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Early taurine administration as a means for halting the cytokine storm progression in COVID-19 patients

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Academic Editor: Esma R. Isenovic, University of Belgrade, Serbia

Received: February 23, 2022 Accepted: May 9, 2022 Published: June 8, 2022

Cite this article: Rubio-Casillas A, Gupta RC, Redwan EM, Uversky VN, Badierah R. Early taurine administration as a means for halting the cytokine storm progression in COVID-19 patients. Explor Med. 2022;3:234–48. https://doi.org/10.37349/emed.2022.00088

Abstract

Around the world, more than 6.2 million individuals have died as a result of coronavirus disease 2019 (COVID-19). According to a recent survey conducted among immunologists, epidemiologists, and virologists, this disease is expected to become endemic. This implies that the disease could have a continuous presence and/or normal frequency in the population. Pharmacological interventions to prevent infection, as well as to treat the patients at an early phase of illness to avoid hospitalization are essential additions to the vaccines. Taurine is known to inhibit the generation of all inflammatory mediators linked to the cytokine storm. It can also protect against lung injury by suppressing increased oxidants production and promoting the resolution of the inflammatory process. Neutrophil lactoferrin degranulation stimulated by taurine may have antiviral effects against SARS-CoV-2, limiting viral replication. It is hypothesized that if taurine is administered early in the onset of COVID-19 disease, it may stop the cytokine storm from progressing, lowering morbidity and mortality.

Keywords

COVID-19, cytokine storm, efferocytosis, hypochlorous acid, macrophage polarization, myeloperoxidase, SARS-CoV-2, taurine

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Introduction

In its critical stage, the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) may provoke deadly pneumonia [1, 2]. Although coronavirus disease 2019 (COVID-19) caused by SARS-CoV-2 infection is characterized by a mortality rate of 2.2% [3], this disease is more severe in aged patients and patients with underlying maladies [4–6]. For example, the mortality rate of patients with underlying health conditions was shown to reach 12% [4]. Furthermore, in COVID-19 patients who developed acute respiratory distress syndrome (ARDS), the mortality rate can be as high as 40% to 50% [7, 8]. Even though the factors involved in COVID-19-mediated lung damage remain mostly unknown, in the published research, the phrase "cytokine storm"; i.e., a hyper-inflammatory state originating from the uncontrolled release of cytokines by a dysregulated immunological system has been frequently associated with the related pathological alterations [9–37]. In general terms, it represents an overactive immunological reaction distinguished by the production of interferon, interleukins, tumor necrosis factors, chemokines, and many other mediators, such compounds are core components of a well-preserved innate immunological response essential for the effective removal of pathogens. The term "cytokine storm" refers to a scenario in which several cytokines released are harmful to the host [9, 38].

One of the most dangerous COVID-19 complications is the emergence of a severe upper respiratory lung disease, which presents a big challenge to physicians in terms of illness treatment [39]. The majority of sick individuals with SARS-CoV-2 infection experienced minor symptoms, according to a study of 41 patients with COVID-19, however, some of them later displayed serious symptoms and eventually died due to organic failure as a consequence of an excessive cytokine storm [2]. It is worth noting that the production of numerous cytokines is closely linked to the onset of the first symptoms. For example, interferon-alpha (IFN- α) release causes fever, headaches, chills, drowsiness, and tiredness. Tumor necrosis factor-alpha (TNF- α) can induce symptoms similar to the flu, but it can also cause vascular fragility, heart disease, and lung injury. Interleukin 6 (IL-6) release may cause complement activation and clotting cascade stimulation, causing severe symptoms including disseminated intravascular coagulation [40–42].

It is interesting to note that IL-6 may cause cardiomyopathy by increasing cardiac failure, which is common in cytokine release syndrome (CRS) patients. Furthermore, endothelial cell activation could be one of the symptoms of critical CRS [43]. Malfunctions in the endothelia are characterized by capillary leakage, low blood pressure, and coagulation alterations [44]. These data show that immunopathological mechanisms induced by SARS-CoV-2 are important in fatal pneumonia that occurs after coronavirus infections [45]. A cytokine storm is defined as "a potentially deadly immune disorder characterized by accelerated growth and enhanced activation of T cells, macrophages, and natural killer cells, and also elevated release of over 150 inflammatory cytokines and signaling molecules released by immune and non-immune cells" [46]. In viral diseases, aberrant inflammatory cytokine release causes epithelial and endothelial cell death, affecting lung microcirculation and alveolar epithelial cell permeability, and resulting in vascular bleeding, alveolar edema, and hypoxia [39]. Other organ systems affected by COVID-19 outside the lungs include the heart, kidneys, vascular system, liver, and brain [47].

In addition, it is well known that patients with COVID-19 may have a worse prognosis if their blood glucose levels are high, increasing the likelihood of mechanical ventilation, shock, and multiple organ failure, requiring intensive care unit (ICU) therapy [48]. During COVID-19 infection, people with type 2 diabetes mellitus had a higher incidence, severity of symptoms, and mortality [49]. According to current findings, glycemic management has a significant impact on the effectiveness of the immunological response in patients receiving a messenger ribonucleic acid (mRNA) vaccine against SARS-CoV-2 [50]. Certainly, hyperglycemia during vaccination impairs the immune response: a recent investigation found that at 21 days after the first vaccine dose, neutralizing antibody titers and CD4 cytokine responses involving type 1 helper T cells were lower in type two diabetic patients with glycosylated hemoglobin (HbA1c) levels > 7% than in individuals with HbA1c levels \leq 7% [50].

In patients with acute myocardial infarction, the significance of SARS-CoV-2 in causing endothelial dysfunction and enhanced coagulation is well documented [51]. Furthermore, enhanced

angiotensin-converting enzyme 2 (ACE2) glycation and transmembrane serine protease 2 (TMPRSS2) expression in cardiac myocytes as a result of diabetes mellitus have been shown to facilitate SARS-CoV-2 entry in the host cell; as a consequence, about 50% of COVID-19 patients with diabetes mellitus who were hospitalized experienced myocardial injury [52].

Taurine is one of the most prevalent non-essential amino acids in mammals, with several physiological activities in the neurological, cardiovascular, renal, endocrine, and immunological systems. Inflammation causes taurine to be halogenated in phagocytes, resulting in taurine chloramine (TauCl) synthesis. It is formed in active neutrophils by a reaction with hypochlorous acid (HOCl) produced by the halide-dependent myeloperoxidase (MPO) system. Following apoptosis, TauCl is released from active neutrophils and inhibits the synthesis of inflammatory mediators such as superoxide anion, nitric oxide, TNF- α , interleukins, and prostaglandins in inflammatory cells and tissues. TauCl also stimulates the expression of antioxidant proteins in macrophages, including heme oxygenase 1, peroxiredoxin, thioredoxin, glutathione peroxidase, and catalase [53]. Because taurine protects many tissues from oxidative stress [54–58], the scientific rationale for employing taurine as a therapeutic in the early stages of COVID-19 will be presented in this work.

The neutrophil leukocytes function in COVID-19 disease

The pathological changes associated with severe COVID-19 disease are characterized by neutrophil quantity, phenotype, and performance alterations. Following SARS-CoV-2 infection, an increased number of neutrophils were found in the nasopharynx [59] and after in the more distant areas of the lung [60]. Elevated neutrophil number in the bloodstream has been recognized as a marker of COVID-19 [8, 61], and neutrophil activation pathways are major components of the severe cases [62, 63]. The neutrophil-to-lymphocyte ratio (NLR), a recognized biomarker of infection and generalized inflammation, has shown that COVID-19 patients have an increased inflammatory reaction [8].

