

#### **Open Access** Review



# Inflammatory bowel diseases: what lactoferrin can do for us?

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## Abstract

Lactoferrin (LF), an iron-binding protein, is found in mammalian milk. LF is also secreted by different cell phenotypes. LF shows a wide range of biological activities, as many preclinical and clinical studies indicate that LF and its derived peptides have many biological functions in host defence, including not only antibacterial, but also antiviral, antifungal, and antiparasitic effects. These results raise the view that these compounds might affect the composition of the intestinal microbiota. LF is generally recognized as safe (GRAS). This protein has been shown in experimental studies to exert beneficial effects on intestinal inflammation. This review will target the beneficial effects of oral LF supplements on the intestinal ecosystem during inflammation and highlight the mechanisms by which LF may contribute to reducing inflammatory flare, and present perspectives for future research.

## **Keywords**

Lactoferrin, oral supplementation, antimicrobial effects, inflammatory bowel diseases

## Introduction

Lactoferrin (LF) is a component of the milk whey protein of most mammals. LF, is a globular iron-binding glycoprotein that belongs to the transferrin family. LF is found in human breast milk at relatively high concentrations (2–3 g/L) [1, 2]. Bovine and human LF share sequence homology (69% amino acid sequence homology) and structure homology as judged from their three-dimensional structures [3]. LF is also found in many other fluids. Many cells within human tissues and organs can produce LF and presence of LF has been confirmed in kidneys, lungs, gallbladder, pancreas, intestine, liver, prostate, and cells of the immune system and in fluids such as saliva, tears, sperm, cerebrospinal fluid, urine, bronchial secretion, synovial fluid, umbilical cord blood, blood plasma, and vagina discharge [3]. LF, in the circulation of healthy individuals, is found in concentrations ranging between 2 and 7  $\mu$ g/mL [4]. Intact bovine milk-derived LF (bLF) can be absorbed by intestinal epithelial cells and enter the systemic circulation in mice and rats [5, 6]. Intact bLF can be recovered in the duodenal (~9.5%) and in the colonic luminal fluids (~7.2%) [7].

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LF mass from milk and colostrum is in the range of 83–87 kDa, while in neutrophils, LF mass is about 87–91 kDa [8]. Among the different physiological functions of LF, as will be presented in the following paragraphs, one of the most important is a direct antimicrobial role. In addition, LF shows effects on the intestinal epithelial cells. Furthermore, the LF's ability to modulate immune response and to protect against viral infections and septic shock has been described [9–14].

bLF appears to be safe for human consumption. Indeed, this compound was approved in 2012 by the European Food Safety Authority (EFSA) as a novel food ingredient [15]. Earlier, in 2001, bLF was considered generally recognized as safe (GRAS). In fact, bLF is GRAS for use as a food additive and dietary supplement in infants and adults by the Food and Drug Administration (FDA) [16]. bLF is currently added to several foods, including infant formulas, yoghurts, beverages, and dietary supplements [17, 18]. The aim of the present review is to present the experimental and clinical arguments that suggest that LF can be beneficial in situations of inflammatory flare, and to present some directions for future research in that field.

## LF and epithelial cells

As LF hydrolysis is minimal in infant intestinal content, oral administration of LF has been shown to improve growth and renewal of the intestinal epithelium of infants during development [19]. Experimental studies show the positive effects of LF on the intestine. LF modulates numerous parameters involved in the physiology of the intestinal epithelium. The modifications of such parameters include an increase in the intestinal epithelial differentiation, as judged from the measurement of the brush-border-associated enzymatic activities and amino acid transporters, an increase in the mouse jejunal villus height, and a decrease in epithelial cell spontaneous apoptosis [20, 21]. In Caco-2 cells, bLF decreases the expression of Cdck2 and increases the expression of TAF1, which regulates cellular proliferation and differentiation [20]. In rat pups, bLF accelerates intestinal maturation, increases expression of tight junction (TJ) proteins, of goblet cell marker mucins, and of several enzymes associated with the intestinal brush border membranes. However, LF ingestion was more efficient for increasing the expression of TJ proteins in the colon than in the ileum [6]. LF is internalized through clathrin-mediated endocytosis, and after entering inside the nucleus, LF increases thymidine incorporation in crypt cells and regulates the transcription of many genes [22]. Moreover, metabolic approaches indicated that higher plasma LF concentration is correlated to higher amino acid concentration, in particular essential ones, that favor protein synthesis, supporting an enhanced growth and maturation of the intestine and a potential therapeutic interest in post-natal development [6]. LF possesses antibacterial and immunomodulatory properties that play a role in the protection of the epithelial barrier. LF plays key roles in the regulation of the intestinal mucosal immune system [23, 24]. The intestinal epithelial cells include not only absorptive cells but also cells that participate in the immune activities, such as those involved in cytokines secretion [25, 26].

# LF partly resists digestion in the gastrointestinal tract and can be partly absorbed in intact form

LF can be administered by different routes: oral, intravenous, or local. LF digestion in the stomach by pepsin is limited. This limited digestion releases bioactive peptides, called collectively lactoferricin, which have been characterized for their antibiotic capacities [27]. LF-derived peptides have been studied for their biochemical properties of lactoferricin peptides may interact with lipidic compounds and negatively charged surfaces of both Gram-negative and Gram-positive bacteria, as well as with fungi, viruses, and parasites thus explaining presumably partly antibacterial, antifungal, antiviral, and antiparasitic activities of these peptides [28, 29]. LF was shown four decades ago to be not fully degraded by the exocrine enzymatic pancreatic protease activities [30], thus raising the view that a part of the ingested LF may remain in intact form in the small intestine luminal fluid. In newborns, LF is absorbed, but in adults, the LF its bioavailability is very low. However, LF can be absorbed in intact form by epithelial cells and enter the systemic circulation, but the exact percentage of LF that is absorbed remains unknown [31, 32]. As free LF

has a half-life of about 12–60 min in blood, many approaches to optimize oral delivery of LF are under development [33].

