH3-only proteins because they depend on Bax/Bak, so Bax activation would be of extreme importance for apoptosis promotion [56]. Many anti-tumoral therapies induce cell death by resorting to Bax activation, so a direct effect and activation of Bax could be also the key for combined therapies [54].

By inducing conformational changes in Bax, they will promote a change from inactive to active state, thus exerting its function. However, some physical constraints such as a negative charge within a hydrophobic alpha-helix, may be an obstacle [57]. Recently, serine 184 (S184) was identified as a critical switch for apoptotic activity control in Bax, and the use of a structural pocket about it was able to provide a docking site for several small molecules of Bax agonists [58]. In this research, Xin et al. [58] were able to develop three compounds—SBMA1, SBMA2, and SBMA3, which induced conformation changes by blocking S184 phosporilation and prevented the creation of a negative charge; Bax was then able to bind with the mitochondrial membrane, with oligomers formation, subsequent cytochrome c release, and apoptosis induction. The anti-tumoral effect was observed in lung cancer cells, but also in *in vivo* models without relevant toxicity in non-tumoral tissue. The activation of Bax directly by small molecules had also been done in 2012 by Gavathiotis et al. [59] but in non-tumoral cells—genetically modified mouse fibroblasts.

The study from Barrott et al. [46] also demonstrated that the specific inhibition of Bcl-2 had a minimal impact on SS. Still, the use of a Bcl-xL inhibitor showed an anti-tumoral activity both in cell lines and animal models, providing evidence that combined therapies could be the solution.

A recent study from Sobol et al. [60] also refers to the therapeutic effect of Bcl-xL inhibitors (ABT-263) as sensitizers for radiotherapy in SS. In this study, the authors found that cell lines treated with the

anti-Bcl-xL showed major radiation-induced damage, even in radiation-resistant cell lines. The potential clinical application of these findings is vast.

The development of new Bcl-2 inhibitors with an affinity for different pro-survival Bcl-2 members and the use of combination therapies targeting also MCL-1 and Bax may be the answer to overcoming these resistances. The Figure 2 represents a SS with Bcl-2 expression.

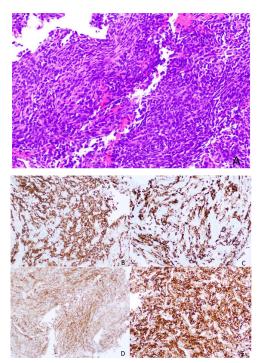


Figure 2. Histology of a SS, with a spindle cell morphology, sometimes with a herringbone pattern. The tumor cells are usually positive for Bcl-2 immunostaining (smaller image in the right lower corner, Figure 2E) *Note.* Reprinted from "Synovial sarcoma of bone: sarcoma typically of soft tissues presenting as a primary bone tumor," by Caracciolo JT, Henderson-Jackson E, Binitie O. Radiol Case Rep. 2018;14:204–7 (https://www.sciencedirect.com/science/article/pii/S1930043318304692?via%3Dihub). CC BY-NC-ND.

Solitary fibrous tumor

Originally described in the pleura, solitary fibrous tumor (SFT) has been described almost in every anatomical location [61]. Recently a *NAB2*-signal transducer and activator of transcription 6 (*STAT6*) gene fusion was discovered as a genetic hallmark of SFT, assessed by immunohistochemistry [62], facilitating the diagnosis, that until then was based on morphology, CD34, and Bcl-2 expression.

The current WHO Classification of Soft Tissue and Bone Tumors classifies SFT as a fibroblastic neoplasm with intermediate (rarely metastasizing) behavior [7] but some cases have been described with features that contribute to malignancy, namely: older age, larger tumor size, increased cellularity, increased mitotic activity ($\geq 4/10$ HPFs or > 2 mitoses/2 mm²), nuclear pleomorphism, tumor necrosis, and infiltrative borders [7, 63]. Interestingly, these features can be seen in recurrences and metastases of SFT that lacked malignant characteristics at the time of primary resection [64]. Several scores have been proposed [7, 65–69], with the one proposed by Demicco EG and colleagues [70] being considered as the most accurate in prognosis.

The Bcl-2 expression in SFT could prompt for targeted therapy, but due to the rarity of this tumor, there is a lack of randomized control trials and no consensus on treatment [71]. Chemotherapy is still the core of the treatment for malignant SFT in neo and adjuvant settings [72]. Anti-angiogenic treatment has been proposed in advanced and aggressive lesions with good results in clinical trials [73, 74], but no targeted Bcl-2 therapies have been described to date.

Ewing sarcoma

Ewing sarcoma (ES) is a high-grade sarcoma, often diagnosed in children and young adults, which may occur both in soft tissue and bone [75]. Morphologically ES have a small, blue, and round cell appearance and are genetically defined by *EWSR1* fusion [76]. Currently, three categories are acknowledged: round cell sarcomas with *EWSR1* gene fusion with non-ETS family members, *CIC*-rearranged sarcomas, and *BCOR* -rearranged sarcomas [7].

The standard care for ES is intensive chemotherapy, neoadjuvant and adjuvant, with good overall survival—75% at five years; however, for patients with relapsed or metastatic disease after treatment, the survival drops to 30% [77]. The most frequent translocation event (in circa 90%) in ES is the *EWSR1-FLI1* t(11;22)(q24;q12), which activates the transcription factor FLI1 [78], but the FLI1 is currently undruggable.

In 2012, studies conducted by Brenner et al. [79] and Garnett et al. [80] demonstrated that ES with the *ESWR1-FLI1* translocation had a high sensitivity to poly ADP ribose polymerase (PARP) inhibitors, which was later confirmed by other groups [81–83]. Despite this promising data, the initial clinical studies were rather disappointing, without objective response to olaparib treatment [84].