Because acute respiratory distress syndrome is the most frequent cause of death in COVID-19 sufferers, high NLR rates predict a grave prognosis [64]. When severe COVID-19 patients who were sent to the ICU were compared to other COVID-19 patients who were not sent to the ICU, the former patients had low lymphocyte count and increased neutrophil number and NLR, according to a recent study [65]. Furthermore, some COVID-19 sick people also showed an increased number of neutrophils and a decreased lymphocyte number through the critical stage [45]. Pulmonary capillaries of the COVID-19 patients had considerable neutrophil infiltration, according to another research [66].

The role of MPO and HOCl in the cytokine storm

MPO, or myeloperoxidase, is an enzyme found mostly in neutrophils and macrophages [67]. When chemically reacts with superoxide, hydrogen peroxide, and chloride, MPO induces the synthesis of HOCl [67]. Although the MPO/HOCl complex is required for phagocytes to kill bacteria and viruses, MPO can be released outside the cell, increasing the risk of tissue harm [68]. HOCl is the oxidant with the highest reactivity and generated in considerable amounts in our bodies, and has a reaction that is many times greater with most substrates than hydrogen peroxide, hydroperoxides, and peroxynitrite [69]. HOCl has been shown *in vitro* to increase nuclear factor-kappa B (NF- κ B) activity, the attachment of a phosphoryl group to tyrosine in T and B lymphocytes, calcium signaling, and TNF- α production [70, 71].

Many COVID-19 patients, especially the elderly, who eventually developed respiratory failure, had hypoxemia and hypocapnia without any indications of respiratory distress [72]. This condition is also called "happy hypoxemia" or "quiet hypoxemia", and it was first seen in patients during the Wuhan outbreak [72]. The exact cause of this asymptomatic hypoxia is unknown, although the accumulation of HOCl in the blood during COVID-19 has been suggested to cause tissue hypoxia due to a lack of normal hemoglobin (Hb) activity [73]. Interestingly, it is notorious that Hb has a higher affinity for HOCl than for O_2 ; that is, HOCl acts as a competitive inhibitor with O_2 for heme binding sites, thus resulting in a decreased percentage of oxygen in the blood [73, 74]. Furthermore, HOCl is involved in a plethora of oxidative reactions, including iron oxidation of the Hb-heme complex, heme breakdown, and the final liberation of free iron, which

together promote harmful tissue damage by generating reactive oxygen species (ROS) and using nitric oxide (NO). Linking these reactions in an integral model might explain the significant cellular injury, vascular constriction, and extreme hypoxemia as reported in COVID-19 sick people with clinical worsening [73].

Notably, taurine has been shown to prevent MPO degranulation selectively, safeguarding organs from oxidative harm caused by exaggerated MPO release. This inhibition of MPO release produced a significant reduction in neutrophil extracellular traps (NETs) [75].

NETs are extracellular webs of DNA, histones, microbicidal proteins, and oxidant enzymes that are released by neutrophils to control infections; however, when not properly regulated, NETs have the potential to initiate and propagate inflammation, thrombosis [76, 77], and respiratory arrest [78, 79]. SARS-CoV-2 does cause, in fact, a prothrombotic condition with dangerous consequences including deep vein thrombosis [80]. Increased NETs, which can propagate inflammation and microvascular thrombosis in individuals with SARS-CoV-2 infection, are primarily responsible for this outcome [81, 82]. In addition, SARS-CoV-2 infection has been shown to directly cause NETs formation and release in healthy neutrophils, which induces lung epithelial cell death *in vitro* [81].

The discovery that taurine-mediated inhibition of MPO release produced a significant reduction in NETs is transcendental [75] since NETs generation was discovered to be increased in severe COVID-19 cases, and it is linked to immuno-thrombosis and the development of acute lung injury/acute respiratory distress syndrome (ALI/ARDS) [83]. Of note, inhibition of excessive neutrophil function and NETs formation by an anti-MPO antibody was protective in various models of influenza-associated ARDS [84].

Taurine reprograms macrophage M1 polarization by inhibiting mitophagy

Alveolar macrophages are known to remove billions of air pollutants, allergens, and microorganisms per day [85, 86]. Macrophages may shift into M1 or M2 phenotypes to preserve alveolar homeostasis. Macrophages have been classified as "polarized" either towards M1 mode (proinflammatory, regulating pathogen resistance) or M2 mode (anti-inflammatory, facilitating tissue remodeling) (Figure 1) [86, 87].



Figure 1. The M1 and M2 macrophage polarization states have distinct phenotypic properties. An inflammatory stimulus in the lungs, such as virus infection, lipopolysaccharide (LPS), dust particles, or IFN- γ , causes local macrophages to transition from M2 to M1 phenotype. Furthermore, circulating monocytes will be attracted into lung tissues and activated by the inflammatory stimulation, with M1 macrophages accounting for the majority of macrophages in the inflammatory zone (M1 dominant). Inducible nitric oxide synthase (iNOS), IL-6, IL-12, IL-1 β , and TNF- α are inflammatory cytokines secreted by M1 macrophages. M2 macrophages release anti-inflammatory cytokines: C-C motif ligand 18 (CCL18), CCL22, and IL-10

Note. Adapted from "Lung macrophage phenotypes and functional responses: role in the pathogenesis of COPD," by Yamasaki K, van Eeden SF. Int J Mol Sci. 2018;19:582 (https://doi.org/10.3390/ijms19020582). CC BY.

The CD206 receptor is routinely expressed by alveolar macrophages, according to the results of a blood cytometry study of 72 donors without lung pathology [88]. Considering that CD206 is a characteristic M2 marker, human alveolar macrophages appear to be biased toward an M2-like phenotype, which is not ideal for virus propagation [89]. SARS-CoV-2 sabotages alveolar macrophages for its reproduction and dissemination, leading to enhanced pathology in the lungs; it was also discovered that SARS-CoV-2 may induce the M1 profile in alveolar macrophages by releasing inflammatory cytokines [89].

These cytokines, such as TNF-α, may promote macrophage polarization toward an M1-like state [90, 91]. Interestingly, the reduction of CD206⁺ M2 macrophages *in vivo* exacerbates acute lung damage, which is associated with neutrophil migration to the lungs [92]. If the viral infection in the alveoli surpasses a certain threshold, SARS-CoV-2 may directly reprogram alveolar macrophages to an M1 phenotype, and the resulting pro-inflammatory cytokines [93] may further promote M1 polarization, allowing viral transmission via alveolar macrophages [89]. In addition, bronchoalveolar liquid from patients with severe COVID-19 disease had greater M1 macrophage counts, but such fluid from less severe infected people and healthy controls had a higher M2 macrophage count [60].

High intracellular MPO activity is required for macrophage activation [94], and MPO-mediated HOCl production has been demonstrated to be a major cause of macromolecular oxidative injury as reported in several investigations [69, 95, 96]. Concretely, cellular contact with HOCl can have a variety of negative consequences by affecting the cellular redox state [97], so avoiding more HOCl generation during the cytokine storm is critical to controlling excessive MPO activity [73]. In this regard, it is known that TauCl "inhibits macrophage production of inflammatory mediators like macrophage inflammatory protein-2 (MIP-2), monocyte chemoattractant protein-1 and 2 (MCP-1 and 2), nitric oxide, nitrites, prostaglandin E2 (PGE2), TNF- α , and IL-6" [98–101]. Monocytes and macrophages are transformed from an inflammatory (M1) to a basal (M2) phagocytic state by these down-regulation pathways [102].