The presence of LF in the intestinal fluid and in blood coincides with effects of this protein on several tissues, including notably small and large intestine. The physiological effects of LF on target cells have been shown to involve the binding to specific receptors. The target cells equipped with receptors to LF include intestinal epithelial cells [33, 34], lymphocytes [35], and macrophages [36].

#### LF displays numerous effects on the intestinal ecosystem

The small and large intestines are well known to be inhabited by a complex mixture of microbes, among which bacteria have been the subject of most studies. The concentrations of bacteria increase from the proximal part of the small intestine to the distal part of the large intestine. In the large intestine, the bacterial concentration is in the range of  $10^9-10^{12}$  colony-forming unit (CFU) per g of content. The spectacular increase in the concentration of bacteria in the distal small and large intestine is notably due to a much slower transit of the intestinal content in the large intestine than in the small intestine, allowing intense metabolism of the available substrates supplied by the host [37]. This microbial population includes not only bacteria, but also other microorganisms are usually not considered members of the intestinal microbiota. They represent a heterogeneous group of living organisms, with several of them being classified as parasites [38].

As previously reported, LF and LF-derived lactoferricin have been demonstrated in numerous studies to exert antibacterial, antiviral, and antifungal, but also antiparasitic effects and to stimulate the immune response of intestinal epithelial cells. These effects are related to the effects of LF on specific immune response regulation [39–41]. Notably, LF can modulate cytokines such as interleukins (IL-1b, IL-6, IL-10, and IL-18) and tumor necrosis factor alpha (TNF- $\alpha$ ) production by intestinal immune cells [42, 43].

The functional role of LF in gastrointestinal inflammation is mainly attributed to its capacity to bind to iron, thus inhibiting the invasion or adhesion of bacteria. However, other mechanisms of action have been shown, LF can bind to LF receptors in intestinal mucosa and lymphatic cells to modulate the antimicrobial effects of LF, but also to specific receptors in bacteria [44] and to bacterial cell walls, leading to the disruption of bacteria integrity and accordingly contributing to bacterial death. Direct interaction with Gram-negative bacteria leading to cell membrane damage has been reported [45, 46]. Bovine LF retards the growth of the pathogenic bacteria *Clostridium difficile* in a model of bacterial infection [47]. This result is of major importance considering the high incidence of *Clostridium difficile* infection in individuals with inflammatory bowel diseases (IBDs). However, we do not know if specific bacteria can degrade LF, a situation which would render bacteria more resistant to the antibacterial effects of LF.

Concerning the antiviral effect of LF, this protein has been shown to be active on both DNA- and RNAviruses, including human rotaviruses [48, 49], which represent a major cause of diarrhea in children younger than five years [50]. The fungicidal activity of LF has been shown against various Candida species, including *Candida albicans*, a common fungus found within the human intestinal microbiota [51–53]. *Candida albicans* exacerbates intestinal inflammation in mice [54]. Lastly, the antiparasitic effect of LF has been demonstrated in vitro against the malaria parasite *Plasmodium falciparum* [55], which is involved in complex interaction with the gut microbes [56]. Lastly, microbicidal effects of LF-derived peptides have been demonstrated against the parasite *Entamoeba histolytica* [57]. This parasite infects humans primarily within the intestinal tract [58], provoking intestinal inflammation and ulceration in infected patients [59].

LF is also active on the host intestinal epithelial cells. In rodents, a model, as well as in an in vitro study with intestinal epithelial cells, LF has been shown to efficiently increase intestinal cell proliferation and differentiation [20]. Interestingly, LF ingestion by the mother during gestation and lactation was shown to promote early pup development in a rodent model [6]. This latter effect coincided in pups with modification of intestinal epithelial physiology. In fact, increased small intestine epithelial cell

differentiation as well as increased colon barrier function are recorded after LF supplementation. The increased epithelial cell differentiation was associated with increased expression of genes coding for the tight-junction proteins. In addition, in such a situation, LF supplementation was associated with higher plasma amino acid concentrations, maybe because of increased expression of amino acid transporters. This latter hypothesis needs to be experimentally tested to confirm or invalidate it.

## LF can be beneficial in situation of intestinal inflammation

IBDs are chronic inflammatory disorders affecting the gastrointestinal tract, with the two main forms being ulcerative colitis (UC) and Crohn's disease. IBD etiology is not fully understood, but the interactions among mucosal immune cells, barrier function, and commensal enteric flora involve both genetic and environmental factors (including dietary parameters) that presumably play significant roles [60]. IBD induced an increase in intestinal permeability, reduction of TJ protein expression, and overproduction of proinflammatory cytokines, followed by accumulation and activation of immune cells. LF may be of interest in individuals prone to IBDs since this protein acts as a protector of the intestinal barrier [61]. The disease activity coincides with increased secretion of pro-inflammatory cytokines, including notably TNF- $\alpha$ , IL-1, IL-6, and IL-8. These molecules play a crucial role in mediating the response to infection [62]. Many studies reported that LF orally administered can bind to LF receptors on intestinal cells and gut-associated lymphatic tissue to modulate cytokine production. Such binding is associated with the stimulation of the synthesis of anti-inflammatory cytokines, including IL-4 and IL-10, these proteins being considered potent anti-inflammatory and immunomodulating compounds involved in the protection of the mucosa against infections and inflammation [63, 64].