To identify the resistance mechanisms, in 2019, a study conducted by Heisey et al. [85] resorting to cell lines and animal xenograft models, was able to demonstrate that the cell line with resistance to olaparib after chemotherapy had a high expression of Bcl-2, which was the responsible for the apoptotic resistance to olaparib. The addition of venetoclax, a Bcl-2 specific inhibitor, did not show sensitization of the tumor cells to olaparib, but the addition of navitoclax, a dual inhibitor of Bcl-2 and Bcl-xL, induced a sensitization of the tumor cells to the PARP inhibitor, with a marked loss of cell viability, even with low doses of olaparib, in two different cell lines, one chemoresistant and other chemosensitive. The experiments in the animal models replicate these findings, with a minimum effect of navitoclax and olaparib in a monotherapy regimen, but when used in combination, they demonstrated a robust inhibition of tumor growth, without augmented hematologic toxicity. Therefore, the co-targeting on Bcl-2 and Bcl-xL can sensitize the ES to olaparib treatment, which may be of benefit in chemotherapy-resistant cases, prompting the need for clinical trials.

A very recent study, from Pascual-Pasto et al. [86] also addressed the question of ES sensitivity to Nab-paclitaxel. Using patient derived xenografts, they were able to demonstrate that low expression of Bcl-2 (assessed by immunohistochemistry) was associated with a high efficacy of Nab-paclitaxel, therefore prompting the use of immunohistochemistry for selecting patients for this therapy.

Pediatric sarcomas

Although rare, pediatric cancer is a major cause of death. Rhabdomyosarcoma is the most frequent sarcoma in children, but neuroblastoma, ES, and osteosarcoma are also common [7]. In haematological malignancies the use of venetoclax is already ongoing in clinical trials for relapsed, refractory or high-risk leukaemia [87, 88]. Based on the previous findings in ES, a study in 2020 by Kehr et al. [89] demonstrated that rhabdomyosarcoma, osteosarcoma, and neuroblastoma cell lines were co-dependent of Bcl-xL and MCL-1 for survival. The single treatment with Bcl-2, Bcl-xL, or MCL-1 inhibitors did not show a valid anti-tumoral response. Once again, the co-treatment with Bcl-xL and MCL-1 inhibitors demonstrated high cellular death induced by caspase-dependent apoptosis. The treatment facilitated the Bax/Bak complex formation and led to a shift in the interaction pattern of the BH3-only proteins BIM and NOXA, disrupting their interactions with Bcl-xL and MCL-1. The co-treatment also displayed anti-tumor activity in the *in vivo* model of rhabdomyosarcoma (embryonic chicken model). Previously, in 2018, Fagar-Uz-Zaman et al. [90] had already stated that BH3 mimetics could be used to increase chemotherapy sensitization in rhabdomyosarcoma, findings that were also stated by Alcon et al. [91] in 2020. This study from 2020 describes that combining BH3 mimetics with conventional chemotherapy (vincristine and doxorubicin) bypasses tumor resistance, with an increase in treatment efficiency and less secondary effects being reported. A very interesting, phase I study, is ongoing, studying the effect of venetoclax added to

chemotherapy in pediatric and young adult patients with any relapsed/refractory tumor type with evidence of BCL-2 expression, which will present additional data to this subject [92].

The role of Bcl-2 as a biomarker in rhabdomyosarcoma has been the subject of several review articles [16, 93, 94] and there are recent papers trying to correlate levels of Bcl-2 and sensitivity to Nab-paclitaxel [86] in rhabdomyosarcoma and ES. New studies should emerge in a short period of time. The Figure 3 represents a rhabdomyosarcoma without changes after conventional treatment.

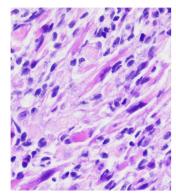


Figure 3. A rhabdomyosarcoma from the orbit with considerable atypia. This case had a posttherapy cytodifferentiation and aggressive clinical course

Note. Adapted from "Embryonal Rhabdomyosarcoma with Posttherapy Cytodifferentiation and Aggressive Clinical Course," by Jeyaraju M, Macatangay RA, Munchel AT, York TA, Montgomery EA, Kallen ME. Case Rep Pathol. 2021;2021:1800854 (https://www.hindawi.com/journals/cripa/2021/1800854/). Copyright © 2021 Maniraj Jeyaraju et al.

Previously, in 2015, a study by Olsen et al. [95] demonstrated that the use of navitoclax (a Bcl-2 family antagonist) was able to sensitize neuroblastoma and ES cell lines to chemotherapy agents, via apoptosis due to Bax activity, especially in N-myc amplified neuroblastoma.

These results should be confirmed using more complex models to ensure the anti-tumoral activity and assess toxicity, namely hepatic, but they pave the way for advanced trials with this co-treatment in pediatric cancer.

Other sarcomas

Despite variable expression of Bcl-2 in soft tissue sarcomas, the anti-Bcl-2 therapy has been studied with promising results. A study from 2020 by Muenchow et al. [96] explored the synergistic association of venetoclax with the proteasome inhibitor bortezomib (BZB). This experiment was based on previous studies that showed that using a proteasome inhibitor stabilized BOK, preventing its ubiquitination and inducing apoptosis [97].

The study by Muenchow et al. [96] was based on tumor cell lines, derived from patients representing six different subtypes of sarcomas: SS, leiomyosarcoma, liposarcoma, rhabdomyosarcoma, chondrosarcoma and osteosarcoma.

The tumor cells had a frequent expression of Bcl-2 and had different p53 status (mutant and wildtype). The study demonstrated a viability reduction of the cells and an induction of apoptotic cell death after therapy with venetoclax and BZB. This effect was mainly due to the accumulation of BOK and the BH3-only protein NOXA, which inhibit the MCL-1 protein (anti-apoptotic). This is a very interesting study since it showed the efficacy of the therapy in different types of sarcomas and was independent of p53 status, with a promising approach for the clinical scenario.