Finally, a recent work discovered that taurine suppresses macrophage M1 polarization by inhibiting mitophagy (mitochondrial autophagy) and increasing the synthesis of M2-markers (CD206 and IL-10) by decreasing the production of M1 inflammatory factor genes [103]. Without an adequate number of mitochondria, macrophages turn to glycolytic metabolism, which is linked to the M1 phenotype; this change is triggered in alveolar macrophages by SARS-CoV2, which favors viral replication [89]. Excessive M1 polarization thus induces a pro-inflammatory state, that can develop in a cytokine storm if it is not adequately managed [20]. Taurine reprograms the metabolic activity of M1 macrophages by sustaining a high number of mitochondria; this restricts the transition of energy metabolism to glycolysis, which is essential for the metabolism of M1 macrophages [103]. All of these findings lead us to propose that if taurine is given early in the disease, it may be able to stop the cytokine storm that is linked with severe COVID-19 cases.

Taurine neutralizes HOCI-induced tissue damage

HOCl is formed within the phagocytic vacuole when neutrophils internalize and destroy bacteria or viruses, and while it is a potent bactericide, it can also be hazardous since it oxidizes many biological components [104]. Taurine chemically reacts with HOCl in leukocytes to produce TauCl (a more balanced, less aggressive, and specific oxidant compared with HOCl) [105–107]. In biological systems, TauCl is thought to operate as a general antioxidant (scavenger), protecting cells from self-destruction during activities that create excessive HOCl [100]. Taurine may also protect epithelial cells of the lungs against HOCl/ hypochlorite (OCl⁻)-mediated lysis by lowering chlorination of cell components, rather than preventing sulfhydryl group oxidation [108].

Notably, TauCl promotes nuclear factor erythroid 2 (Nrf2) to enter the nucleus and triggers the production of several anti-oxidant proteins [109], and prevents the generation of inflammatory cytokines [110, 111]. Because NF- κ B is known to influence inflammatory cytokine gene production [112], TauCl is thought to decrease NF- κ B activation. The chemical mechanism underpinning TauCl-induced suppression of NF- κ B activation was unknown at the time, but it was eventually discovered to be oxidization of the inhibitor of nuclear factor kappa B alpha protein (I κ B α) at methionine⁴⁵ [112].

Taurine protects against lung damage by mitigating oxidative stress

Taurine is a functionally important molecule, and it has been discovered to be a powerful cell guardian, lowering oxidative stress and inflammatory factor production [113, 114]. Taurine pre-treatment of rats before acute ozone exposure reduces oxidant-mediated lung damage by preventing the generation of nitrite and the release of TNF- α , which are both related to tissue damage [101]. Taurine has been shown to counteract paraquat-mediated reductions in glutathione (GSH) level and GSH peroxidase function in other studies [113], inhibits oxidative stress-mediated cell apoptosis and also lowers malondialdehyde (MDA) production [115]. In research mimicking limb ischemia reperfusion-induced lung damage, taurine increased superoxide dismutase (SOD) and catalase functions by decreasing stress in the endoplasmic reticulum [115].

Taurine also synergized the protective effect of dexmedetomidine against sepsis-induced lung damage by blocking the NF- κ B pathway and lowering IL-6 and IL-1 expression levels [114]. In a recent study, taurine's beneficial effects on sepsis-mediated lung injury were investigated, and it was discovered that taurine administration reduced sepsis-mediated elevations in TNF- α and IL-1 expression in lung cells. These beneficial effects were linked to the control of inflammation and oxidant stress via interruption of the p38/mitogen-activated protein kinase (MAPK) and NF- κ B regulatory pathways [116].

Acute lung damage is a frequent manifestation of Gram-negative bacterial infections and a primary origin of disease complications and deaths in humans [117]. The internal toxin LPS, which is located in the external membranes of bacteria, is a frequent cause of ALI [118]. In contrast to control animals' lungs, LPS-treated mice's lungs exhibited increased cellular apoptosis, lipid peroxidation, reduced concentrations of glutathione catalase, glutathione peroxidase, and superoxide dismutase, and inflammation limited to the parenchyma [119]. All of these changes were greatly reduced after taurine supplementation, showing that this amino acid has anti-inflammatory and antioxidant capabilities that help defend the lungs from the toxic effects of Gram-negative bacterial endotoxin [119].

TauCl contributes to the resolution of inflammation

The death of neutrophils is required for inflammation to be resolved; however, this does not always occur, and neutrophils keep driving the inflammatory process [120]. MPO has been found to exert an unforeseen function in affecting neutrophil destiny and, as a result, the persistence of inflammation—MPO extended the lifespan of neutrophils by regulating the constitutive cell death pathway (apoptosis), postponing the resolution of inflammation [120]. Individuals suffering from sepsis-associated ARDS were recently discovered to have dysfunctional alveolar macrophage efferocytosis (phagocytic engulfment of apoptotic neutrophils), thus provoking an important inflammatory reaction as the result of subsequent neutrophil necrosis, which also leads to a severe clinical course, including death [121].

Previous research has established that TauCl stimulates the resolution of inflammation by boosting efferocytosis [122] and decreasing MPO production by neutrophils [75]. Macrophages secrete more IL-10, a cytokine that modulates excessive inflammatory reactions, and less of the cytokines that promote inflammation, like the TNF- α , IL-1, and IL-12 during efferocytosis [123]. A contemporary study demonstrated that sick people with COVID-19 had fewer airway regulatory T cells (Tregs) than healthy controls, reinforcing the hypothesis that a Treg deficit at the lung location is contributing to more severe illness [124]. Interestingly, taurine has been recently shown to increase the generation of CD4⁺CD25⁺FOXP3⁺ Tregs [125].

TauCl stimulates lactoferrin release

Acute inflammation is the body's active reaction to microbial pathogens and physical trauma; when neutrophils are drawn to an inflamed region, they get active and experience an oxidative burst, which is a key event in the protective immunity. As a result, there are abundant reactive oxygen species, of the kind that neutrophil leucocytes use to destroy and remove pathogenic microorganisms [122]. The major molecule MPO, second molecule lactoferrin (LF), and third molecule matrix metalloproteinase (MMP)-9 are discharged to the fluid that surrounds cells during degranulation [75].

Neutrophil LF degranulation is also induced by TauCl [75]. LF is a glycoprotein present in a variety of excretory fluids from mammals, and within neutrophil secondary granules [126]. LF exhibits antibacterial and antiviral effects on a wide range of pathogens, including several kinds of bacteria, as well as encapsulated and non-encapsulated viruses [127–129]. LF is an antiviral molecule that works mostly during the initial stage of infection but also at the internal cellular level (for example, during hepatitis C infection) [130–132]. According to the generally accepted framework, LF can prevent SARS-CoV-2 infection of permissive cells by inhibiting the spike protein attachment, adhesion to ACE2 receptors, or by exerting a competitive inhibition with molecules such as heparan sulfate and sialoside glycosaminoglycans, or by impairing viral replication. LF may be able to inhibit the triggering of the cytokine storm cascade, preventing systemic effects and illness worsening, due to its high ability to penetrate cells and get into the nucleus [126].