bLF oral administration in animal colitis model can diminish signs of intestinal inflammation, probably partly by modulating the immune system and by reducing the pro-inflammatory cytokine production and secretion in the colonic tissue. Furthermore, LF can increase the expression of TJ proteins [65–67]. LF has also been shown to activate the mitogen-activated protein kinase (MAPK) to promote cells proliferation and differentiation [68]. In addition, bLF and its derived peptides, when given orally, reduce signs of experimentally induced colitis in mice when given orally [69]. Indeed, in this latter experimental work, bLF reduces the presence of occult blood in feces and the number of TNF- $\alpha$ -producing cells in the distal colon. Oral delivery of a *Lactococcus lactis* strain secreting LF-derived peptides can diminish the development of colitis in mice [70]. Furthermore, in the model of colitis induced by dextran sulfate sodium (DSS), bLF was found to reduce inflammation and, accordingly, the impairment of colonic epithelial barrier function. These effects were paralleled by changes in the composition of the flora, such changes being hypothesized to improve the intestinal barrier regeneration [71].

The effects of LF supplementation have also been tested in the model of endotoxemia provoked by administration of lipopolysaccharide (LPS). This model is characterized by a systemic inflammatory response [72]. LPSs are glycolipids that are components of the bacterial surface. LPS produced by Gramnegative bacteria can be partly transferred from the luminal intestinal fluid to the bloodstream [73]. Within the intestine, LPS systemic administration has been shown to increase intestinal permeability, epithelial cell apoptosis and shedding, while provoking villus shortening and overall diarrhea [74–77]. These effects of LPS are associated with several intestinal dysfunctions, such as decreased mucosal oxygen consumption and amino acid absorption [78, 79]. LF ingestion diminishes the systemic inflammation induced by LPS and reduces the associated intestinal damages in different in vivo experimental models [80–83].

Conversely, a recent study has shown that LF deficiency further aggravates LPS-induced inflammation [84]. From a mechanistic point of view, absorbed bLF or bLF peptides can bind to LPS, preventing systemic excessive LPS signaling. Such effect results in the inhibition of the nuclear transcription factor kappa B (NF- $\kappa$ B). Such inhibition then reduces TNF- $\alpha$  secretion by TLR4 (monocytes/macrophages/dendritic cells) and then finally limits the interaction of TNF- $\alpha$  with the TNRF1 (tumor necrosis factor receptor 1) [85]. Such interaction would reduce enterocyte apoptosis and shedding, as well as TJ damage. LF supplementation, when performed before LPS administration, has been recently shown to reduce the circulating concentration of the inflammatory cytokine TNF- $\alpha$ , to prevent the increase intestinal permeability, and to

maintain the morphology of the intestinal mucosa when compared with controls without LF [14]. Moreover, this study reported a good correlation between blood TNF- $\alpha$  level and intestinal permeability, suggesting a direct anti-inflammatory action of LF in the intestine and that the sensitivity of the jejunum and the colon to the beneficial effect of LF against LPS challenge is not similar [14]. LF is also able to inhibit intestinal dysfunction in inflammation by modulating the MAPK pathway, which is essential to maintain intestinal integrity and to reduce production of proinflammatory factors [86]. Another anti-inflammatory effect of LF is related to its ability to reduce production of reactive oxygen species by granulocytes [87]. The ability of LF to protect the intestinal tract against systemic inflammation may involve not only an anti-inflammatory effect but also a direct protective effect on epithelial cells' growth, differentiation, and TJ formation. Figure 1 schematically shows the main beneficial effects of LF.



**Figure 1. Schematic presentation of lactoferrin effects on intestinal inflammation.** LF: lactoferrin; TJ: tight junction; MAPK: mitogen-activated protein kinase; NF-kB: nuclear transcription factor kappa B

# LF is considered an indicator of intestinal mucosa inflammation

Incidentally, but importantly, the amount of LF in feces, which represents neutrophil infiltration in case of intestinal inflammation, is used in addition to calprotectin, as a fecal marker of inflammation [88]. This is because the amounts of LF released by neutrophils have been shown to correlate with the severity of inflammation in the gastrointestinal tract [89]. In that latter case, it is worth to note that LF release from neutrophils is from endogenous but not dietary origin. It is possible that in a healthy situation, a part of LF in the fecal material originates from the diet. In any case, the presence of LF in feces suggests that this protein is not extensively degraded by the intestinal microbiota. Evaluation of the resistance of LF to the different proteases and peptidases equipping the bacteria within the large intestine luminal fluid needs further work.