The genetic background is a major player in this setting, since some alterations such as chromosomal rearrangement, *WT1*, and isocitrate dehydrogenase 1 and 2 (*IDH1/2*) point mutations have been associated with high sensitivity to venetoclax [98, 99]. The IDH1/2 mutations are very interesting in sarcomas since they are a hallmark of chondrosarcomas and negatively impact overall survival [100]. Due to the resistance of chondrosarcomas to conventional chemotherapy and radiotherapy [101], the IDH1/2 mutations could

represent a viable option for Bcl-2 inhibitors. Co-therapy may also be a possible solution, using the BH3 mimetic ABT-737, capable of Bcl-2 and Bcl-xL inhibition, rendering the chondrosarcoma sensitive to chemotherapy (doxorubicin or cisplatin) regimens, according to the findings of van Oosterwijk et al. [102], in which they described the high expression of Bcl-2 and Bcl-xL in the mesenchymal, clear cell, and dedifferentiated chondrosarcoma subtypes. Recently, the role of microRNA (miRNA) in regulating Bcl-2 families has been unveiled. Veys et al. [103], in 2021, discovered that Bcl-2 was a direct target of miR-342-5p and that Bcl-xL was a direct target of miR-342-5p and miR-491-5p. The study found that, under hypoxia, miR-342-5p decreased the expression of Bak, and miR-491-5p was associated with Bak upregulation. These findings make miRNA a possible and valuable tool for therapy and shifting resistant chondrosarcomas to more sensitive forms.

Interestingly, in osteosarcomas, there were reports that Bcl-2 could be modulated by miRNA expression, namely miR-143. A study from 2010 [104] showed that miR-143 was downregulated in osteosarcoma cell lines and tumor samples, and its levels of restoration allowed a direct targeting of Bcl-2, reducing cell viability and promoting apoptosis. The interaction of miRNA and Bcl-2 was further studied in 2015 [105], where hypoxia was associated with low levels of miR-15a, promoting invasion and migration of tumor cells. The overexpression of miR-15a was found to repress the tumor cells' invasion and migration via regulation of Bcl-2 and mitochondrial membrane potential. More recently, miR-190b has been shown to have a capacity for inhibiting tumor cell proliferation and promoting apoptosis via Bcl-2 regulation [106]. Pan-inhibitors of Bcl-2 have also been tested in animal models with promising results [107].

Similarly to other sarcoma subtypes, some studies have focused on Bcl-xL and the sensitization of osteosarcoma cell lines to chemotherapy [108], with the use of WEHI-539 (a Bcl-xL selective BH3 mimetic) obtaining a potent increase in apoptosis even with low doses of doxorubicin [109].

Still, in osteosarcomas, a study from 2020 [110] investigated the effects of rapamycin on the apoptosis and proliferation of osteosarcomas cell lines. They found that rapamycin, especially when administered with a Beclin-1 plasmid transfection group, demonstrated an increase of the pro-apoptotic factor Bax and low levels of the anti-apoptotic Bcl-2, thus decreasing the viability of tumor cells and promoting apoptosis. Some studies have focused on the effects of anti-angiogenic drugs, such as apatinib, which exert its effects via vascular endothelial growth factor receptor-2 (*VEGFR2*). A project from Liu et al. [111] on tumor samples (human samples and xenograft models) found that apatinib showed osteosarcoma growth due to inhibition of VEGFR2 and consequently suppressing the STAT3/Bcl-2 signalling pathway, ultimately with low Bcl-2 levels, inducing autophagy and apoptosis. These findings reinforce the role of Bcl-2 in sarcoma progression and therapeutic target.

Leiomyosarcomas are malignant tumors with smooth muscle differentiation [7]. In patients with advanced tumors, there are few therapeutic options, and overall survival is variable according to location—from 15% to 60% [112]. Leiomyosarcomas have a low response to chemo and radiotherapy [113] and despite some good results with trabectedin [114, 115] there are still from the ideal. An example of leiomyosarcoma can be seen in Figure 4.

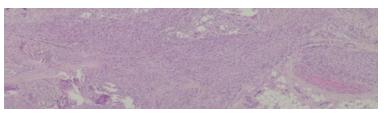


Figure 4. Leiomyosarcoma histological appearance. These tumors are rather resistant to conventional chemotherapy *Note*. Adapted from "Long-Term Response after 94 Cycles of Trabectedin in a Patient with Metastatic Leiomyosarcoma of the Lower Extremity," by Cordeiro M, Casanova JM, Rodrigues J, Freitas J, Fonseca R, Caetano de Oliveira R, et al. Case Rep Oncol. 2020;13:113–9 (https://karger.com/cro/article/13/1/113/95231/Long-Term-Response-after-94-Cycles-of-Trabectedin). CC-BY-NC.

Marieke de Graaff and colleagues [116] explored the immunohistochemical expression of Bcl-2, Bcl-xL, and Bcl-W and found a high expression of these proteins in leiomyosarcomas, not related to histological grade. Based on the high expression of the proteins in most leiomyosarcomas, they explored the response to doxorubicin and ABT-737, and a synergistic effect was assessed, with cell viability diminution and an increase in apoptosis. The data showed an increased sensitivity of leiomyosarcoma to doxorubicin, due to the ABT-737 administration. Of note, in this study, there was also a relatively high expression of Bcl-2, Bcl-xL, and Bcl-W by immunohistochemistry in myxofibrosarcomas and undifferentiated pleomorphic/ spindle cell sarcomas, which may prompt similar studies in these entities.

Conclusions

The inhibition of members of the Bcl-2 family has been evolving and now encompasses a wider range of tumors and is no longer limited to hematological malignancies. The available data supports the use of these therapies in sarcomas, both in the soft tissues and in the bone, especially when used in combination. The co-treatments regimens are promising since they sensitize the tumors to standard chemotherapy regimens, with good biosafety profile. This possibility will be of extreme value, especially in high-grade sarcomas and in chemo resistant subtypes. Studies with more complex models and clinical trials are essential to assess the clinical translational possibilities.