Discussion

If adequate therapy is not given at the proper moment, the COVID-19 disease can be lethal. According to research findings, "approximately 20% of COVID-19 patients tend to have a severe or critical disease, with a mortality rate of 50% or more in critical cases" [133–135]. The polarization (change from the M2 to the M1) of alveolar macrophages caused by SARS-CoV-2 could disrupt efferocytosis, preventing neutrophil death and perpetuating the inflammatory process [121, 136]. It has been postulated that much of the damage to lung cells in complicated COVID-19 cases is associated with an increase in HOCl generation by neutrophils and M1 macrophages rather than by the virus itself [73]. Taurine is a powerful scavenger for HOCl, and so neutralizes its harmful effects, according to compelling data [100]. Taurine also restricts the generation of all pro-inflammatory molecules linked to the cytokine storm, prevents damage to the lungs by decreasing oxidative stress, and promotes the resolution of the inflammatory process [98–101]. Neutrophil LF degranulation driven by taurine may have direct antiviral actions against SARS-CoV-2, suppressing viral replication [126].

Several studies have reported that taurine levels in mild/moderate COVID-19 patients are usually higher during active disease compared to healthy subjects [137], while others have found decreased taurine levels in asymptomatic, mild, moderate, severe, and critical patients [138]. The latter authors proposed that because taurine is abundant in leucocytes, a lack of it is thought to impair immune cell activity, thus in COVID-19 physiopathology, sepsis is comparable to the cytokine storm, and the low taurine level in this study could be one of the causes of the aberrant immune reaction in COVID-19 sufferers [138]. Our explanation of the different taurine levels found in COVID-19 patients is that during the acute phase, taurine levels increase as a response directed at trying to neutralize excessive oxidative damage. Taurine concentration in organs is derived from two sources: biosynthesis and diet. Most severely affected patients do not feed correctly, so the taurine's external supply is compromised. Furthermore, it has been reported that hyperglycemia downregulates the human taurine transporter [139]. Interestingly, a group of researchers discovered that an increased glucose concentration significantly promoted viral proliferation and inflammatory cytokine production, demonstrating the essential role of glycolysis. These findings can explain why the uncontrolled serum glucose levels in diabetes patients are a determinant factor for complicated COVID-19 cases [140]. So, during the severe and critical phases, taurine levels could be depleted by SARS-CoV-2-induced hyperglycemia, thus explaining the decreased taurine levels found in several studies. During the recovery phase, inflammation is controlled and taurine levels gradually increase, except in long COVID-19 patients.

Based on the data reported in this review, we propose that taurine may be able to prevent the cytokine storm from occurring if administered at the early COVID-19 stages. Interestingly, taurine derivatives like taurolidine reduce inflammation in peripheral blood mononuclear cells by inhibiting the release of the proinflammatory cytokines TNF- α , IL-1 β , IL-6, and IL-8; and a previous work postulated that taurine derivatives should be considered a promising available treatment strategy in the management of COVID-19 patients [141]. To test the validity of this hypothesis, clinical trials are required.

Abbreviations

ARDS: acute respiratory distress syndrome COVID-19: coronavirus disease 2019 Hb: hemoglobin HOCI: hypochlorous acid ICU: intensive care unit IL-6: interleukin 6 LF: lactoferrin LPS: lipopolysaccharide MPO: myeloperoxidase NETs: neutrophil extracellular traps NF-κB: nuclear factor-kappa B NLR: neutrophil-to-lymphocyte ratio SARS-CoV-2: severe acute respiratory syndrome coronavirus 2 TauCl: taurine chloramine TNF-α: tumor necrosis factor-alpha Tregs: regulatory T cells

Declarations

Author contributions

ARC and EMR contributed to conceptualization and project design. ARC, RCG and RB contributed to the literature analysis. VNU, EMR and RB contributed to the acquisition and interpretation of data, data curation, formal analysis, writing, and review of the manuscript. All authors have read and agreed to the published version of the manuscript.

Conflicts of interest

The authors declare they have no conflicts of interest.

Ethical approval Not applicable.

Consent to participate Not applicable. Consent to publication

Not applicable. Availability of data and materials

Not applicable.

Funding Not applicable.

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References

- Chen N, Zhou M, Dong X, Qu J, Gong F, Han Y, et al. Epidemiological and clinical characteristics of 99 cases of 2019 novel coronavirus pneumonia in Wuhan, China: a descriptive study. Lancet. 2020;395:507–13.
- 2. Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. Lancet. 2020;395:497–506.
- Coronavirus disease (COVID-19) pandemic [Internet]. Geneva: World Health Organization; c2022 [cited 2022 Apr 04]. Available from: https://www.who.int/emergencies/diseases/novelcoronavirus-2019
- 4. Choi WY. Mortality rate of patients with COVID-19 based on underlying health conditions. Disaster Med Public Health Prep. 2021;139:1–6.
- 5. Bertsimas D, Lukin G, Mingardi L, Nohadani O, Orfanoudaki A, Stellato B, et al.; Hellenic COVID-19 Study Group. COVID-19 mortality risk assessment: an international multi-center study. PLoS One. 2020;15:e0243262.
- 6. Clark A, Jit M, Warren-Gash C, Guthrie B, Wang HHX, Mercer SW, et al.; Centre for the Mathematical Modelling of Infectious Diseases COVID-19 working group. Global, regional, and national estimates of the population at increased risk of severe COVID-19 due to underlying health conditions in 2020: a modelling study. Lancet Glob Health. 2020;8:e1003–17.
- 7. Wu C, Chen X, Cai Y, Xia J, Zhou X, Xu S, et al. Risk factors associated with acute respiratory distress syndrome and death in patients with coronavirus disease 2019 pneumonia in Wuhan, China. JAMA Intern Med. 2020;180:934–43.
- 8. Qin C, Zhou L, Hu Z, Zhang S, Yang S, Tao Y, et al. Dysregulation of immune response in patients with coronavirus 2019 (COVID-19) in Wuhan, China. Clin Infect Dis. 2020;71:762–8.
- 9. Sinha P, Matthay MA, Calfee CS. Is a "cytokine storm" relevant to COVID-19? JAMA Intern Med. 2020;180:1152–4.
- 10. Abdin SM, Elgendy SM, Alyammahi SK, Alhamad DW, Omar HA. Tackling the cytokine storm in COVID-19, challenges and hopes. Life Sci. 2020;257:118054.
- 11. de la Rica R, Borges M, Gonzalez-Freire M. COVID-19: in the eye of the cytokine storm. Front Immunol. 2020;11:558898.
- 12. Fara A, Mitrev Z, Rosalia RA, Assas BM. Cytokine storm and COVID-19: a chronicle of pro-inflammatory cytokines. Open Biol. 2020;10:200160.
- 13. Garg S, Garg M, Prabhakar N, Malhotra P, Agarwal R. Unraveling the mystery of Covid-19 cytokine storm: from skin to organ systems. Dermatol Ther. 2020;33:e13859.
- 14. Hojyo S, Uchida M, Tanaka K, Hasebe R, Tanaka Y, Murakami M, et al. How COVID-19 induces cytokine storm with high mortality. Inflamm Regen. 2020;40:37.
- 15. Lin SH, Zhao YS, Zhou DX, Zhou FC, Xu F. Coronavirus disease 2019 (COVID-19): cytokine storms, hyper-inflammatory phenotypes, and acute respiratory distress syndrome. Genes Dis. 2020;7:520–7.
- 16. Moradian N, Gouravani M, Salehi MA, Heidari A, Shafeghat M, Hamblin MR, et al. Cytokine release syndrome: inhibition of pro-inflammatory cytokines as a solution for reducing COVID-19 mortality. Eur Cytokine Netw. 2020;31:81–93.
- 17. Nile SH, Nile A, Qiu J, Li L, Jia X, Kai G. COVID-19: pathogenesis, cytokine storm and therapeutic potential of interferons. Cytokine Growth Factor Rev. 2020;53:66–70.
- 18. Peter AE, Sandeep BV, Rao BG, Kalpana VL. Calming the storm: natural immunosuppressants as adjuvants to target the cytokine storm in COVID-19. Front Pharmacol. 2021;11:583777.
- 19. Quirch M, Lee J, Rehman S. Hazards of the cytokine storm and cytokine-targeted therapy in patients with COVID-19: review. J Med Internet Res. 2020;22:e20193.