# **Conclusion and perspectives**

Although there is a set of data indicating a beneficial role of bLF oral ingestion (usually 10 g per kg of diet) for reducing the signs and severity of colitis in several experimental models, we need clinical trials with

volunteers prone to chronic intestinal mucosa inflammation are obviously required to test if such dose may prove to be active for the reduction of the severity of inflammation. Obviously, LF per se cannot substitute for pharmacological treatments in patients with different pathologies resulting in chronic intestinal inflammation, but LF supplementation may prove to be effective in patients as adjunctive therapy in future randomized-controlled clinical trials. More information is needed on the precise dispatching of ingested LF between the intestinal luminal fluid and the bloodstream according to the dose of LF used. Such results are necessary for our understanding of the origin of LF, either from the intestinal content or bloodstream, for the observed effects. Notably, the measurement of the concentrations of LF after dietary supplementation in the small and large intestine, respectively, would be very helpful. Indeed, if, as shown in several experimental studies, LF is partly resistant to protease activities in the gastrointestinal tract, it can be expected that a part of LF will be transferred through the ileocecal junction from the small to the large intestine. Measurement of the resistance of LF to protease and peptidase activities from the intestinal bacteria present in the large intestine luminal fluid also needs to be tested. Randomized controlled clinical studies with increasing doses of oral LF and assay of LF in feces and blood in healthy subjects will be obviously instructive to answer this question. As a significant part of orally ingested LF is degraded within the gastrointestinal tract, new formulations such as encapsulation and nanoparticular carriers that can reduce LF degradation within the gastrointestinal luminal fluid and presumably enhance its absorption may prove to be helpful.

We also need to get more information on the impact of LF in oral supplements and at increasing doses on the microbiota composition and metabolic activity. Indeed, if the available results clearly indicate antimicrobial effects of LF, we do not know the effects of increasing the amounts of oral LF on the fecal microbiota composition. The efficiency of LF for modulating growth and virulence of pathogenic bacteria, fungi, viral, and parasite infections could be usefully tested in animal models.

The encouraging results obtained after oral supplementation with LF in situations of experimental intestinal inflammation should allow utilization of LF in clinical trials with volunteers prone to chronic intestinal inflammation or in remission phase. Progress in the identification of LF-derived peptides and their biochemical characteristics (heat resistance, resistance to bacterial proteases) and physiological characteristics (absorption through the intestinal epithelium, dose-effect curves on pathophysiological parameters) is another important goal. This finding would help development of LF peptides efficient in situations of inflammatory states, whose efficacy will be first tested in experimental studies to provide insights for clinical studies. Moreover, as bioavailability of orally administered LF is low oral formulation resistant to gastric digestion is also needed.

Finally, although LF has been recognized as safe by the American and European authorities, possible side effects of LF supplementation need to be considered and investigated in the different human subpopulations, such as infants and the elderly, to fully evaluate the potential of LF in situations of intestinal inflammation.

Indeed, it is only fair to recognize that most of the knowledge on the effects of LF on the small and large intestine mostly originates from preclinical experimental studies.

#### Abbreviations

bLF: bovine milk-derived lactoferrin
GRAS: generally recognized as safe
IBDs: inflammatory bowel diseases
IL: interleukin
LF: lactoferrin
LPS: lipopolysaccharide
MAPK: mitogen-activated protein kinase

TJ: tight junction TNF-α: tumor necrosis factor alpha

## **Declarations**

#### Author contributions

AB: Conceptualization, Visualization, Writing—original draft, Writing—review & editing.

### **Conflicts of interest**

The author declares that there are no conflicts of interest.

#### Ethical approval

Not applicable.

Consent to participate

Not applicable.

**Consent to publication** 

Not applicable.

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# References

- 1. Kowalczyk P, Kaczyńska K, Kleczkowska P, Bukowska-Ośko I, Kramkowski K, Sulejczak D. The Lactoferrin Phenomenon—A Miracle Molecule. Molecules. 2022;27:2941. [DOI] [PubMed] [PMC]
- Mastromarino P, Capobianco D, Campagna G, Laforgia N, Drimaco P, Dileone A, et al. Correlation between lactoferrin and beneficial microbiota in breast milk and infant's feces. Biometals. 2014;27: 1077–86. [DOI] [PubMed]
- 3. Legrand D, Pierce A, Elass E, Carpentier M, Mariller C, Mazurier J. Lactoferrin Structure and Functions. Adv Exp Med Biol. 2008;606:163–94. [DOI] [PubMed]
- 4. Naot D, Grey A, Reid IR, Cornish J. Lactoferrin—A Novel Bone Growth Factor. Clin Med Res. 2005;3: 93–101. [DOI] [PubMed] [PMC]
- Fischer R, Debbabi H, Blais A, Dubarry M, Rautureau M, Boyaka PN, et al. Uptake of ingested bovine lactoferrin and its accumulation in adult mouse tissues. Int Immunopharmacol. 2007;7:1387–93.
   [DOI] [PubMed]
- 6. Blais A, Lan A, Boluktas A, Grauso-Culetto M, Chaumontet C, Blachier F, et al. Lactoferrin Supplementation during Gestation and Lactation Is Efficient for Boosting Rat Pup Development. Nutrients. 2022;14:2814. [DOI] [PubMed] [PMC]