Abbreviation

BAK: B-cell lymphoma 2 homologous antagonist/killer BAX: B-cell lymphoma 2-associated X protein Bcl-2: B-cell lymphoma 2 BCL-W: B-cell lymphoma 2-like protein 2 BCL-xL: B-cell lymphoma-extra large BH3: B-cell lymphoma 2 homology 3 BOK: B-cell lymphoma 2 related ovarian killer ES: Ewing sarcoma IDH1/2: isocitrate dehydrogenase 1 and 2 MCL-1: myeloid leukemia cell differentiation protein miRNA: microRNA NOXA: Phorbol-12-myristate-13-acetate-induced protein 1 SFT: solitary fibrous tumor SS: synovial sarcoma

Declarations

Author contributions

RCO: Conceptualization, Investigation, Writing—original draft. JG: Investigation, Writing—original draft. JC: Supervision, Writing—review & editing.

Conflicts of interest

The authors declare that they have no conflicts of interest.

Ethical approval

Not applicable.

Consent to participate

Not applicable.

Consent to publication

Not applicable.

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References

- 1. Meier P, Finch A, Evan G. Apoptosis in development. Nature. 2000;407:796–801.
- 2. Elmore S. Apoptosis: a review of programmed cell death. Toxicol Pathol. 2007;35:495–516.
- 3. Campbell KJ, Tait SWG. Targeting BCL-2 regulated apoptosis in cancer. Open Biol. 2018;8:180002.
- 4. Gamboa AC, Gronchi A, Cardona K. Soft-tissue sarcoma in adults: an update on the current state of histiotype-specific management in an era of personalized medicine. CA Cancer J Clin. 2020;70:200–29.
- 5. van Eck I, den Hollander D, Desar IME, Soomers VLMN, van de Sande MAJ, de Haan JJ, et al. Unraveling the heterogeneity of sarcoma survivors' health-related quality of life regarding primary sarcoma location: results from the SURVSARC study. Cancers (Basel). 2020;12:3083.
- 6. Du XH, Wei H, Zhang P, Yao WT, Cai QQ. Heterogeneity of soft tissue sarcomas and its implications in targeted therapy. Front Oncol. 2020;10:564852.
- 7. WHO Classification of Tumours Editorial Board. WHO classification of tumours, 5th edition, volume3: soft tissue and bone tumours. 2020.
- 8. Weitz J, Antonescu CR, Brennan MF. Localized extremity soft tissue sarcoma: improved knowledge with unchanged survival over time. J Clin Oncol. 2003;21:2719–25.
- Strauss SJ, Frezza AM, Abecassis N, Bajpai J, Bauer S, Biagini R, et al.; ESMO Guidelines Committee; EURACAN; GENTURIS; ERN PaedCan. Bone sarcomas: ESMO-EURACAN-GENTURIS-ERN PaedCan Clinical Practice Guideline for diagnosis, treatment and follow-up[☆]. Ann Oncol. 2021;32:1520–36.
- Gronchi A, Miah AB, Dei Tos AP, Abecassis N, Bajpai J, Bauer S, et al.; ESMO Guidelines Committee; EURACAN; GENTURIS. Soft tissue and visceral sarcomas: ESMO-EURACAN-GENTURIS Clinical Practice Guidelines for diagnosis, treatment and follow-up[☆]. Ann Oncol. 2021;32:1348–65.
- Tap WD, Papai Z, Van Tine BA, Attia S, Ganjoo KN, Jones RL, et al. Doxorubicin plus evofosfamide versus doxorubicin alone in locally advanced, unresectable or metastatic soft-tissue sarcoma (TH CR-406/SARC021): an international, multicentre, open-label, randomised phase 3 trial. Lancet Oncol. 2017;18:1089–103. Erratum in: Lancet Oncol. 2018;19:e78.
- 12. Tsujimoto Y, Finger LR, Yunis J, Nowell PC, Croce CM. Cloning of the chromosome breakpoint of neoplastic B cells with the t(14;18) chromosome translocation. Science. 1984;226:1097–9.
- 13. Vaux DL, Cory S, Adams JM. *Bcl-2* gene promotes haemopoietic cell survival and cooperates with *c*-*myc* to immortalize pre-B cells. Nature. 1988;335:440–2.
- 14. Kale J, Osterlund EJ, Andrews DW. BCL-2 family proteins: changing partners in the dance towards death. Cell Death Differ. 2018;25:65–80.
- 15. Billen LP, Shamas-Din A, Andrews DW. Bid: a Bax-like BH3 protein. Oncogene. 2008;27:S93–104.