- 20. Ragab D, Salah Eldin H, Taeimah M, Khattab R, Salem R. The COVID-19 cytokine storm; what we know so far. Front Immunol. 2020;11:1446.
- 21. Tang Y, Liu J, Zhang D, Xu Z, Ji J, Wen C. Cytokine storm in COVID-19: the current evidence and treatment strategies. Front Immunol. 2020;11:1708.
- 22. Ye Q, Wang B, Mao J. The pathogenesis and treatment of the 'cytokine storm' in COVID-19. J Infect. 2020;80:607–13.
- 23. Aslani M, Mortazavi-Jahromi SS, Mirshafiey A. Cytokine storm in the pathophysiology of COVID-19: possible functional disturbances of miRNAs. Int Immunopharmacol. 2021;101:108172.
- 24. Caricchio R, Gallucci M, Dass C, Zhang X, Gallucci S, Fleece D, et al. Temple University COVID-19 Research Group. Preliminary predictive criteria for COVID-19 cytokine storm. Ann Rheum Dis. 2021;80:88–95.
- 25. Chong ZZ, Souayah N. SARS-CoV-2 induced neurological manifestations entangles cytokine storm that implicates for therapeutic strategies. Curr Med Chem. 2022;29:2051–74.
- 26. Hanna J, Tipparaju P, Mulherkar T, Lin E, Mischley V, Kulkarni R, et al. Risk factors associated with the clinical outcomes of COVID-19 and its variants in the context of cytokine storm and therapeutics/vaccine development challenges. Vaccines (Basel). 2021;9:938.
- 27. Kaur S, Bansal R, Kollimuttathuillam S, Gowda AM, Singh B, Mehta D, et al. The looming storm: blood and cytokines in COVID-19. Blood Rev. 2021;46:100743.
- 28. Mehta P, Fajgenbaum DC. Is severe COVID-19 a cytokine storm syndrome: a hyperinflammatory debate. Curr Opin Rheumatol. 2021;33:419–30.
- 29. Melo AKG, Milby KM, Caparroz ALMA, Pinto ACPN, Santos RRP, Rocha AP, et al. Biomarkers of cytokine storm as red flags for severe and fatal COVID-19 cases: a living systematic review and meta-analysis. PLoS One. 2021;16:e0253894.
- 30. Nidadavolu LS, Walston JD. Underlying vulnerabilities to the cytokine storm and adverse COVID-19 outcomes in the aging immune system. J Gerontol A Biol Sci Med Sci. 2021;76:e13–8.
- 31. Premkumar M, Kedarisetty CK. Cytokine storm of COVID-19 and its impact on patients with and without chronic liver disease. J Clin Transl Hepatol. 2021;9:256–64.
- 32. Zhao Z, Wei Y, Tao C. An enlightening role for cytokine storm in coronavirus infection. Clin Immunol. 2021;222:108615.
- 33. Jiang Y, Rubin L, Peng T, Liu L, Xing X, Lazarovici P, et al. Cytokine storm in COVID-19: from viral infection to immune responses, diagnosis and therapy. Int J Biol Sci. 2022;18:459–72.
- 34. Kumar A, Sharma A, Tirpude NV, Sharma S, Padwad YS, Kumar S. Pharmaco-immunomodulatory interventions for averting cytokine storm-linked disease severity in SARS-CoV-2 infection. Inflammopharmacology. 2022;30:23–49.
- 35. Wahid B, Rani N, Idrees M. Cytokine storm syndrome in SARS-CoV-2: a review. Z Naturforsch C J Biosci. 2021;77:65–9.
- 36. Xu R, Liu C, Xu X, Hu Y, Zhu B, Yang C. Role of cytokine storm in coronavirus infections: culprit or accomplice? Front Biosci (Landmark Ed). 2022;27:102.
- 37. Zanza C, Romenskaya T, Manetti AC, Franceschi F, La Russa R, Bertozzi G, et al. Cytokine storm in COVID-19: immunopathogenesis and therapy. Medicina (Kaunas). 2022;58:144.
- 38. Yang L, Xie X, Tu Z, Fu J, Xu D, Zhou Y. The signal pathways and treatment of cytokine storm in COVID-19. Signal Transduct Target Ther. 2021;6:255.
- 39. Sun X, Wang T, Cai D, Hu Z, Chen J, Liao H, et al. Cytokine storm intervention in the early stages of COVID-19 pneumonia. Cytokine Growth Factor Rev. 2020;53:38–42.
- 40. Shimabukuro-Vornhagen A, Gödel P, Subklewe M, Stemmler HJ, Schlöβer HA, Schlaak M, et al. Cytokine release syndrome. J Immunother Cancer. 2018;6:56.