- Yao X, Bunt C, Cornish J, Quek SY, Wen J. Stability of Bovine Lactoferrin in Luminal Extracts and Mucosal Homogenates from Rat Intestine: A Prelude to Oral Absorption. Chem Biol Drug Des. 2014; 84:676–84. [DOI] [PubMed]
- Dyrda-Terniuk T, Pomastowski P. The Multifaceted Roles of Bovine Lactoferrin: Molecular Structure, Isolation Methods, Analytical Characteristics, and Biological Properties. J Agric Food Chem. 2023;71: 20500–31. [DOI] [PubMed] [PMC]
- Bukowska-Ośko I, Popiel M, Kowalczyk P. The Immunological Role of the Placenta in SARS-CoV-2 Infection—Viral Transmission, Immune Regulation, and Lactoferrin Activity. Int J Mol Sci. 2021;22: 5799. [DOI] [PubMed] [PMC]
- Baker HM, Baker EN. A structural perspective on lactoferrin function. Biochem Cell Biol. 2012;90: 320–8. [DOI] [PubMed]
- Sinopoli A, Isonne C, Santoro MM, Baccolini V. The effects of orally administered lactoferrin in the prevention and management of viral infections: A systematic review. Rev Med Virol. 2022;32:e2261.
   [DOI] [PubMed] [PMC]
- 12. Costagliola G, Nuzzi G, Spada E, Comberiati P, Verduci E, Peroni DG. Nutraceuticals in Viral Infections: An Overview of the Immunomodulating Properties. Nutrients. 2021;13:2410. [DOI] [PubMed] [PMC]
- 13. Valenti P, Antonini G. Lactoferrin: an important host defence against microbial and viral attack. Cell Mol Life Sci. 2005;62:2576–87. [DOI] [PubMed] [PMC]
- Blais A, Takakura N, Grauso M, Puel-Artero C, Blachier F, Lan A. Dietary Bovine Lactoferrin Reduces the Deleterious Effects of Lipopolysaccharide Injection on Mice Intestine. Nutrients. 2024;16:4040.
   [DOI] [PubMed] [PMC]
- 15. EFSA Panel on Dietetic Products, Nutrition and Allergies (NDA). Scientific Opinion on bovine lactoferrin. EFSA J. 2012;10:2701. [DOI]
- 16. Superti F. Lactoferrin from Bovine Milk: A Protective Companion for Life. Nutrients. 2020;12:2562. [DOI] [PubMed] [PMC]
- 17. Tomita M, Wakabayashi H, Shin K, Yamauchi K, Yaeshima T, Iwatsuki K. Twenty-five years of research on bovine lactoferrin applications. Biochimie. 2009;91:52–7. [DOI] [PubMed]
- Wakabayashi H, Yamauchi K, Takase M. Lactoferrin research, technology and applications. Int Dairy J. 2006;16:1241–51. [DOI]
- 19. Carr LE, Virmani MD, Rosa F, Munblit D, Matazel KS, Elolimy AA, et al. Role of Human Milk Bioactives on Infants' Gut and Immune Health. Front Immunol. 2021;12:604080. [DOI] [PubMed] [PMC]
- Blais A, Fan C, Voisin T, Aattouri N, Dubarry M, Blachier F, et al. Effects of lactoferrin on intestinal epithelial cell growth and differentiation: an in vivo and in vitro study. Biometals. 2014;27:857–74. [DOI] [PubMed]
- 21. Liao Y, Jiang R, Lönnerdal B. Biochemical and molecular impacts of lactoferrin on small intestinal growth and development during early life. Biochem Cell Biol. 2012;90:476–84. [DOI] [PubMed]
- Zhang P, Sawicki V, Lewis A, Hanson L, Nuijens JH, Neville MC. Human Lactoferrin in the Milk of Transgenic Mice Increases Intestinal Growth in Ten-Day-Old Suckling Neonates. Adv Exp Med Biol. 2001;501:107–13. [DOI] [PubMed]
- 23. Puddu P, Latorre D, Valenti P, Gessani S. Immunoregulatory role of lactoferrin-lipopolysaccharide interactions. Biometals. 2010;23:387–97. [DOI] [PubMed]
- 24. Cutone A, Ianiro G, Lepanto MS, Rosa L, Valenti P, Bonaccorsi di Patti MC, et al. Lactoferrin in the Prevention and Treatment of Intestinal Inflammatory Pathologies Associated with Colorectal Cancer Development. Cancers (Basel). 2020;12:3806. [DOI] [PubMed] [PMC]
- 25. Demmelmair H, Prell C, Timby N, Lönnerdal B. Benefits of Lactoferrin, Osteopontin and Milk Fat Globule Membranes for Infants. Nutrients. 2017;9:817. [DOI] [PubMed] [PMC]
- 26. Wu J, Zang M, Wang S, Qiao X, Zhao B, Bai J, et al. Lactoferricin, an antimicrobial motif derived from lactoferrin with food preservation potential. Crit Rev Food Sci Nutr. 2024;64:9032–44. [DOI] [PubMed]