- 16. Hafezi S, Rahmani M. Targeting BCL-2 in cancer: advances, challenges, and perspectives. Cancers (Basel). 2021;13:1292.
- 17. Jeng PS, Inoue-Yamauchi A, Hsieh JJ, Cheng EH. BH3-dependent and independent activation of BAX and BAK in mitochondrial apoptosis. Curr Opin Physiol. 2018;3:71–81.
- 18. Townsend PA, Kozhevnikova MV, Cexus ONF, Zamyatnin AA Jr, Soond SM. BH3-mimetics: recent developments in cancer therapy. J Exp Clin Cancer Res. 2021;40:355.
- 19. Schlesinger PH, Saito M. The Bax pore in liposomes, biophysics. Cell Death Differ. 2006;13:1403–8.
- 20. Qian S, Wang W, Yang L, Huang HW. Structure of transmembrane pore induced by Bax-derived peptide: evidence for lipidic pores. Proc Natl Acad Sci U S A. 2008;105:17379–83.
- 21. Zhang M, Zheng J, Nussinov R, Ma B. Release of cytochrome C from Bax pores at the mitochondrial membrane. Sci Rep. 2017;7:2635.
- 22. Westphal D, Dewson G, Menard M, Frederick P, Iyer S, Bartolo R, et al. Apoptotic pore formation is associated with in-plane insertion of Bak or Bax central helices into the mitochondrial outer membrane. Proc Natl Acad Sci U S A. 2014;111:E4076–85.
- 23. Hata AN, Engelman JA, Faber AC. The BCL2 family: key mediators of the apoptotic response to targeted anticancer therapeutics. Cancer Discov. 2015;5:475–87.
- 24. Einsele-Scholz S, Malmsheimer S, Bertram K, Stehle D, Johänning J, Manz M, et al. Bok is a genuine multi-BH-domain protein that triggers apoptosis in the absence of Bax and Bak. J Cell Sci. 2016;129:2213–23. Erratum in: J Cell Sci. 2016;129:3054.
- 25. Souers AJ, Leverson JD, Boghaert ER, Ackler SL, Catron ND, Chen J, et al. ABT-199, a potent and selective BCL-2 inhibitor, achieves antitumor activity while sparing platelets. Nat Med. 2013;19:202–8.
- 26. Hanahan D. Hallmarks of cancer: new dimensions. Cancer Discov. 2022;12:31–46.
- 27. Troppan K, Wenzl K, Neumeister P, Deutsch A. Molecular pathogenesis of MALT lymphoma. Gastroenterol Res Pract. 2015;2015:102656.
- 28. Matulis SM, Boise LH. BCL2 dependency in diffuse large B-cell lymphoma: it's a family affair. Haematologica. 2020;105:1993–6.
- 29. Wei Y, Cao Y, Sun R, Cheng L, Xiong X, Jin X, et al. Targeting Bcl-2 proteins in acute myeloid leukemia. Front Oncol. 2020;10:584974.
- 30. Johnson NA, Savage KJ, Ludkovski O, Ben-Neriah S, Woods R, Steidl C, et al. Lymphomas with concurrent *BCL2* and *MYC* translocations: the critical factors associated with survival. Blood. 2009;114:2273–9.
- 31. Gandhi L, Camidge DR, Ribeiro de Oliveira M, Bonomi P, Gandara D, Khaira D, et al. Phase I study of navitoclax (ABT-263), a novel Bcl-2 family inhibitor, in patients with small-cell lung cancer and other solid tumors. J Clin Oncol. 2011;29:909–16.
- Zhang H, Xue J, Hessler P, Tahir SK, Chen J, Jin S, et al. Genomic analysis and selective small molecule inhibition identifies BCL-X_L as a critical survival factor in a subset of colorectal cancer. Mol Cancer. 2015;14:126.
- 33. Park D, Magis AT, Li R, Owonikoko TK, Sica GL, Sun SY, et al. Novel small-molecule inhibitors of Bcl-XL to treat lung cancer. Cancer Res. 2013;73:5485–96.
- 34. Bessou M, Lopez J, Gadet R, Deygas M, Popgeorgiev N, Poncet D, et al. The apoptosis inhibitor Bcl-xL controls breast cancer cell migration through mitochondria-dependent reactive oxygen species production. Oncogene. 2020;39:3056–74.
- 35. Leverson JD, Phillips DC, Mitten MJ, Boghaert ER, Diaz D, Tahir SK, et al. Exploiting selective BCL-2 family inhibitors to dissect cell survival dependencies and define improved strategies for cancer therapy. Sci Transl Med. 2015;7:279ra40.
- 36. Vogler M. Targeting BCL2-proteins for the treatment of solid tumours. Adv Med. 2014;2014:943648.

- 37. Wang L, Doherty GA, Judd AS, Tao ZF, Hansen TM, Frey RR, et al. Discovery of A-1331852, a first-inclass, potent, and orally-bioavailable $BCL-X_{L}$ inhibitor. ACS Med Chem Lett. 2020;11:1829–36.
- 38. Scherr AL, Mock A, Gdynia G, Schmitt N, Heilig CE, Korell F, et al. Identification of BCL-XL as highly active survival factor and promising therapeutic target in colorectal cancer. Cell Death Dis. 2020;11:875.
- 39. Lopez A, Reyna DE, Gitego N, Kopp F, Zhou H, Miranda-Roman MA, et al. Co-targeting of BAX and BCL-XL proteins broadly overcomes resistance to apoptosis in cancer. Nat Commun. 2022;13:1199.
- 40. Senichkin VV, Pervushin NV, Zamaraev AV, Sazonova EV, Zuev AP, Streletskaia AY, et al. Bak and Bcl-xL participate in regulating sensitivity of solid tumor derived cell lines to Mcl-1 inhibitors. Cancers (Basel). 2022;14:181.
- 41. Hardwick JM, Soane L. Multiple functions of BCL-2 family proteins. Cold Spring Harb Perspect Biol. 2013;5:a008722.
- 42. Berman SB, Chen YB, Qi B, McCaffery JM, Rucker EB 3rd, Goebbels S, et al. $Bcl-x_{\perp}$ increases mitochondrial fission, fusion, and biomass in neurons. J Cell Biol. 2009;184:707–19.
- 43. Ofengeim D, Chen YB, Miyawaki T, Li H, Sacchetti S, Flannery RJ, et al. N-terminally cleaved Bcl-x_L mediates ischemia-induced neuronal death. Nat Neurosci. 2012;15:574–80.
- 44. Oakes SA, Scorrano L, Opferman JT, Bassik MC, Nishino M, Pozzan T, et al. Proapoptotic BAX and BAK regulate the type 1 inositol trisphosphate receptor and calcium leak from the endoplasmic reticulum. Proc Natl Acad Sci U S A. 2005;102:105–10.
- 45. White C, Li C, Yang J, Petrenko NB, Madesh M, Thompson CB, et al. The endoplasmic reticulum gateway to apoptosis by Bcl-X_L modulation of the InsP₃R. Nat Cell Biol. 2005;7:1021–8. Erratum in: Nat Cell Biol. 2006;8:299.
- 46. Barrott JJ, Zhu JF, Smith-Fry K, Susko AM, Nollner D, Burrell LD, et al. The influential role of BCL2 family members in synovial sarcomagenesis. Mol Cancer Res. 2017;15:1733–40.
- 47. Fairchild CK Jr, Floros KV, Jacob S, Coon CM, Puchalapalli M, Hu B, et al. Unmasking BCL-2 addiction in synovial sarcoma by overcoming low NOXA. Cancers (Basel). 2021;13:2310.
- 48. Bose P, Gandhi V, Konopleva M. Pathways and mechanisms of venetoclax resistance. Leuk Lymphoma. 2017;58:2026–39.
- 49. Punnoose EA, Leverson JD, Peale F, Boghaert ER, Belmont LD, Tan N, et al. Expression profile of BCL-2, BCL-X₁, and MCL-1 predicts pharmacological response to the BCL-2 selective antagonist venetoclax in multiple myeloma models. Mol Cancer Ther. 2016;15:1132–44.
- 50. Rahmani M, Nkwocha J, Hawkins E, Pei X, Parker RE, Kmieciak M, et al. Cotargeting BCL-2 and PI3K induces BAX-dependent mitochondrial apoptosis in AML cells. Cancer Res. 2018;78:3075–86.
- 51. Birkinshaw RW, Gong JN, Luo CS, Lio D, White CA, Anderson MA, et al. Structures of BCL-2 in complex with venetoclax reveal the molecular basis of resistance mutations. Nat Commun. 2019;10:2385.
- 52. Blombery P, Thompson ER, Nguyen T, Birkinshaw RW, Gong JN, Chen X, et al. Multiple *BCL2* mutations cooccurring with Gly101Val emerge in chronic lymphocytic leukemia progression on venetoclax. Blood. 2020;135:773–7.
- 53. Fresquet V, Rieger M, Carolis C, García-Barchino MJ, Martinez-Climent JA. Acquired mutations in BCL2 family proteins conferring resistance to the BH3 mimetic ABT-199 in lymphoma. Blood. 2014;123:4111–9.
- 54. Liu Z, Ding Y, Ye N, Wild C, Chen H, Zhou J. Direct activation of Bax protein for cancer therapy. Med Res Rev. 2016;36:313–41.
- 55. Wei MC, Zong WX, Cheng EH, Lindsten T, Panoutsakopoulou V, Ross AJ, et al. Proapoptotic BAX and BAK: a requisite gateway to mitochondrial dysfunction and death. Science. 2001;292:727–30.