- 41. Tanaka T, Narazaki M, Kishimoto T. Immunotherapeutic implications of IL-6 blockade for cytokine storm. Immunotherapy. 2016;8:959–70.
- 42. Hunter CA, Jones SA. IL-6 as a keystone cytokine in health and disease. Nat Immunol. 2015;16:448–57.
- 43. Pathan N, Hemingway CA, Alizadeh AA, Stephens AC, Boldrick JC, Oragui EE, et al. Role of interleukin 6 in myocardial dysfunction of meningococcal septic shock. Lancet. 2004;363:203–9.
- 44. Hay KA, Hanafi LA, Li D, Gust J, Liles WC, Wurfel MM, et al. Kinetics and biomarkers of severe cytokine release syndrome after CD19 chimeric antigen receptor-modified T-cell therapy. Blood. 2017;130:2295–306.
- 45. Wang D, Hu B, Hu C, Zhu F, Liu X, Zhang J, et al. Clinical characteristics of 138 hospitalized patients with 2019 novel coronavirus-infected pneumonia in Wuhan, China. JAMA. 2020;323:1061–9.
- 46. Teijaro JR, Walsh KB, Rice S, Rosen H, Oldstone MB. Mapping the innate signaling cascade essential for cytokine storm during influenza virus infection. Proc Natl Acad Sci U S A. 2014;111:3799–804.
- 47. Jain U. Effect of COVID-19 on the organs. Cureus. 2020;12:e9540.
- 48. Sardu C, D'Onofrio N, Balestrieri ML, Barbieri M, Rizzo MR, Messina V, et al. Outcomes in patients with hyperglycemia affected by COVID-19: can we do more on glycemic control? Diabetes Care. 2020;43:1408–15.
- 49. Sardu C, Gargiulo G, Esposito G, Paolisso G, Marfella R. Impact of diabetes mellitus on clinical outcomes in patients affected by Covid-19. Cardiovasc Diabetol. 2020;19:76.
- 50. Marfella R, D'Onofrio N, Sardu C, Scisciola L, Maggi P, Coppola N, et al. Does poor glycaemic control affect the immunogenicity of the COVID-19 vaccination in patients with type 2 diabetes: the CAVEAT study. Diabetes Obes Metab. 2022;24:160–5.
- 51. Choudry FA, Hamshere SM, Rathod KS, Akhtar MM, Archbold RA, Guttmann OP, et al. High thrombus burden in patients with COVID-19 presenting with ST-segment elevation myocardial infarction. J Am Coll Cardiol. 2020;76:1168–76.
- 52. D'Onofrio N, Scisciola L, Sardu C, Trotta MC, De Feo M, Maiello C, et al. Glycated ACE2 receptor in diabetes: open door for SARS-COV-2 entry in cardiomyocyte. Cardiovasc Diabetol. 2021;20:99.
- 53. Kim C, Cha YN. Taurine chloramine produced from taurine under inflammation provides anti-inflammatory and cytoprotective effects. Amino Acids. 2014;46:89–100.
- 54. Qaradakhi T, Gadanec LK, McSweeney KR, Abraham JR, Apostolopoulos V, Zulli A. The anti-inflammatory effect of taurine on cardiovascular disease. Nutrients. 2020;12:2847.
- 55. Goc Z, Kapusta E, Formicki G, Martiniaková M, Omelka R. Effect of taurine on ethanol-induced oxidative stress in mouse liver and kidney. Chin J Physiol. 2019;62:148–56.
- 56. Niu X, Zheng S, Liu H, Li S. Protective effects of taurine against inflammation, apoptosis, and oxidative stress in brain injury. Mol Med Rep. 2018;18:4516–22.
- 57. Chesney RW, Han X, Patters AB. Taurine and the renal system. J Biomed Sci. 2010;17 Suppl 1:S4.
- 58. Kim SJ, Gupta RC, Lee HW. Taurine-diabetes interaction: from involvement to protection. Curr Diabetes Rev. 2007;3:165–75.
- 59. Chua RL, Lukassen S, Trump S, Hennig BP, Wendisch D, Pott F, et al. COVID-19 severity correlates with airway epithelium-immune cell interactions identified by single-cell analysis. Nat Biotechnol. 2020;38:970–9.
- 60. Liao M, Liu Y, Yuan J, Wen Y, Xu G, Zhao J, et al. Single-cell landscape of bronchoalveolar immune cells in patients with COVID-19. Nat Med. 2020;26:842–4.
- 61. Guan WJ, Ni ZY, Hu Y, Liang WH, Ou CQ, He JX, et al.; China Medical Treatment Expert Group for Covid-19. Clinical characteristics of coronavirus disease 2019 in China. N Engl J Med. 2020;382:1708–20.
- 62. Meizlish ML, Pine AB, Bishai JD, Goshua G, Nadelmann ER, Simonov M, et al. A neutrophil activation signature predicts critical illness and mortality in COVID-19. Blood Adv. 2021;5:1164–77.

- 63. Aschenbrenner AC, Mouktaroudi M, Krämer B, Oestreich M, Antonakos N, Nuesch-Germano M, et al.; German COVID-19 Omics Initiative (DeCOI). Disease severity-specific neutrophil signatures in blood transcriptomes stratify COVID-19 patients. Genome Med. 2021;13:7.
- 64. Lagunas-Rangel FA. Neutrophil-to-lymphocyte ratio and lymphocyte-to-C-reactive protein ratio in patients with severe coronavirus disease 2019 (COVID-19): a meta-analysis. J Med Virol. 2020;92:1733–4.
- 65. Sun S, Cai X, Wan H, He G, Lin Y, Lu B, et al. Abnormalities of peripheral blood system in patients with COVID-19 in Wenzhou, China. Clin Chim Acta. 2020;507:174–80.
- 66. Barnes BJ, Adrover JM, Baxter-Stoltzfus A, Borczuk A, Cools-Lartigue J, Crawford JM, et al. Targeting potential drivers of COVID-19: neutrophil extracellular traps. J Exp Med. 2020;217:e20200652.
- 67. Klebanoff SJ. Myeloperoxidase-halide-hydrogen peroxide antibacterial system. J Bacteriol. 1968;95:2131–8.
- 68. Klebanoff SJ. Myeloperoxidase: friend and foe. J Leukoc Biol. 2005;77:598–625.
- 69. Winterbourn CC, van den Berg JJ, Roitman E, Kuypers FA. Chlorohydrin formation from unsaturated fatty acids reacted with hypochlorous acid. Arch Biochem Biophys. 1992;296:547–55.
- 70. Schieven GL, de Fex H, Stephenson L. Hypochlorous acid activates tyrosine phosphorylation signal pathways leading to calcium signaling and TNFalpha production. Antioxid Redox Signal. 2002;4:501–7.
- 71. Schoonbroodt S, Legrand-Poels S, Best-Belpomme M, Piette J. Activation of the NF-kappaB transcription factor in a T-lymphocytic cell line by hypochlorous acid. Biochem J. 1997;321:777–85.
- 72. Xie J, Tong Z, Guan X, Du B, Qiu H. Clinical characteristics of patients who died of coronavirus disease 2019 in China. JAMA Netw Open. 2020;3:e205619.
- 73. Goud PT, Bai D, Abu-Soud HM. A multiple-hit hypothesis involving reactive oxygen species and myeloperoxidase explains clinical deterioration and fatality in COVID-19. Int J Biol Sci. 2021;17:62–72.
- 74. Maitra D, Byun J, Andreana PR, Abdulhamid I, Diamond MP, Saed GM, et al. Reaction of hemoglobin with HOCl: mechanism of heme destruction and free iron release. Free Radic Biol Med. 2011;51:374–86.
- 75. Kim DG, Kwon YM, Kang IS, Kim C. Taurine chloramine selectively regulates neutrophil degranulation through the inhibition of myeloperoxidase and upregulation of lactoferrin. Amino Acids. 2020;52:1191–9.
- 76. Twaddell SH, Baines KJ, Grainge C, Gibson PG. The emerging role of neutrophil extracellular traps in respiratory disease. Chest. 2019;156:774–82.
- 77. Porto BN, Stein RT. Neutrophil extracellular traps in pulmonary diseases: too much of a good thing? Front Immunol. 2016;7:311.
- 78. Potey PM, Rossi AG, Lucas CD, Dorward DA. Neutrophils in the initiation and resolution of acute pulmonary inflammation: understanding biological function and therapeutic potential. J Pathol. 2019;247:672–85.
- 79. Frantzeskaki F, Armaganidis A, Orfanos SE. Immunothrombosis in acute respiratory distress syndrome: cross talks between inflammation and coagulation. Respiration. 2017;93:212–25.
- 80. Klok FA, Kruip MJHA, van der Meer NJM, Arbous MS, Gommers DAMPJ, Kant KM, et al. Incidence of thrombotic complications in critically ill ICU patients with COVID-19. Thromb Res. 2020;191:145–7.
- 81. Veras FP, Pontelli MC, Silva CM, Toller-Kawahisa JE, de Lima M, Nascimento DC, et al. SARS-CoV-2-triggered neutrophil extracellular traps mediate COVID-19 pathology. J Exp Med. 2020;217:e20201129.
- 82. Zuo Y, Yalavarthi S, Shi H, Gockman K, Zuo M, Madison JA, et al. Neutrophil extracellular traps in COVID-19. JCI Insight. 2020;5:e138999.
- 83. Zuo Y, Yalavarthi S, Navaz S, Hoy C, Harbaugh A, Gockman K, et al. Autoantibodies stabilize neutrophil extracellular traps in COVID-19. JCI Insight. 2021;6:e150111.