- Zarzosa-Moreno D, Avalos-Gómez C, Ramírez-Texcalco LS, Torres-López E, Ramírez-Mondragón R, Hernández-Ramírez JO, et al. Lactoferrin and Its Derived Peptides: An Alternative for Combating Virulence Mechanisms Developed by Pathogens. Molecules. 2020;25:5763. [DOI] [PubMed] [PMC]
- 28. Drago-Serrano ME, Campos-Rodriguez R, Carrero JC, de la Garza M. Lactoferrin and Peptidederivatives: Antimicrobial Agents with Potential Use in Nonspecific Immunity Modulation. Curr Pharm Des. 2018;24:1067–78. [DOI] [PubMed]
- 29. Brines RD, Brock JH. The effect of trypsin and chymotrypsin on the in vitro antimicrobial and ironbinding properties of lactoferrin in human milk and bovine colostrum. Unusual resistance of human apolactoferrin to proteolytic digestion. Biochim Biophys Acta. 1983;759:229–35. [DOI] [PubMed]
- 30. Li W, Liu B, Lin Y, Xue P, Lu Y, Song S, et al. The application of lactoferrin in infant formula: The past, present and future. Crit Rev Food Sci Nutr. 2024;64:5748–67. [DOI] [PubMed]
- 31. Manzoni P. Clinical Benefits of Lactoferrin for Infants and Children. J Pediatr. 2016;173:S43–52. [DOI] [PubMed]
- 32. Teraguchi S, Wakabayashi H, Kuwata H, Yamauchi K, Tamura Y. Protection against infections by oral lactoferrin: Evaluation in animal models. Biometals. 2004;17:231–4. [DOI] [PubMed]
- 33. Suzuki YA, Lopez V, Lönnerdal B. Mammalian lactoferrin receptors: structure and function. Cell Mol Life Sci. 2005;62:2560–75. [DOI] [PubMed] [PMC]
- 34. Takayama Y, Aoki R, Uchida R, Tajima A, Aoki-Yoshida A. Role of CXC chemokine receptor type 4 as a lactoferrin receptor. Biochem Cell Biol. 2017;95:57–63. [DOI] [PubMed]
- 35. Mazurier J, Legrand D, Leveugle B, Rochard E, Montreuil J, Spik G. Study on the Binding of Lactotransferrin (Lactoferrin) to Human PHA-Activated Lymphocytes and Non-Activated Platelets. In: Hutchens TW, Rumball SV, editors. Localisation and description of the receptor-binding site. Boston, MA: Springer US; 1994. pp. 111–9. [DOI] [PubMed]
- 36. Curran CS, Demick KP, Mansfield JM. Lactoferrin activates macrophages via TLR4-dependent and independent signaling pathways. Cell Immunol. 2006;242:23–30. [DOI] [PubMed]
- 37. Blachier F, Kong X. Metabolism of alimentary compounds by the intestinal microbiota and consequences for gut health. J Food Nutr Diet Sci. 2023;1:3–19.
- 38. Burgess SL, Gilchrist CA, Lynn TC, Petri WA Jr. Parasitic Protozoa and Interactions with the Host Intestinal Microbiota. Infect Immun. 2017;85:e00101–17. [DOI] [PubMed] [PMC]
- 39. Actor JK, Hwang SA, Kruzel ML. Lactoferrin as a Natural Immune Modulator. Curr Pharm Des. 2009; 15:1956–73. [DOI] [PubMed] [PMC]
- 40. Kawasaki Y, Sato K, Shinmoto H, Dosako S. Role of Basic Residues of Human Lactoferrin in the Interaction with B Lymphocytes. Biosci Biotechnol Biochem. 2000;64:314–8. [DOI] [PubMed]
- 41. Legrand D, Elass E, Carpentier M, Mazurier J. Lactoferrin: a modulator of immune and inflammatory responses. Cell Mol Life Sci. 2005;62:2549–59. [DOI] [PubMed] [PMC]
- 42. Takakura N, Wakabayashi H, Yamauchi K, Takase M. Influences of orally administered lactoferrin on IFN-γ and IL-10 production by intestinal intraepithelial lymphocytes and mesenteric lymph-node cells. Biochem Cell Biol. 2006;84:363–8. [DOI] [PubMed]
- 43. Sfeir RM, Dubarry M, Boyaka PN, Rautureau M, Tomé D. The Mode of Oral Bovine Lactoferrin Administration Influences Mucosal and Systemic Immune Responses in Mice. J Nutr. 2004;134:403–9.
   [DOI] [PubMed]
- 44. Ostan NKH, Moraes TF, Schryvers AB. Lactoferrin receptors in Gram-negative bacteria: an evolutionary perspective. Biochem Cell Biol. 2021;99:102–8. [DOI] [PubMed]
- 45. Ashraf MF, Zubair D, Bashir MN, Alagawany M, Ahmed S, Shah QA, et al. Nutraceutical and Health-Promoting Potential of Lactoferrin, an Iron-Binding Protein in Human and Animal: Current Knowledge. Biol Trace Elem Res. 2024;202:56–72. [DOI] [PubMed] [PMC]
- 46. Ellison RT 3rd, Giehl TJ, LaForce FM. Damage of the outer membrane of enteric gram-negative bacteria by lactoferrin and transferrin. Infect Immun. 1988;56:2774–81. [DOI] [PubMed] [PMC]