- 56. Zong WX, Lindsten T, Ross AJ, MacGregor GR, Thompson CB. BH3-only proteins that bind pro-survival Bcl-2 family members fail to induce apoptosis in the absence of Bax and Bak. Genes Dev. 2001;15:1481–6.
- 57. Bird GH, Bernal F, Pitter K, Walensky LD. Chapter 22 Synthesis and biophysical characterization of stabilized α-helices of BCL-2 domains. Methods Enzymol. 2008;446:369–86.
- 58. Xin M, Li R, Xie M, Park D, Owonikoko TK, Sica GL, et al. Small-molecule Bax agonists for cancer therapy. Nat Commun. 2014;5:4935.
- 59. Gavathiotis E, Reyna DE, Bellairs JA, Leshchiner ES, Walensky LD. Direct and selective smallmolecule activation of proapoptotic BAX. Nat Chem Biol. 2012;8:639–45.
- 60. Sobol B, Azzam Nieto O, Eberlein EL, Scherr AL, Ismail L, Kessler A, et al. Specific targeting of antiapoptotic Bcl-2 proteins as a radiosensitizing approach in solid tumors. Int J Mol Sci. 2022;23:7850.
- 61. Tariq MU, Din NU, Abdul-Ghafar J, Park YK. The many faces of solitary fibrous tumor; diversity of histological features, differential diagnosis and role of molecular studies and surrogate markers in avoiding misdiagnosis and predicting the behavior. Diagn Pathol. 2021;16:32.
- 62. Doyle LA, Vivero M, Fletcher CD, Mertens F, Hornick JL. Nuclear expression of STAT6 distinguishes solitary fibrous tumor from histologic mimics. Mod Pathol. 2014;27:390–5.
- 63. Ronchi A, Cozzolino I, Zito Marino F, Accardo M, Montella M, Panarese I, et al. Extrapleural solitary fibrous tumor: a distinct entity from pleural solitary fibrous tumor. An update on clinical, molecular and diagnostic features. Ann Diagn Pathol. 2018;34:142–50.
- 64. Rao N, Colby TV, Falconieri G, Cohen H, Moran CA, Suster S. Intrapulmonary solitary fibrous tumors: clinicopathologic and immunohistochemical study of 24 cases. Am J Surg Pathol. 2013;37:155–66.
- 65. Kim JM, Choi YL, Kim YJ, Park HK. Comparison and evaluation of risk factors for meningeal, pleural, and extrapleural solitary fibrous tumors: a clinicopathological study of 92 cases confirmed by STAT6 immunohistochemical staining. Pathol Res Pract. 2017;213:619–25.
- 66. Pasquali S, Gronchi A, Strauss D, Bonvalot S, Jeys L, Stacchiotti S, et al. Resectable extra-pleural and extra-meningeal solitary fibrous tumours: a multi-centre prognostic study. Eur J Surg Oncol. 2016;42:1064–70.
- 67. Demicco EG, Park MS, Araujo DM, Fox PS, Bassett RL, Pollock RE, et al. Solitary fibrous tumor: a clinicopathological study of 110 cases and proposed risk assessment model. Mod Pathol. 2012;25:1298–306.
- 68. Park HK, Yu DB, Sung M, Oh E, Kim M, Song JY, et al. Molecular changes in solitary fibrous tumor progression. J Mol Med (Berl). 2019;97:1413–25.
- 69. Bianchi G, Sambri A, Pedrini E, Pazzaglia L, Sangiorgi L, Ruengwanichayakun P, et al. Histological and molecular features of solitary fibrous tumor of the extremities: clinical correlation. Virchows Arch. 2020;476:445–54.
- 70. Demicco EG, Wagner MJ, Maki RG, Gupta V, Iofin I, Lazar AJ, et al. Risk assessment in solitary fibrous tumors: validation and refinement of a risk stratification model. Mod Pathol. 2017;30:1433–42.
- 71. Davanzo B, Emerson RE, Lisy M, Koniaris LG, Kays JK. Solitary fibrous tumor. Transl Gastroenterol Hepatol. 2018;3:94.
- 72. de Bernardi A, Dufresne A, Mishellany F, Blay JY, Ray-Coquard I, Brahmi M. Novel therapeutic options for solitary fibrous tumor: antiangiogenic therapy and beyond. Cancers (Basel). 2022;14:1064.
- 73. Martin-Broto J, Stacchiotti S, Lopez-Pousa A, Redondo A, Bernabeu D, de Alava E, et al. Pazopanib for treatment of advanced malignant and dedifferentiated solitary fibrous tumour: a multicentre, singlearm, phase 2 trial. Lancet Oncol. 2019;20:134–44.
- 74. Martin-Broto J, Cruz J, Penel N, Le Cesne A, Hindi N, Luna P, et al. Pazopanib for treatment of typical solitary fibrous tumours: a multicentre, single-arm, phase 2 trial. Lancet Oncol. 2020;21:456–66.