- 84. Narasaraju T, Yang E, Samy RP, Ng HH, Poh WP, Liew AA, et al. Excessive neutrophils and neutrophil extracellular traps contribute to acute lung injury of influenza pneumonitis. Am J Pathol. 2011;179:199–210.
- 85. Kopf M, Schneider C, Nobs SP. The development and function of lung-resident macrophages and dendritic cells. Nat Immunol. 2015;16:36–44.
- 86. Yamasaki K, Eeden SFV. Lung macrophage phenotypes and functional responses: role in the pathogenesis of COPD. Int J Mol Sci. 2018;19:582.
- 87. Hussell T, Bell TJ. Alveolar macrophages: plasticity in a tissue-specific context. Nat Rev Immunol. 2014;14:81–93.
- 88. Desch AN, Gibbings SL, Goyal R, Kolde R, Bednarek J, Bruno T, et al. Flow cytometric analysis of mononuclear phagocytes in nondiseased human lung and lung-draining lymph nodes. Am J Respir Crit Care Med. 2016;193:614–26.
- 89. Lv J, Wang Z, Qu Y, Zhu H, Zhu Q, Tong W, et al. Distinct uptake, amplification, and release of SARS-CoV-2 by M1 and M2 alveolar macrophages. Cell Discov. 2021;7:24.
- 90. Lawrence T, Natoli G. Transcriptional regulation of macrophage polarization: enabling diversity with identity. Nat Rev Immunol. 2011;11:750–61.
- 91. Kalliolias GD, Ivashkiv LB. TNF biology, pathogenic mechanisms and emerging therapeutic strategies. Nat Rev Rheumatol. 2016;12:49–62.
- 92. Kambara K, Ohashi W, Tomita K, Takashina M, Fujisaka S, Hayashi R, et al. *In vivo* depletion of CD206⁺ M2 macrophages exaggerates lung injury in endotoxemic mice. Am J Pathol. 2015;185:162–71.
- 93. Boumaza A, Gay L, Mezouar S, Bestion E, Diallo AB, Michel M, et al. Monocytes and macrophages, targets of severe acute respiratory syndrome coronavirus 2: the clue for coronavirus disease 2019 immunoparalysis. J Infect Dis. 2021;224:395–406.
- 94. Rodrigues MR, Rodriguez D, Russo M, Campa A. Macrophage activation includes high intracellular myeloperoxidase activity. Biochem Biophys Res Commun. 2002;292:869–73.
- 95. Hawkins CL, Pattison DI, Davies MJ. Hypochlorite-induced oxidation of amino acids, peptides and proteins. Amino Acids. 2003;25:259–74.
- 96. Kawai Y, Morinaga H, Kondo H, Miyoshi N, Nakamura Y, Uchida K, et al. Endogenous formation of novel halogenated 2'-deoxycytidine. Hypohalous acid-mediated DNA modification at the site of inflammation. J Biol Chem. 2004;279:51241–9.
- 97. Woods CG, Fu J, Xue P, Hou Y, Pluta LJ, Yang L, et al. Dose-dependent transitions in Nrf2-mediated adaptive response and related stress responses to hypochlorous acid in mouse macrophages. Toxicol Appl Pharmacol. 2009;238:27–36.
- 98. Quinn MR, Barua M, Liu Y, Serban V. Taurine chloramine inhibits production of inflammatory mediators and iNOS gene expression in alveolar macrophages; a tale of two pathways: part I, NF-kappaB signaling. Adv Exp Med Biol. 2003;526:341–8.
- 99. Barua M, Liu Y, Quinn MR. Taurine chloramine inhibits inducible nitric oxide synthase and TNF-alpha gene expression in activated alveolar macrophages: decreased NF-kappaB activation and IkappaB kinase activity. J Immunol. 2001;167:2275–81.
- 100. Marcinkiewicz J, Grabowska A, Bereta J, Stelmaszynska T. Taurine chloramine, a product of activated neutrophils, inhibits *in vitro* the generation of nitric oxide and other macrophage inflammatory mediators. J Leukoc Biol. 1995;58:667–74.
- 101. Schuller-Levis G, Quinn MR, Wright C, Park E. Taurine protects against oxidant-induced lung injury: possible mechanism(s) of action. Adv Exp Med Biol. 1994;359:31–9.
- 102. McGrath MS, Kodelja V. Balanced macrophage activation hypothesis: a biological model for development of drugs targeted at macrophage functional states. Pathobiology. 1999;67:277–81.