- 47. Crowther GS, Chilton CH, Longshaw C, Todhunter SL, Ewin D, Vernon J, et al. Efficacy of vancomycin extended-dosing regimens for treatment of simulated *Clostridium difficile* infection within an *in vitro* human gut model. J Antimicrob Chemother. 2016;71:986–91. [DOI] [PubMed]
- 48. Florisa R, Recio I, Berkhout B, Visser S. Antibacterial and Antiviral Effects of Milk Proteins and Derivatives Thereof. Curr Pharm Des. 2003;9:1257–75. [DOI] [PubMed]
- 49. van der Strate BW, Beljaars L, Molema G, Harmsen MC, Meijer DK. Antiviral activities of lactoferrin. Antiviral Res. 2001;52:225–39. [DOI] [PubMed]
- 50. Jagirdhar GSK, Pulakurthi YS, Chigurupati HD, Surani S. Gastrointestinal tract and viral pathogens. World J Virol. 2023;12:136–50. [DOI] [PubMed] [PMC]
- 51. Velliyagounder K, Rozario SD, Fine DH. The effects of human lactoferrin in experimentally induced systemic candidiasis. J Med Microbiol. 2019;68:1802–12. [DOI] [PubMed]
- 52. Krupińska AM, Bogucki Z. Lactoferrin as a potential therapeutic for the treatment of Candidaassociated denture stomatitis. J Oral Biosci. 2024;66:308–13. [DOI] [PubMed]
- 53. Ponde NO, Lortal L, Ramage G, Naglik JR, Richardson JP. *Candida albicans* biofilms and polymicrobial interactions. Crit Rev Microbiol. 2021;47:91–111. [DOI] [PubMed] [PMC]
- 54. Jawhara S, Thuru X, Standaert-Vitse A, Jouault T, Mordon S, Sendid B, et al. Colonization of Mice by *Candida albicans* Is Promoted by Chemically Induced Colitis and Augments Inflammatory Responses through Galectin-3. J Infect Dis. 2008;197:972–80. [DOI] [PubMed]
- 55. Fritsch G, Sawatzki G, Treumer J, Jung A, Spira DT. *Plasmodium falciparum*: Inhibition *in vitro* with lactoferrin, desferriferrithiocin, and desferricrocin. Exp Parasitol. 1987;63:1–9. [DOI] [PubMed]
- 56. Sriboonvorakul N, Chotivanich K, Silachamroon U, Phumratanaprapin W, Adams JH, Dondorp AM, et al. Intestinal injury and the gut microbiota in patients with *Plasmodium falciparum* malaria. PLoS Pathog. 2023;19:e1011661. [DOI] [PubMed] [PMC]
- 57. López-Soto F, León-Sicairos N, Nazmi K, Bolscher JG, de la Garza M. Microbicidal effect of the lactoferrin peptides Lactoferricin17–30, Lactoferrampin265–284, and Lactoferrin chimera on the parasite *Entamoeba histolytica*. Biometals. 2010;23:563–8. [DOI] [PubMed]
- 58. Guillén N. Pathogenicity and virulence of *Entamoeba histolytica*, the agent of amoebiasis. Virulence. 2023;14:2158656. [DOI] [PubMed] [PMC]
- 59. Zhang Z, Wang L, Seydel KB, Li E, Ankri S, Mirelman D, et al. *Entamoeba histolytica* cysteine proteinases with interleukin-1 beta converting enzyme (ICE) activity cause intestinal inflammation and tissue damage in amoebiasis. Mol Microbiol. 2000;37:542–8. [DOI] [PubMed]
- 60. Démaris A, Widigson ESK, Ilvemark JFKF, Steenholdt C, Seidelin JB, Huisinga W, et al. Ulcerative Colitis and Acute Severe Ulcerative Colitis Patients Are Overlooked in Infliximab Population Pharmacokinetic Models: Results from a Comprehensive Review. Pharmaceutics. 2022;14:2095. [DOI] [PubMed] [PMC]
- 61. Liu N, Feng G, Zhang X, Hu Q, Sun S, Sun J, et al. The Functional Role of Lactoferrin in Intestine Mucosal Immune System and Inflammatory Bowel Disease. Front Nutr. 2021;8:759507. [DOI] [PubMed] [PMC]
- 62. Nakase H. Optimizing the Use of Current Treatments and Emerging Therapeutic Approaches to Achieve Therapeutic Success in Patients with Inflammatory Bowel Disease. Gut Liver. 2020;14:7–19. [DOI] [PubMed] [PMC]
- Haiwen Z, Rui H, Bingxi Z, Qingfeng G, Jifeng Z, Xuemei W, et al. Oral Administration of Bovine Lactoferrin-Derived Lactoferricin (Lfcin) B Could Attenuate Enterohemorrhagic *Escherichia coli* 0157: H7 Induced Intestinal Disease through Improving Intestinal Barrier Function and Microbiota. J Agric Food Chem. 2019;67:3932–45. [DOI] [PubMed]
- 64. Aly E, López-Nicolás R, Darwish AA, Ros-Berruezo G, Frontela-Saseta C. *In vitro* effectiveness of recombinant human lactoferrin and its hydrolysate in alleviating LPS-induced inflammatory response. Food Res Int. 2019;118:101–7. [DOI] [PubMed]