- 75. Sbaraglia M, Righi A, Gambarotti M, Dei Tos AP. Ewing sarcoma and Ewing-like tumors. Virchows Arch. 2020;476:109–19.
- 76. Folpe AL, Goldblum JR, Rubin BP, Shehata BM, Liu W, Dei Tos AP, et al. Morphologic and immunophenotypic diversity in Ewing family tumors: a study of 66 genetically confirmed cases. Am J Surg Pathol. 2005;29:1025–33.
- 77. Balamuth NJ, Womer RB. Ewing's sarcoma. Lancet Oncol. 2010;11:184–92.
- 78. Tomazou EM, Sheffield NC, Schmidl C, Schuster M, Schönegger A, Datlinger P, et al. Epigenome mapping reveals distinct modes of gene regulation and widespread enhancer reprogramming by the oncogenic fusion protein EWS-FLI1. Cell Rep. 2015;10:1082–95.
- 79. Brenner JC, Feng FY, Han S, Patel S, Goyal SV, Bou-Maroun LM, et al. PARP-1 inhibition as a targeted strategy to treat Ewing's sarcoma. Cancer Res. 2012;72:1608–13.
- 80. Garnett MJ, Edelman EJ, Heidorn SJ, Greenman CD, Dastur A, Lau KW, et al. Systematic identification of genomic markers of drug sensitivity in cancer cells. Nature. 2012;483:570–5.
- 81. Stewart E, Goshorn R, Bradley C, Griffiths LM, Benavente C, Twarog NR, et al. Targeting the DNA repair pathway in Ewing sarcoma. Cell Rep. 2014;9:829–40.
- Engert F, Schneider C, Weiβ LM, Probst M, Fulda S. PARP inhibitors sensitize Ewing sarcoma cells to temozolomide-induced apoptosis via the mitochondrial pathway. Mol Cancer Ther. 2015;14:2818–30.
- 83. Lee HJ, Yoon C, Schmidt B, Park DJ, Zhang AY, Erkizan HV, et al. Combining PARP-1 inhibition and radiation in Ewing sarcoma results in lethal DNA damage. Mol Cancer Ther. 2013;12:2591–600.
- 84. Choy E, Butrynski JE, Harmon DC, Morgan JA, George S, Wagner AJ, et al. Phase II study of olaparib in patients with refractory Ewing sarcoma following failure of standard chemotherapy. BMC Cancer. 2014;14:813.
- 85. Heisey DAR, Lochmann TL, Floros KV, Coon CM, Powell KM, Jacob S, et al. The Ewing family of tumors relies on BCL-2 and BCL-X_L to escape PARP inhibitor toxicity. Clin Cancer Res. 2019;25:1664–75.
- 86. Pascual-Pasto G, Resa-Pares C, Castillo-Ecija H, Aschero R, Baulenas-Farres M, Vila-Ubach M, et al. Low Bcl-2 is a robust biomarker of sensitivity to nab-paclitaxel in Ewing sarcoma. Biochem Pharmacol. 2023;208:115408.
- 87. Pikman Y, Tasian SK, Sulis ML, Stevenson K, Blonquist TM, Apsel Winger B, et al. Matched targeted therapy for pediatric patients with relapsed, refractory, or high-risk leukemias: a report from the LEAP consortium. Cancer Discov. 2021;11:1424–39.
- 88. Obszański P, Kozłowska A, Wańcowiat J, Twardowska J, Lejman M, Zawitkowska J. Moleculartargeted therapy of pediatric acute myeloid leukemia. Molecules. 2022;27:3911.
- 89. Kehr S, Haydn T, Bierbrauer A, Irmer B, Vogler M, Fulda S. Targeting BCL-2 proteins in pediatric cancer: dual inhibition of BCL-X_L and MCL-1 leads to rapid induction of intrinsic apoptosis. Cancer Lett. 2020;482:19–32.
- 90. Faqar-Uz-Zaman SF, Heinicke U, Meister MT, Vogler M, Fulda S. BCL-x_L-selective BH3 mimetic sensitizes rhabdomyosarcoma cells to chemotherapeutics by activation of the mitochondrial pathway of apoptosis. Cancer Lett. 2018;412:131–42.
- 91. Alcon C, Manzano-Muñoz A, Prada E, Mora J, Soriano A, Guillén G, et al. Sequential combinations of chemotherapeutic agents with BH3 mimetics to treat rhabdomyosarcoma and avoid resistance. Cell Death Dis. 2020;11:634.
- 92. Place AE, Goldsmith K, Bourquin JP, Loh ML, Gore L, Morgenstern DA, et al. Accelerating drug development in pediatric cancer: a novel phase I study design of venetoclax in relapsed/refractory malignancies. Future Oncol. 2018;14:2115–29.
- 93. Salah HT, DiNardo CD, Konopleva M, Khoury JD. Potential biomarkers for treatment response to the BCL-2 inhibitor venetoclax: state of the art and future directions. Cancers (Basel). 2021;13:2974.