- 103. Meng L, Lu C, Wu B, Lan C, Mo L, Chen C, et al. Taurine antagonizes macrophages M1 polarization by mitophagy-glycolysis switch blockage via dragging SAM-PP2Ac transmethylation. Front Immunol. 2021;12:648913.
- 104. Kettle AJ, Albrett AM, Chapman AL, Dickerhof N, Forbes LV, Khalilova I, et al. Measuring chlorine bleach in biology and medicine. Biochim Biophys Acta. 2014;1840:781–93.
- 105. Giese T, McGrath MS, Stumm S, Schempp H, Elstner E, Meuer SC. Differential effects on innate *versus* adaptive immune responses by WF10. Cell Immunol. 2004;229:149–58.
- 106. Schempp H, Reim M, Dornisch, K, Elstner EF. Chlorite-hemoprotein interaction as key role for the pharmacological activity of the chlorite-based drug WF10. Arzneimittelforschung. 2001;51:554–62.
- 107. Weiss SJ, Klein R, Slivka A, Wei M. Chlorination of taurine by human neutrophils. Evidence for hypochlorous acid generation. J Clin Invest. 1982;70:598–607.
- 108. Cantin AM. Taurine modulation of hypochlorous acid-induced lung epithelial cell injury *in vitro*. Role of anion transport. J Clin Invest. 1994;93:606–14.
- 109. Sun Jang J, Piao S, Cha YN, Kim C. Taurine chloramine activates Nrf2, increases HO-1 expression and protects cells from death caused by hydrogen peroxide. J Clin Biochem Nutr. 2009;45:37–43.
- 110. Marcinkiewicz J, Grabowska A, Bereta J, Bryniarski K, Nowak B. Taurine chloramine down-regulates the generation of murine neutrophil inflammatory mediators. Immunopharmacology. 1998;40:27–38.
- 111. Kontny E, Szczepańska K, Kowalczewski J, Kurowska M, Janicka I, Marcinkiewicz J, et al. The mechanism of taurine chloramine inhibition of cytokine (interleukin-6, interleukin-8) production by rheumatoid arthritis fibroblast-like synoviocytes. Arthritis Rheum. 2000;43:2169–77.
- 112. Kanayama A, Inoue J, Sugita-Konishi Y, Shimizu M, Miyamoto Y. Oxidation of Ikappa Balpha at methionine 45 is one cause of taurine chloramine-induced inhibition of NF-kappa B activation. J Biol Chem. 2002;277:24049–56.
- 113. Li S, Wang J, Wei BK, Dong G, Wang X. Protective effect of taurine on paraquat-induced lung epithelial cell injury. Adv Exp Med Biol. 2019;1155:739–46.
- 114. Zhao W, Jia L, Yang HJ, Xue X, Xu WX, Cai JQ, et al. Taurine enhances the protective effect of Dexmedetomidine on sepsis-induced acute lung injury via balancing the immunological system. Biomed Pharmacother. 2018;103:1362–8.
- 115. Men X, Han S, Gao J, Cao G, Zhang L, Yu H, et al. Taurine protects against lung damage following limb ischemia reperfusion in the rat by attenuating endoplasmic reticulum stress-induced apoptosis. Acta Orthop. 2010;81:263–7.
- 116. Chen J, Xue X, Cai J, Jia L, Sun B, Zhao W. Protective effect of taurine on sepsis-induced lung injury via inhibiting the p38/MAPK signaling pathway. Mol Med Rep. 2021;24:653.
- 117. Mei SH, McCarter SD, Deng Y, Parker CH, Liles WC, Stewart DJ. Prevention of LPS-induced acute lung injury in mice by mesenchymal stem cells overexpressing angiopoietin 1. PLoS Med. 2007;4:e269.
- 118. Caroff M, Karibian D. Structure of bacterial lipopolysaccharides. Carbohydr Res. 2003;338:2431–47.
- 119. Bhavsar TM, Patel SN, Lau-Cam CA. Protective action of taurine, given as a pretreatment or as a posttreatment, against endotoxin-induced acute lung inflammation in hamsters. J Biomed Sci. 2010;17 Suppl 1:S19.
- 120. El Kebir D, József L, Pan W, Filep JG. Myeloperoxidase delays neutrophil apoptosis through CD11b/CD18 integrins and prolongs inflammation. Circ Res. 2008;103:352–9.
- 121. Mahida RY, Scott A, Parekh D, Lugg ST, Hardy RS, Lavery GG, et al. Acute respiratory distress syndrome is associated with impaired alveolar macrophage efferocytosis. Eur Respir J. 2021;58:2100829.
- 122. Kim W, Kim HU, Lee HN, Kim SH, Kim C, Cha YN, et al. Taurine chloramine stimulates efferocytosis through upregulation of Nrf2-mediated heme oxygenase-1 expression in murine macrophages: possible involvement of carbon monoxide. Antioxid Redox Signal. 2015;23:163–77.

- 123. Voll RE, Herrmann M, Roth EA, Stach C, Kalden JR, Girkontaite I. Immunosuppressive effects of apoptotic cells. Nature. 1997;390:350–1.
- 124. Szabo PA, Dogra P, Gray JI, Wells SB, Connors TJ, Weisberg SP, et al. Longitudinal profiling of respiratory and systemic immune responses reveals myeloid cell-driven lung inflammation in severe COVID-19. Immunity. 2021;54:797–814.e6.
- 125. Zhou J, Lu Y, Wu W, Feng Y. Taurine promotes the production of CD4⁺CD25⁺FOXP3⁺ Treg cells through regulating IL-35/STAT1 pathway in a mouse allergic rhinitis model. Allergy Asthma Clin Immunol. 2021;17:59.
- 126. Mattar EH, Elrashdy F, Almehdar HA, Uversky VN, Redwan EM. Natural resources to control COVID-19: could lactoferrin amend SARS-CoV-2 infectivity? PeerJ. 2021;9:e11303.
- 127. Redwan EM, Uversky VN, El-Fakharany EM, Al-Mehdar H. Potential lactoferrin activity against pathogenic viruses. C R Biol. 2014;337:581–95.
- 128. Albar AH, Almehdar HA, Uversky VN, Redwan EM. Structural heterogeneity and multifunctionality of lactoferrin. Curr Protein Pept Sci. 2014;15:778–97.
- 129. Redwan EM, El-Fakharany EM, Uversky VN, Linjawi MH. Screening the anti infectivity potentials of native N- and C-lobes derived from the camel lactoferrin against hepatitis C virus. BMC Complement Altern Med. 2014;14:219.
- 130. El-Fakharany EM, Sánchez L, Al-Mehdar HA, Redwan EM. Effectiveness of human, camel, bovine and sheep lactoferrin on the hepatitis C virus cellular infectivity: comparison study. Virol J. 2013;10:199.
- 131. Liao Y, El-Fakkarany E, Lönnerdal B, Redwan EM. Inhibitory effects of native and recombinant full-length camel lactoferrin and its N and C lobes on hepatitis C virus infection of Huh7.5 cells. J Med Microbiol. 2012;61:375–83.
- 132. Redwan el-RM, Tabll A. Camel lactoferrin markedly inhibits hepatitis C virus genotype 4 infection of human peripheral blood leukocytes. J Immunoassay Immunochem. 2007;28:267–77.
- 133. Richardson S, Hirsch JS, Narasimhan M, Crawford JM, McGinn T, Davidson KW; the Northwell COVID-19 Research Consortium; Barnaby DP, Becker LB, Chelico JD, Cohen SL, Cookingham J, Coppa K, et al. Presenting characteristics, comorbidities, and outcomes among 5,700 patients hospitalized with COVID-19 in the New York city area. JAMA. 2020;323:2052–9.
- 134. Zhang J, Wang X, Jia X, Li J, Hu K, Chen G, et al. Risk factors for disease severity, unimprovement, and mortality in COVID-19 patients in Wuhan, China. Clin Microbiol Infect. 2020;26:767–72.
- 135. Chen T, Wu D, Chen H, Yan W, Yang D, Chen G, et al. Clinical characteristics of 113 deceased patients with coronavirus disease 2019: retrospective study. BMJ. 2020;368:m1091.
- 136. Mosser DM, Edwards JP. Exploring the full spectrum of macrophage activation. Nat Rev Immunol. 2008;8:958–69.
- 137. Singh Y, Trautwein C, Fendel R, Krickeberg N, Berezhnoy G, Bissinger R, et al.; Deutsche COVID-19 OMICS Initiate (DeCOI). SARS-CoV-2 infection paralyzes cytotoxic and metabolic functions of the immune cells. Heliyon. 2021;7:e07147.
- 138. Atila A, Alay H, Yaman ME, Akman TC, Cadirci E, Bayrak B, et al. The serum amino acid profile in COVID-19. Amino Acids. 2021;53:1569–88.
- 139. Stevens MJ, Hosaka Y, Masterson JA, Jones SM, Thomas TP, Larkin DD. Downregulation of the human taurine transporter by glucose in cultured retinal pigment epithelial cells. Am J Physiol. 1999;277:E760–71.
- 140. Codo AC, Davanzo GG, Monteiro LB, de Souza GF, Muraro SP, Virgilio-da-Silva JV, et al. Elevated glucose levels favor SARS-CoV-2 infection and monocyte response through a HIF- 1α /glycolysis-dependent axis. Cell Metab. 2020;32:437–46.e5.
- 141. Iwegbulem O, Wang J, Pfirrmann RW, Redmond HP. The role of taurine derivatives in the putative therapy of COVID-19-induced inflammation. Ir J Med Sci. 2022;191:485–6.