- 65. Togawa J, Nagase H, Tanaka K, Inamori M, Umezawa T, Nakajima A, et al. Lactoferrin reduces colitis in rats via modulation of the immune system and correction of cytokine imbalance. Am J Physiol Gastrointest Liver Physiol. 2002;283:G187–95. [DOI] [PubMed]
- 66. Togawa J, Nagase H, Tanaka K, Inamori M, Nakajima A, Ueno N, et al. Oral administration of lactoferrin reduces colitis in rats via modulation of the immune system and correction of cytokine imbalance. J Gastroenterol Hepatol. 2002;17:1291–8. [DOI] [PubMed]
- 67. Li L, Ren F, Yun Z, An Y, Wang C, Yan X. Determination of the effects of lactoferrin in a preclinical mouse model of experimental colitis. Mol Med Rep. 2013;8:1125–9. [DOI] [PubMed]
- Sun Y, Liu WZ, Liu T, Feng X, Yang N, Zhou HF. Signaling pathway of MAPK/ERK in cell proliferation, differentiation, migration, senescence and apoptosis. J Recept Signal Transduct Res. 2015;35:600–4.
   [DOI] [PubMed]
- Håversen LA, Baltzer L, Dolphin G, Hanson LA, Mattsby-Baltzer I. Anti-Inflammatory Activities of Human Lactoferrin in Acute Dextran Sulphate-Induced Colitis in Mice. Scand J Immunol. 2003;57: 2–10. [DOI] [PubMed]
- 70. Song L, Xie W, Liu Z, Guo D, Zhao D, Qiao X, et al. Oral delivery of a *Lactococcus lactis* strain secreting bovine lactoferricin-lactoferrampin alleviates the development of acute colitis in mice. Appl Microbiol Biotechnol. 2019;103:6169–86. [DOI] [PubMed]
- 71. Wang S, Zhou J, Xiao D, Shu G, Gu L. Bovine Lactoferrin Protects Dextran Sulfate Sodium Salt Mice Against Inflammation and Impairment of Colonic Epithelial Barrier by Regulating Gut Microbial Structure and Metabolites. Front Nutr. 2021;8:660598. [DOI] [PubMed] [PMC]
- 72. Pool R, Gomez H, Kellum JA. Mechanisms of Organ Dysfunction in Sepsis. Crit Care Clin. 2018;34: 63–80. [DOI] [PubMed] [PMC]
- 73. Lebrun LJ, Pallot G, Nguyen M, Tavernier A, Dusuel A, Pilot T, et al. Increased Weight Gain and Insulin Resistance in HF-Fed PLTP Deficient Mice Is Related to Altered Inflammatory Response and Plasma Transport of Gut-Derived LPS. Int J Mol Sci. 2022;23:13226. [DOI] [PubMed] [PMC]
- 74. Williams JM, Duckworth CA, Watson AJ, Frey MR, Miguel JC, Burkitt MD, et al. A mouse model of pathological small intestinal epithelial cell apoptosis and shedding induced by systemic administration of lipopolysaccharide. Dis Model Mech. 2013;6:1388–99. [DOI] [PubMed] [PMC]
- 75. Stephens M, von der Weid PY. Lipopolysaccharides modulate intestinal epithelial permeability and inflammation in a species-specific manner. Gut Microbes. 2020;11:421–32. [DOI] [PubMed] [PMC]
- 76. Chambon-Savanovitch C, Farges MC, Raul F, Blachier F, Davot P, Cynober L, et al. Can a glutamateenriched diet counteract glutamine depletion in endotoxemic rats? J Nutr Biochem. 1999;10:331–7.
   [DOI] [PubMed]
- Hietbrink F, Besselink MG, Renooij W, de Smet MB, Draisma A, van der Hoeven H, et al. Systemic inflammation increases intestinal permeability during experimental human endotoxemia. Shock. 2009;32:374–8. [DOI] [PubMed]
- King CJ, Tytgat S, Delude RL, Fink MP. Ileal mucosal oxygen consumption is decreased in endotoxemic rats but is restored toward normal by treatment with aminoguanidine. Crit Care Med. 1999;27: 2518–24. [DOI] [PubMed]
- 79. Boutry C, Matsumoto H, Bos C, Moinard C, Cynober L, Yin Y, et al. Decreased glutamate, glutamine and citrulline concentrations in plasma and muscle in endotoxemia cannot be reversed by glutamate or glutamine supplementation: a primary intestinal defect? Amino Acids. 2012;43:1485–98. [DOI] [PubMed]
- 80. Kruzel ML, Harari Y, Chen CY, Castro GA. Lactoferrin Protects Gut Mucosal Integrity During Endotoxemia Induced by Lipopolysaccharide in Mice. Inflammation. 2000;24:33–44. [DOI] [PubMed]
- 81. Kruzel ML, Harari Y, Mailman D, Actor JK, Zimecki M. Differential effects of prophylactic, concurrent and therapeutic lactoferrin treatment on LPS-induced inflammatory responses in mice. Clin Exp Immunol. 2002;130:25–31. [DOI] [PubMed] [PMC]

- Li C, Liu X, Huang Z, Zhai Y, Li H, Wu J. Lactoferrin Alleviates Lipopolysaccharide-Induced Infantile Intestinal Immune Barrier Damage by Regulating an ELAVL1-Related Signaling Pathway. Int J Mol Sci. 2022;23:13719. [DOI] [PubMed] [PMC]
- 83. Doursout MF, Horton H, Hoang L, Liang Y, Hwang SA, Boyd S, et al. Lactoferrin moderates LPS-induced hypotensive response and gut injury in rats. Int Immunopharmacol. 2013;15:227–31. [DOI] [PubMed]
- 84. Liu C, Peng Q, Wei L, Li Z, Zhang X, Wu Y, et al. Deficiency of Lactoferrin aggravates lipopolysaccharide-induced acute inflammation via recruitment macrophage in mice. Biometals. 2023;36:549–62. [DOI] [PubMed] [PMC]
- 85. Håversen L, Ohlsson BG, Hahn-Zoric M, Hanson LA, Mattsby-Baltzer I. Lactoferrin down-regulates the LPS-induced cytokine production in monocytic cells via NF-κB. Cell Immunol. 2002;220:83–95. [DOI] [PubMed]
- 86. Hu P, Zong Q, Zhao Y, Gu H, Liu Y, Gu F, et al. Lactoferrin Attenuates Intestinal Barrier Dysfunction and Inflammation by Modulating the MAPK Pathway and Gut Microbes in Mice. J Nutr. 2022;152:2451–60.
   [DOI] [PubMed]
- 87. Sabra S, Agwa MM. Lactoferrin, a unique molecule with diverse therapeutical and nanotechnological applications. Int J Biol Macromol. 2020;164:1046–60. [DOI] [PubMed] [PMC]
- Zhou XL, Xu W, Tang XX, Luo LS, Tu JF, Zhang CJ, et al. Fecal lactoferrin in discriminating inflammatory bowel disease from irritable bowel syndrome: a diagnostic meta-analysis. BMC Gastroenterol. 2014; 14:121. [DOI] [PubMed] [PMC]
- 89. Abraham BP. Fecal Lactoferrin Testing. Gastroenterol Hepatol (N Y). 2018;14:713–6. [PubMed] [PMC]