- 94. Chen C, Dorado Garcia H, Scheer M, Henssen AG. Current and future treatment strategies for rhabdomyosarcoma. Front Oncol. 2019;9:1458.
- 95. Olsen RR, Mary-Sinclair MN, Yin Z, Freeman KW. Antagonizing Bcl-2 family members sensitizes neuroblastoma and Ewing's sarcoma to an inhibitor of glutamine metabolism. PLoS One. 2015;10:e0116998.
- 96. Muenchow A, Weller S, Hinterleitner C, Malenke E, Bugl S, Wirths S, et al. The BCL-2 selective inhibitor ABT-199 sensitizes soft tissue sarcomas to proteasome inhibition by a concerted mechanism requiring BAX and NOXA. Cell Death Dis. 2020;11:701.
- 97. Llambi F, Wang YM, Victor B, Yang M, Schneider DM, Gingras S, et al. BOK is a non-canonical BCL-2 family effector of apoptosis regulated by ER-associated degradation. Cell. 2016;165:421–33.
- 98. Chan SM, Thomas D, Corces-Zimmerman MR, Xavy S, Rastogi S, Hong WJ, et al. Isocitrate dehydrogenase 1 and 2 mutations induce BCL-2 dependence in acute myeloid leukemia. Nat Med. 2015;21:178–84.
- 99. Campos EDV, Pinto R. Targeted therapy with a selective BCL-2 inhibitor in older patients with acute myeloid leukemia. Hematol Transfus Cell Ther. 2019;41:169–77.
- 100. Vuong HG, Ngo TNM, Dunn IF. Prognostic importance of IDH mutations in chondrosarcoma: an individual patient data meta-analysis. Cancer Med. 2021;10:4415–23.
- 101. Nazeri E, Gouran Savadkoohi M, Majidzadeh-A K, Esmaeili R. Chondrosarcoma: an overview of clinical behavior, molecular mechanisms mediated drug resistance and potential therapeutic targets. Crit Rev Oncol Hematol. 2018;131:102–9.
- 102. van Oosterwijk JG, Meijer D, van Ruler MA, van den Akker BE, Oosting J, Krenács T, et al. Screening for potential targets for therapy in mesenchymal, clear cell, and dedifferentiated chondrosarcoma reveals Bcl-2 family members and TGFβ as potential targets. Am J Pathol. 2013;182:1347–56.
- 103. Veys C, Benmoussa A, Contentin R, Duchemin A, Brotin E, Lafont JE, et al. Tumor suppressive role of miR-342-5p in human chondrosarcoma cells and 3D organoids. Int J Mol Sci. 2021;22:5590.
- 104. Zhang H, Cai X, Wang Y, Tang H, Tong D, Ji F. microRNA-143, down-regulated in osteosarcoma, promotes apoptosis and suppresses tumorigenicity by targeting Bcl-2. Oncol Rep. 2010;24:1363–9.
- 105. Leng J, Song Q, Zhao Y, Wang Z. miR-15a represses cancer cell migration and invasion under conditions of hypoxia by targeting and downregulating Bcl-2 expression in human osteosarcoma cells. Int J Oncol. 2018;52:1095–104.
- 106. Kang M, Xia P, Hou T, Qi Z, Liao S, Yang X. MicroRNA-190b inhibits tumor cell proliferation and induces apoptosis by regulating Bcl-2 in U2OS osteosarcoma cells. Pharmazie. 2017;72:279–82.
- 107. Masuelli L, Benvenuto M, Izzi V, Zago E, Mattera R, Cerbelli B, et al. *In vivo* and *in vitro* inhibition of osteosarcoma growth by the pan Bcl-2 inhibitor AT-101. Invest New Drugs. 2020;38:675–89.
- 108. Wang ZX, Yang JS, Pan X, Wang JR, Li J, Yin YM, et al. Functional and biological analysis of Bcl-xL expression in human osteosarcoma. Bone. 2010;47:445–54.
- 109. Baranski Z, de Jong Y, Ilkova T, Peterse EF, Cleton-Jansen AM, van de Water B, et al. Pharmacological inhibition of Bcl-xL sensitizes osteosarcoma to doxorubicin. Oncotarget. 2015;6:36113–25.
- 110. Yu WX, Lu C, Wang B, Ren XY, Xu K. Effects of rapamycin on osteosarcoma cell proliferation and apoptosis by inducing autophagy. Eur Rev Med Pharmacol Sci. 2020;24:915–21.
- 111. Liu K, Ren T, Huang Y, Sun K, Bao X, Wang S, et al. Apatinib promotes autophagy and apoptosis through VEGFR2/STAT3/BCL-2 signaling in osteosarcoma. Cell Death Dis. 2017;8:e3015.
- 112. Wang Z, Shi N, Naing A, Janku F, Subbiah V, Araujo DM, et al. Survival of patients with metastatic leiomyosarcoma: the MD Anderson Clinical Center for targeted therapy experience. Cancer Med. 2016;5:3437–44.
- 113. Villar VH, Vögler O, Martínez-Serra J, Ramos R, Calabuig-Fariñas S, Gutiérrez A, et al. Nilotinib counteracts P-glycoprotein-mediated multidrug resistance and synergizes the antitumoral effect of doxorubicin in soft tissue sarcomas. PLoS One. 2012;7:e37735.

- 114. Nakamura T, Sudo A. The role of trabectedin in soft tissue sarcoma. Front Pharmacol. 2022;13:777872.
- 115. Cordeiro M, Casanova JM, Rodrigues J, Freitas J, Fonseca R, Caetano de Oliveira R, et al. Long-term response after 94 cycles of trabectedin in a patient with metastatic leiomyosarcoma of the lower extremity. Case Rep Oncol. 2020;13:113–9.
- 116. de Graaff MA, de Rooij MAJ, van den Akker BEWM, Gelderblom H, Chibon F, Coindre JM, et al. Inhibition of Bcl-2 family members sensitises soft tissue leiomyosarcomas to chemotherapy. Br J Cancer. 2016;114:1219–26.